

One has to reckon on the luck of finding the desired TS in the allowed region. In contrast, the empirical MINDO-type procedures are universal and well suited for dealing with equilibrium molecular geometries, but their extrapolation into the intermolecular interaction region can result in very serious errors. The need for a reliable semiempirical method to treat chemical reactions is still far from being exhausted.

References and Notes

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- (23) The invariance is slightly disturbed because an empirical parametrization of one-center exchange integrals cannot exactly reproduce the transformational properties of AO's. However, we have agreed to neglect these integrals.

Proton-Electron Hyperfine Coupling Constants of the Chlorophyll *a* Cation Radical by ENDOR Spectroscopy

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Abstract: In this paper we describe the assignment of the major coupling constants in monomer chlorophyll *a* cation free radical by ENDOR spectroscopy. To facilitate chemical manipulation methylpyrochlorophyllide *a* has been used as a stand-in, and a suite of six selectively deuterated derivatives have been subjected to ENDOR investigation. Details of the synthesis of these compounds are described. To study the effect of structural features on the spin distribution in the free radicals, six additional chlorophyll derivatives have been studied. Five coupling constants have been assigned, which account for about 80% of the observed electron spin resonance line width in the chlorophyll *a* monomer cation radical. The spin distribution appears to be highly asymmetric.

ENDOR¹ spectroscopy has provided important support²⁻⁸ for the original ESR interpretation⁹⁻¹¹ that the primary donor of photosynthesis contains two special chlorophylls. Detailed comparisons of in vivo and in vitro ENDOR data have provided an experimental basis for a variety of special pair chlorophyll models^{9,12-24} as the primary donor in photosynthesis. In all previous ENDOR studies an interpretation of the spin density distribution in terms of a specific molecular structure has not been documented. For conclusions based on the in vivo ENDOR to be the most convincing, the ENDOR spectrum should account for the whole of the unpaired spin density rather than reflect only a small fraction of an electron distribution. Incomplete interpretation is a likely possibility since all sites of spin density are not necessarily observable by ENDOR spectroscopy.²⁵ Thus a molecular basis for assignment of the ENDOR spectra of the chlorophyll cations is important.

In this paper we report the detailed analysis of the proton-ENDOR spectrum of the cation radical of chlorophyll *a* (**1**), methylpyrochlorophyllide *a* (**9**), and of a series of related magnesium chlorins. These data provide a clear description of specific assignments of the electron-proton hyperfine coupling constants of in vitro chlorophyll *a* π cation free radical and provide firm support for the view that the ENDOR spectrum of the Chl⁺ free radical accounts essentially for all of the

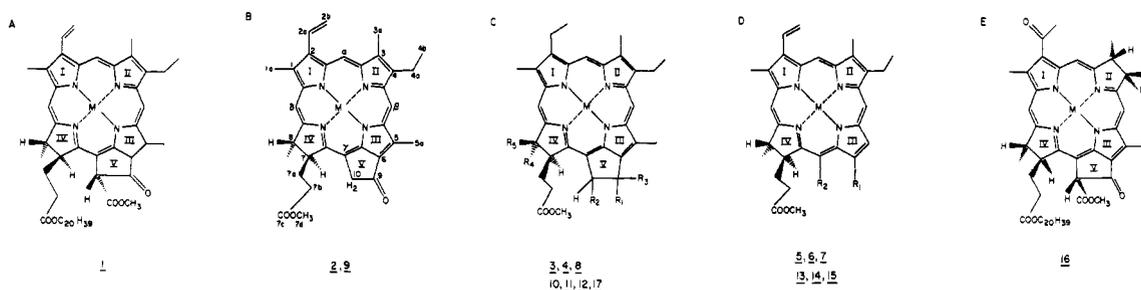
protons on the chlorophyll macrocycle. Comparisons between in vivo and in vitro ENDOR data therefore do in fact appear to be essentially complete.

The model compound (**9**) is essentially identical to chlorophyll *a* insofar as spin distribution in the cation free radicals is concerned, but it offers a much greater variety of possibilities for structural modifications and for the selective introduction of deuterium. The results of the ENDOR spectroscopy on a wide range of selectively deuterated and structurally modified chlorins establish that chlorophyll *a* π cation free radical has low spin density at the four methine bridges and the nitrogen atoms of the pyrrole rings. Although the results are in general agreement with open shell calculations for chlorins, the chlorophyll *a* cation radical shows a strongly asymmetric spin distribution that cannot as yet be completely accounted for by current theory. Finally we show experimentally that this asymmetry is the result of ring V of the chlorophyll macrocycle and is not a result of other various functional side groups.

Experimental Section

Spectra. ENDOR spectra were recorded on a Varian E-700 spectrometer with a Varian large access cavity and at a temperature of 100 K on samples prepared on the vacuum line. Compounds (0.5-1 mg) were first dried by dissolution in benzene, followed by evaporation

Chart I



Compd No. ^a		Modifications					Remarks	Location
Free base ^c (M = H ₂)	Chlorophyllide (M = Mg)	R ₁ , R ₃ ^b	R ₂	R ₄	R ₅			
1, 1a	1, 1a					Chlorophyll <i>a</i>	A	
2, 2a, 2b, 2c	9, 9a, 9b					Basic Chl <i>a</i> derivative	B	
3, 3a	12, 12a, 12b, 12c	=O	H	CH ₃	H		C	
4	10	H ₂	H	CH ₃	H	Racemic	C	
5	14	COOCH ₃	CH ₂ COOCH ₃			No ring V, no R ₃	D	
6	15	H	CH ₂ COOCH ₃			No ring V, no R ₃	D	
7	13	COOCH ₃	H			No ring V, no R ₃	D	
8	11	=O	H	H	CH ₃	Racemic	C	
	16					Bacteriochlorophyll	E	
	17	=O	OH	CH ₃	H	C-10 epimers	C	

^a 1a, 2a, 2b, etc., refer to the corresponding compound with isotopic alterations. ^b Compounds 5, 6, 7, 13, 14, and 15 contain no R₃ group. ^c In the free base Mg has been replaced by H₂.

to dryness at 60 °C in a stream of N₂. Additional drying was accomplished by codistillation with methylene chloride and/or cyclohexane and heating under high vacuum. The sample was then dissolved in 0.75 ml of C²H₂Cl₂ and 0.25 ml of C²H₃O²H, and successive small portions of iodine were added until an optimal ESR signal was obtained. The evacuated sample was then sealed off and stored at 77 K. Chemical oxidation was used to generate chlorin cation free radicals because light cannot be readily introduced into this Varian ENDOR cavity. The free radicals produced by chemical oxidation have been shown to be identical with those produced by photoexcitation.¹⁰

Electron excitation spectra (UV-visible) were recorded on a Cary 14 RI spectrometer. The extinction coefficient of methylpyrochlorophyllide *a* (9) was measured directly, and those of all the other Mg chlorins were determined indirectly by the following method: One milligram of the compound was dissolved in 10 ml of diethyl ether and the spectrum was recorded. The solution was shaken with 5 ml of 2 N HCl and washed with NaHCO₃, the ethereal solution was evaporated to dryness, and the residue was redissolved in 10 ml of a suitable solvent. A spectrum of the thus formed free chlorin was recorded and the extinction coefficient (ϵ) of the original Mg-containing compound was calculated relative to the literature value of ϵ of the free chlorin. With methylpyrochlorophyllide *a* (9) this procedure gave results identical within experimental error with those obtained by conventional procedures.

NMR spectra were recorded on a Varian HR 220 spectrometer operated in the pulse-FT mode. Chemical shifts are given in δ (ppm) relative to TMS or HMS as internal standard. In the exchange experiments, the isotopic compositions were determined by calibration against the signal of the NH proton(s). These are completely exchanged during work-up. **Mass spectra** were obtained on an AEI MS-9 spectrometer. **Mg analysis** was carried out by atomic absorption spectroscopy. Sample solutions were obtained by demetalation of the Mg chlorins dissolved in CH₂Cl₂, and the Mg²⁺ was extracted into water by three extractions with 0.5 N hydrochloric acid.

General Conditions. All solvents were reagent grade. Peroxides were removed by chromatography over basic alumina (Woelm). Magnesium perchlorate (G. F. Smith) was dried for 6 h at 225 °C at $p \leq 10^{-5}$ Torr. "Work-up" of the preparations consisted of thorough washing of the organic phase with water (if acidic, first with NaHCO₃ solution), drying with NaCl, and evaporation under vacuum. The preparative TLC of the free bases were carried out on 20 × 20 cm plates, coated with 0.75 mm Adsorbosil 5 (Applied Science Laboratories), loaded with 10 mg of the crude product. The chlorophyllides were

chromatographed on powdered sugar columns or by HPLC (Waters Associates).

Compounds. See Chart I for structures of numbered compounds. Chlorophyll *a* (1), deuteriochlorophyll *a* ((1)-²H), and methylpyropheophorbide *a* (2) were obtained by standard procedures.^{26,27} Deuterated methylpyropheophorbide *a* ((2)-²H) was obtained from 1-²H by carrying out the transesterification to methylpheophorbide *a* in ²HCl/C²H₃O²H and the subsequent pyrolysis in pyridine containing ²H₂O.

¹H NMR shows 0.4 protons at position 10, 0.14 protons at position 5a, and 0.14 protons in the δ position. *meso*-Methylpyropheophorbide *a* (3) was obtained from 2 by catalytic hydrogenation,²⁸ and *rac*-3 and *rac*-9-deoxy-*meso*-methylpyropheophorbide *a* (4) by reaction of 2 under Wolff-Kishner conditions.²⁹ Compounds with an open isocyclic ring, chlorine *e*₄ trimethyl ester (5), isochlorin *e*₄ dimethyl ester (6), and rhodochlorin dimethyl ester (7) were obtained by suitable modifications of the methods of Fischer.^{30,31} 7,8-*cis-meso*-Methylpyropheophorbide *a* (8) was obtained by Krasnovskii reduction of zinc phyloerythrin.³²

Selective Labeling of Methylpyropheophorbide *a* (2). (a) 10-C²H₂: 20 mg of 2 was refluxed for 48 h under N₂ in 1 ml of pyridine-²H₅/2% ²H₂O. Work-up and TLC on silica 95:5 CCl₄/acetone yielded 18 mg (90%) of the 10-C²H₂ compound. UV-visible and TLC were identical with that of 2; ¹H NMR showed no 10-CH₂ signal (<5%). Under the same conditions, (2)-²H yielded with protic reagents a compound that shows by ¹H NMR 2.2 H at position 10, 0.26 H at position δ , and 0.8 H at position 5a.

(b) 5-C²H₃, 10-C²H₂: according to the procedure of Inhoffen and Mengler³³ 50 mg of 2 was refluxed for 14 days under N₂ in 2 ml of pyridine-²H₅/2% ²H₂O. Work-up and TLC on silica yielded 42 mg (84%) of the 5-C²H₃, 10-C²H₂ compound. UV-visible and TLC were identical with that of 2; ¹H NMR showed no 10-CH₂ (<5%) and 5-CH₃ (<10%). Under the same conditions, (2)-²H yielded with protic reagents a compound which shows by ¹H NMR 2.1 H at position 10, 0.24 H at position δ , and 2.3 H at position 5.

(c) δ -²H, 7''-C²H₃: 20 mg of 2 was refluxed for 1 h with 1 ml of C²H₃O²H/5% ²H₂SO₄. Work-up and TLC yielded 17 mg (85%) of the δ -²H, 7-C²H₃ compound. UV-visible and TLC were identical with that of 2; ¹H NMR showed no δ -H or 7''-CH₃ signal (<5%). Under the same conditions, (2)-²H yielded with protic reagents a compound that shows 3.1 H at position 7'', 1.7 H at position 10, and 0.9 H at the methine position δ by ¹H NMR.

Selective Deuteration of *rac-meso*-Methylpyropheophorbide *a* (3).

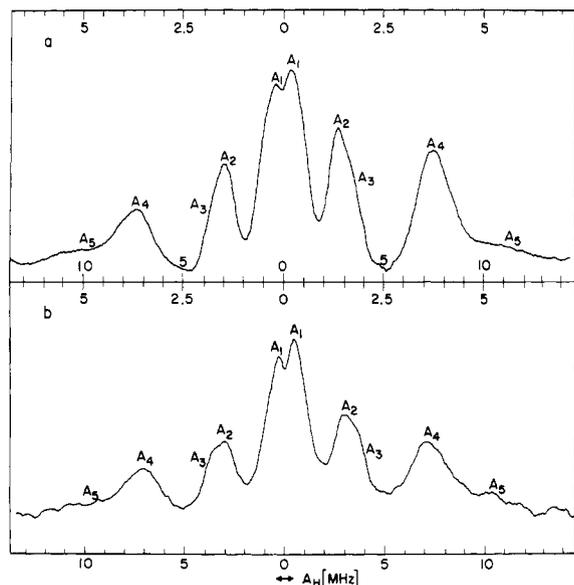


Figure 1. Comparison of (a) ENDOR of chlorophyll *a* (**1**) with (b) ENDOR of methylpyrochlorophyllide *a* (**9**). Lower scale of each spectrum is labeled such that coupling constants of each peak *A* can be read directly in MHz. Upper scale of each spectrum is labeled in true units of MHz frequency shift from the free proton resonance value. In other words one unit of the upper scale corresponds to two units of the lower scale.

Isotopically substituted **3** was prepared by a modification of the method of Fischer and Gibian.²⁹ (**2**)-²H (19 mg) was dissolved in 0.5 ml of pyridine and 2 ml of a freshly prepared saturated solution of sodium methoxide in methanol and 2 ml of 85% hydrazine hydrate was added. The mixture was purged with N₂ and heated in a sealed ampule to 100 °C for 8 h. The crude product was hydrolyzed for 1 h with 25 ml of 25% HCl at 100 °C and the chlorin was then extracted with ether after dilution of the mixture to 2% HCl. Refluxing for 1 h with C²H₃O²H/5% ²H₂SO₄ yielded the methyl esters, which after work-up were separated by TLC on silica (95:5 CCl₄/acetone). Labeled *rac*-*meso*-methylpyropheophorbide *a* (**3**) was obtained in 26% yield, the main by-product being the corresponding 9-desoxo compound (**4**) (18%). UV-visible and TLC of the labeled **3** and of its methylpheophorbide were identical with those of an authentic sample. ¹H NMR: 9.39 (s, β); 9.12 (s, α); 8.38 (s, δ); 5.21, 5.03 (AB pattern, *J* = 18 Hz, 10-CH₂); 4.36 (s, 8-H); 4.18 (s, 7-H); 3.58, 3.53, 3.22 (aromatic CH₃); 2.50, 2.18 (AB, *J* = 16 Hz, 7b-H).

Insertion of Magnesium. A solution of 5 mg of the free base chlorin in 2 ml of pyridine/1% water is heated under N₂. After reflux begins 300 mg of dry magnesium perchlorate is added at once. The refluxing is continued, and the metalation reaction is followed spectrophotometrically to completion (2–10 min). The slurry obtained after cooling was extracted with pyridine and separated by centrifugation. The pyridine extract (1 part) was then partitioned against a mixture of petroleum ether (1 part), water (1 part), and methylene chloride (0.2 parts).

The upper phase was washed four times with water, dried with NaCl, and evaporated. Any residual water and pyridine is removed by co-distillation with benzene at 60 °C. The residue is then dissolved in dry benzene and chromatographed. After the free base chlorin is washed off the column with dry benzene, the Mg complex is eluted with benzene/0.5% 1-propanol, washed with water, and dried. The yield of metalated product is about 80%.

Methylpyrochlorophyllide *a* (9**).** UV-visible (ether): λ_{max} (ε × 10⁻³) = 659 (71.0), 613 (11.2), 428 (104), 407 (63.3), 382 nm (39.8). Mg analysis: found, 4.75%; calcd, 4.13%; ¹H NMR, IR, and TLC identical with an authentic sample.

***rac*-*trans*-9-Deoxy-*meso*-methylpyrochlorophyllide *a* (**10**).** UV-visible (ether): λ_{max} (ε × 10⁻³) = 617 (70.6), 570 (6.1), 406 (354), 386 (100).

***rac*-7,8-*cis*-*meso*-Methylpyrochlorophyllide *a* (**11**).** UV-visible (ether): λ_{max} (ε × 10⁻³) = 651 (92.3), 611 (12.0), 603 (11.6), 572 (6.1), 436 (121), 405 (73.6), 380 nm (41.0). UV-visible (pyridine): λ_{max} (ε × 10⁻³) = 659 (67.0), 631 (12.9), 608 (10.6), 437 (100.5), 415 (58.7), 393 nm (33.1). ¹H NMR: δ [ppm] in acetone ²H₆ = 9.55

(s, β); 9.12 (s, α); 8.29 (s, δ); 5.00, 4.95 (dd, 10-H₂); 4.75 (m, 7-H); 4.62 (m, 8-H); 3.76, 3.73 (q, 2,4-CH₂); 3.53 (s, 7d-CH₃); 3.33, 3.25, 3.17 (s, 1,3,5-CH₃); 2.57 (m, 7a-CH₂); 2.15 (d, 8-CH₃); 1.68 (t, 2a,4a-CH₃); δ (ppm) in pyridine ²H₅ = 9.82 (s, β); 9.28 (s, α); 8.40 (s, δ); 5.22 5.12 (dd, 10-H₂); 4.69 (q, 8-H); 4.42 (m, 7-H); 3.68, 3.65 (q, 2,4-CH₂); 3.70 (7d-CH₃); 3.26, 3.12, 3.05 (1,3,5-CH₃); 2.41 (7a-CH₂); 1.93 (d, 8-CH₃); 1.61, 1.55 (t, 2a,4a-CH₃).

7,8-*trans*-*meso*-Methylpyrochlorophyllide *a* (12**).** UV-visible (ether): λ_{max} (ε × 10⁻³) = 647 (95.1), 607 (12.4), 598 (11.6), 567 (5.9), 423 (136), 403 (78.9), 378 nm (41.9). UV-visible (pyridine): λ_{max} (ε × 10⁻³) = 656 (74.0), 630 (13.2), 607 (11.1), 436 (114), 412 (72.8), 393 nm (43.5). ¹H NMR: δ (ppm) in acetone ²H₆ = 9.51 (s, β); 9.07 (s, α); 8.25 (s, δ); 4.44 (q, 8-H); 4.18 (m, 7-H); 3.69, 3.63 (q, 2,4-CH₂); 3.63 (7d-CH₃); 3.47, 3.11 (s, aromatic CH₃); ~2.3 (m, 7-CH₂CH₂); 1.59 (d, 8-CH₃); 1.51, 1.47 (t, 2,4-CH₃).

Rhodochlorin Dimethyl Ester Magnesium (13**).** UV-visible (ether): λ_{max} (ε × 10⁻³) = 646 (87.0), 598 (11.8), 414 (257), 395 nm (90.4). UV-visible (pyridine): λ_{max} (ε × 10⁻³) = 654 (83.2), 617 (17.3), 430 (198), 409 nm (93.8). Mg analysis: calcd, 4.12%; found, 3.82%. ¹H NMR: 9.69, 9.55, 9.53, 8.99 (s, α, β, γ, δ); 6.54, 5.98 (vinyl H_A, H_B), 4.27, 3.78, 3.55, 3.38, 3.32 (s, aromatic and ester CH₃), 3.60 (4-CH₂), 1.71 (t, 4-CH₃), 1.69 (d, 8-CH₃).

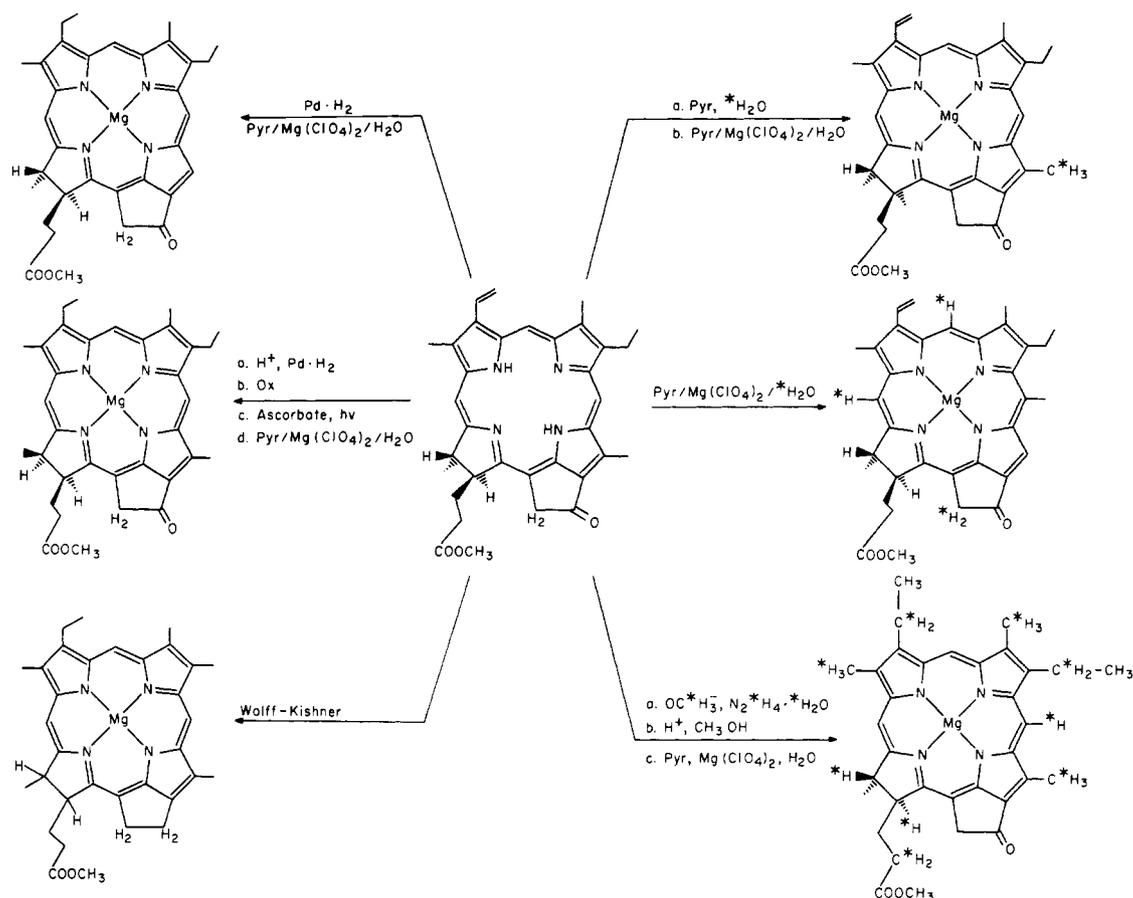
Chlorin e₆ Trimethyl Ester Magnesium (14**).** UV-visible (ether): λ_{max} (ε × 10⁻³) = 642 (76.0), 597 (11.3), 522 (6.8), 414 (203), 395 nm (96.0). UV-visible (pyridine): λ_{max} (ε × 10⁻³) = 649 (66.0), 616 (13.7), 430 (208), 408 nm (86.5). Mg analysis: calcd, 3.68%; found, 3.20%. ¹H NMR: 9.58, 9.55, 8.38 (s, α, β, δ), 5.34 (s, γ-CH₂), 7.88, 6.26, 6.00 (vinyl H_X, H_A, H_B), 3.76, 3.61, 3.45, 3.36, 3.33 (s, aromatic and ester CH₃), 1.67 (t, 4-CH₃), 3.76 (q, 4-CH₂), 1.50 (d, 8-CH₃).

Isochlorin e₄ Dimethyl Ester Magnesium (15**).** UV-visible(ether): λ_{max} (ε × 10⁻³) = 635 (91.6), 591 (11.3), 412 (277), 393 nm (106). UV-visible (pyridine): λ_{max} = 642, 607, 427, 409 nm. Relative intensities = 1:0.18:3.64:1.31. Mg Analysis: calcd, 4.03%; found, 3.97%.

Results and Discussion

The Model Compound. The choice of a model compound for the selective deuteration experiments was dictated by two principal factors: (a) chlorophyll *a* itself (**1**) is unstable toward acids, bases, and heat, and (b) the demetalated chlorophyll *a* (cf. pheophytin *a*), although in principle much easier to modify chemically, cannot as yet be remetalated in reasonable yields. The model compound which overcomes both problems is methylpyrochlorophyllide *a* (**9**). The chemistry of the metal-free methylpyropheophorbide *a* (**2**) has been thoroughly investigated, and, because the enolizable β-ketoester system is lacking, the compound can be smoothly remetalated to the chlorophyllide.^{34,35} In methylpyrochlorophyllide *a* (**9**) the propionic acid phytylester has been transesterified with methanol, and the 10-carbomethoxy group has been removed by pyrolysis. While the propionic ester is several carbon atoms away from the π system and thus is not expected to couple noticeably with the free electron, this is not necessarily true for the changes which result from the removal of the 10-carbomethoxy group and the addition of an additional new β proton (ESR terminology) at C-10. The coupling constant of the 10-protons is very small (see below), however, and thus the substitution of H for COOCH₃ has negligible influence on the ENDOR spectrum. The ENDOR spectra of chlorophyll *a* (**1**) and methylpyrochlorophyllide *a* (**9**) are virtually indistinguishable (Figure 1), and this establishes methylpyrochlorophyllide (**9**) as an appropriate model compound for chlorophyll *a* (**1**).

Insertion of Magnesium. Magnesium, like most divalent metals, is easily incorporated into porphyrins,³⁶ but its insertion into pheophorbides has always been a problem.³⁷ As demonstrated recently, the resistance to Mg insertion is not related to the reduced 7,8 double bond or the isocyclic ring but rather is to be attributed to the enolizable β-ketoester system present in most chlorophylls and their derivatives.³⁴ Thus, the metal is easily incorporated into methylpyropheophorbide *a* (**2**) which lacks the β-ketoester system by the refluxing with anhydrous Mg(ClO₄) (or other Mg salts) in pyridine. The re-

Chart II. Reaction Scheme for Selective Deuteration and Derivative Preparations from Methylpyropheophorbide *a* (2)

action can be followed spectrophotometrically, the best spectral region for this being between 550 and 470 nm, where the two overlapping intermediate intense bands ($\epsilon \sim 12\,000$)³⁸ disappear upon complexation. Magnesium insertion is markedly accelerated by water. Under reasonably available anhydrous conditions (~ 0.1 mol $\text{H}_2\text{O}/\text{l}$), methylpyropheophorbide *a* is metalated to 90% within 1.5 h. Gradual increase of the water concentration to ~ 0.6 M decreases this time to 5–10 min, and the reaction is virtually complete within 15 min.

The Mg complex is partially purified during the work-up procedure, the final separation from by-products and the pheophorbide adduct being achieved by chromatography on powdered sugar. In contrast to the less polar chlorophylls and pheophytins, the 7d methyl esters are considerably more strongly adsorbed. Thus, the columns can be more highly loaded, and a benzene–propanol mixture is suitable for elution.

During the metalation reaction, not only the 10 protons but also the methine protons are exchanged. This observation is paralleled by results of Smith et al.,⁴⁰ who reported the unexpected exchange of the methine hydrogens in porphyrins during metalation with Mg. Both the accelerated Mg insertion and the isotope exchange are possibly due to acid catalysis by water. The δ and 10-protons are easily exchangeable under acidic conditions, but the α - and β -methine protons are observed to exchange usually only at very low pH.^{37,41} There are indications, however, of a considerably increased acidity of water bound to Mg^{2+} in the pyridine/ $\text{Mg}(\text{ClO}_4)_2$ system. At low water concentrations, the water ^1H NMR resonance is shifted to very low field ($\delta > 10$ ppm), and with increasing amounts of water the signal shifts back to its usual value in pyridine at 5 ppm.⁴² Such a low-field shift is characteristic for acidic protons.

Selective Deuterium Exchange (See Chart II) and Additional

Model Compounds. As a general rule, all benzylic protons in pheophorbides are exchangeable under basic conditions. However, the rates of exchange show differences distinct enough to allow selective labeling at least for some of them. See Chart II. In Chl *a* (1) the 10-proton is easily exchanged, being part of the enolizable β -ketoester system.⁴³ In methylpyropheophorbide (2), this exchange occurs in refluxing pyridine with traces of water, via enolization of the 9-CO group.⁴⁴ Prolonged refluxing under these conditions leads to a selective exchange of the 5- CH_3 protons as well, which can be rationalized by a vinologous enolization mechanism.³³ Under the highly basic conditions of the Wolff–Kishner reduction,²⁹ all benzylic positions as well as the enolic 7b-protons are exchanged. As shown by mirror image experiments (see Experimental Section), the C-3 and C-5 methyl protons, the 10-methylene protons, the methine protons, the “chlorin” protons at C-7 and C-8, and the enolic 7b-methylene protons are completely exchanged under these experimental conditions, while the 1- CH_3 as well as the 2- and 4- CH_2 protons are only partially exchanged. This reaction also results in racemization at C-7 and C-8, and in reduction of the 2-vinyl group. Under acidic conditions, the δ -H is substituted⁴¹ and the 7-propionic acid side chain can be transesterified.³⁰ All these exchange experiments have been carried out both by starting from ^1H -methylpyropheophorbide *a* and exchange with deuterium and by starting with the fully deuterated compound and exchanging with protons. The latter approach is of considerable advantage in the assignment of ENDOR lines otherwise hidden.

As mentioned above, the final insertion of magnesium provides a further possibility of exchange (or reexchange) of some of the hydrogen atoms. In this reaction, the 10- CH_2 protons and the α - and δ -methine protons are simultaneously exchanged, which poses limitations on some of the selective ex-

Table I. ENDOR Proton-Electron Coupling Constants (MHz) of Magnesium Chlorins

Compd	A ₁	A ₂	A ₃	A ₄	A ₅	Remarks
(1) Chlorophyll <i>a</i>	0.76	2.83	3.72	7.56	11	
(9) Methylpyrochlorophyllide <i>a</i>						
(9a) Normal isotopic composition	0.81	3.02	3.81	7.22	10.9	
(9b) 5-C ² H ₃ , 10-C ² H ₂	n.v.	2.83	3.50		7-10	Protic solvent
(9c) Only α-H, δ-H, 10-CH ₂	0.5/1.67(sh)					
(12) 7,8- <i>trans</i> - <i>meso</i> -Methylpyrochlorophyllide <i>a</i>						
(12a) Normal isotopic composition	0.65	2.77	3.69	7.25	11.1	2-Vinyl missing
(12b) 3,5-C ² H ₃ , 7,8- ² H	0.62	3.0				1-CH ₃ , 2,4-CH ₂ , part exch.
(12c) 3,5-C ¹ H ₃ , 7,8- ¹ H only	0.26	2.40	2.92	7.06	10	1-CH ₃ , 2,4-CH ₂ , part exch.
(11) 7,8- <i>cis</i> - <i>meso</i> -Methylpyrochlorophyllide <i>a</i>	0.57	2.52	3.35	6.45	8/19.8	
(10) 9-Deoxy- <i>meso</i> - methylpyrochlorophyllide <i>a</i>	1.44	2.91	3.54	8.26	11.5	No 9-CO, 2-vinyl
(17) 10-Hydroxychlorophyll <i>a</i>	n.v.	3.30		7.34	n.v.	Protic solvent
(14) Chlorin e ₆ -TME-Mg	0.36/0.84	3.00		3.57	~7.20	Ring V opened
(13) Rhodochlorin-DME-Mg	0.44(sh), 0.98	2.77	3.54	4.69	8(?)	No γ substituent
(15) Isochlorin e ₄ -DME-Mg	0.44(sh), 0.86	3.16	3.91		7.69	No 6 substituent

change experiments with respect to the final Mg complexes. The isotopic composition of the final products (Table I) has been established from their ¹H NMR spectra, in which all signals in question are well separated and fully assigned.⁴⁵

To determine the influence of the various substituents on the proton-electron coupling constants, several additional structural modifications were made on the model compound **2**. These compounds are tabulated in Chart I. The *meso*-pheophorbide (**3**) was obtained by hydrogenation of the 2-vinyl group,²⁸ the 9-carbonyl group in the 9-deoxopheophorbide (**4**) was removed by Wolff-Kishner reduction,²⁹ and the 7,8-*cis*-pheophorbide (**8**) was obtained by Krasnovskii reduction.³² The influence of the isocyclic ring was studied with compounds in which ring V has been opened, viz., the magnesium complexes of chlorin e₆ trimethyl ester (**14**), isochlorin e₄ dimethyl ester (**15**), and rhodochlorin dimethyl ester (**13**) (Chart I).

The ENDOR Spectrum of Chlorophyll *a* and Methylpyrochlorophyllide *a*. The ENDOR spectrum of the cation radical of Chl *a* (Figure 1a, Table I) shows three well resolved bands (A₁, A₂, A₄), a shoulder (A₃) at the A₂ band, and a broad band (A₅) partially hidden by A₄ and extending as far as 7 MHz. The spectrum is reasonably symmetric with respect to the free proton frequency, indicating no contributions from couplings with nuclei other than protons in the frequency range studied. However, different intensities for the respective high- and low-frequency ENDOR lines are often encountered. These variations in intensity not only arise because of the operation of different relaxation mechanisms but involve also a nonlinear rf sweep, the exact position of the magnetic field *H*₀ with respect to the ESR transition, and perhaps other factors. The ESR spectrum is very similar in dry aggregating and dis-aggregating solvents which indicates a negligible spin delocalization in chlorophyll oligomers formed by keto C=O...Mg interactions.⁹ The spectrum shown in Figure 1 is typical for deuterated or, more precisely, ¹H-free solvents. ¹H-containing solvents give rise to a pronounced matrix peak²⁵ at *A* ~ 0 MHz, which usually obscures the A₁ peaks, and which arises from residual coupling of the free electron with solvent molecules.

The ENDOR spectrum of the cation radical of methylpyrochlorophyllide *a* (**9**) (Figure 1b) is essentially identical with that of Chl *a*⁺. This indicates not only a negligible coupling constant for an unpaired electron with the remote protons of the esterifying alcohol at C-7b, but also suggests a very small coupling constant and low intensity for the C-10 proton(s) coupling constant.

Aromatic Methyl and Methylene Coupling Constants. ¹H/²H exchange of the 5-methyl group (compound **9b**) results in the loss of the major ENDOR line A₄, with only the underlying broad resonance corresponding to the shoulder A₅ in the spectrum of **9** remaining in the spectrum. Obviously, this major ENDOR line in **9** represents coupling only with a single methyl group at C-5. The favorable cross-relaxation time of rotating methyl groups as compared to the more rigid α-protons was first described by Hyde et al.²⁵ For frozen solutions, intense bell-shaped lines were predicted and experimentally verified. The ENDOR spectra of both Chl *a* (**1**) and Bchl *a* (**16**) cation radicals are each dominated by two such lines, and for **16** it can indeed be shown by biosynthetic insertion of deuterium that these peaks arise from the methyl groups in positions 1 and 5.^{46,47} The assignment of the A₄ line to the 5-CH₃ group makes it very probable that the similarly shaped A₂ line likewise originates from interactions involving other methyl groups. In contrast to **16**, **1** has one additional aromatic methyl group at position C-3. This suggests that both the 1- and 3-CH₃ resonances of Chl *a*⁺ are contained in the second (methyl) ENDOR line A₂. This assignment is supported by the relative intensities of A₄ and A₂. In Bchl *a*⁺ both lines represent electron couplings with one methyl group each. The line corresponding to the larger coupling, which we assign by analogy with Chl *a*⁺ to the 5-CH₃ coupling, is about twice as intense as the 1-CH₃ ENDOR line. This is a reflection of the general rule that, given similar relaxation, the intensity of an ENDOR line increases with the magnitude of the coupling constants. In Chl *a*⁺, the ENDOR line, A₂, corresponding to the smaller coupling is increased about twofold for intensity and becomes comparable in size to A₄, thus supporting the assignment of one methyl group (5-CH₃) to A₄ and two methyl groups (1- and 3-CH₃) to A₂.

This assignment is further supported by ²H exchange experiments of **12**. In compound **12b**, the only β type protons that are not completely exchanged are those of the partially exchanged 2- and 4-CH₂ and the 1-CH₃ groups. The ENDOR spectrum of **12b** shows a single line corresponding to A₂ in the ENDOR spectra of compounds **1**, **9**, and **12**. It can be concluded from this experiment that not only the 1- and 3-CH₃ protons but also the 4-CH₂ protons give rise to the A₂ ENDOR line and the A₃ shoulder as well. Assuming comparable spin density distributions in **9** and **12**, this further indicates that the shoulder A₃ arises from coupling with the 3-CH₃ protons. These conclusions are again supported by the corresponding mirror image experiment. Exchange of ²H by ¹H (in compound

(12)-²H) leads to a pronounced diminution of the A₂ resonance in the ENDOR spectrum and the A₃ wing (as compared to the A₄ resonance), corresponding to the only partial exchange of the 1-CH₃, 2-CH₂, and 4-CH₂ deuterons under the exchange conditions.

Methine Protons. Exchange of the δ - and β -methine protons and the 10-CH₂ protons with ²H causes no significant changes in the ENDOR spectrum. Obviously, none of the major lines are related to couplings with these protons. Direct information about coupling with protons in these positions can be obtained, however, from (mirror) isotope inversion studies. Compound **9c**, which contains protons only in the α , δ , and 10 positions (the latter only to the extent of about 50%), gives rise to a single unresolved ENDOR line corresponding to $A \approx 0.5$ MHz with a shoulder at $A = 1.67$ MHz. All three of these sites may contribute to the A₁ line in chlorophyll *a*. During the Mg insertion step all three positions are exchanged simultaneously and without pronounced discrimination, which precludes further discrimination between these resonances from exchange experiments alone. As indicated below, the shoulder on A₁ probably arises from the C-10 protons. The α - and δ -methine interactions are then assigned to the $A = 0.5$ MHz ENDOR line. A small coupling constant involving the third methine proton, i.e., β -H, is indicated by an exchange experiment with ²H-chlorophyll. Occupation of the β , δ , and 10-H positions can be effected by a reversible (Krasnovskii) photo-reduction with H₂S.⁴⁸ The ENDOR spectrum of ²H-chlorophyll in which β , δ , and 10 positions are occupied by ¹H shows only a single center peak corresponding to couplings ≤ 0.5 MHz.

7,8-Protons. An unambiguous assignment of the 7,8-proton couplings is not possible only from selective deuteration experiments. The 7,8-protons can be exchanged under the strongly basic conditions of the Wolff-Kishner reduction, but under these conditions most of the benzylic protons (1,3,5-CH₃, 4- and 10-CH₂) are exchanged as well, thus precluding a sufficiently selective deuteration. In **12b**, the A₄ ENDOR line is missing, and the intensity of A₂ as compared to A₁ is diminished, which as mentioned above aided in the assignment of the latter resonance to interactions with methyl groups. The spectrum of this compound in addition lacks the broad resonance A₅. This resonance is present and unchanged in all other modified chlorophyllides except for one, which indicates that it does not belong to the vinyl, C-10, or methine protons, and thus an assignment to the 7,8-protons suggested itself. This assignment is supported by the spectrum of compound **11** (Figure 2a), which is the only chlorophyllide whose ENDOR spectrum shows a pronounced change in the A₅ signal. Compound **11** is a stereoisomer of the *meso*-chlorophyllide (**12**), in which the 7 and 8 protons are cisoid to each other³² in contradistinction to the transoid configuration present in chlorophylls.^{29,45,49-51} The ¹H NMR spectrum of **11** indicates pronounced conformational changes in ring IV, while the ¹H NMR resonances of the remaining protons in the molecules are less perturbed (cf. ref 32). In the ENDOR spectrum of **11** the most pronounced change is the appearance of a broad resonance with the unusually large coupling constant of $A = 19.9$ MHz. This large coupling leads to a doublet in the ESR spectrum, and indeed the cation radical of **11** is the only one of the compounds investigated with an intact ring V that shows any ESR fine structure at all. The coupling constant obtained from the ESR spectrum ($A \approx 20$ MHz) agrees well with that observed by ENDOR, and the multiplicity of the signal proves interaction with one proton to be responsible for the coupling. As the pronounced configurational and conformational changes of ring IV³² must be responsible for this new coupling, we assign it to one of the "extra hydrogens" at C-7 and C-8. This strongly suggests that the similarly large and broadened A₅ resonance in the 7,8-*trans*-chlorophyllides is due as well

to coupling with these protons.

The assignment of the largest coupling constant in Chl *a*⁺ to the "extra" hydrogens at C-7 and C-8 is paralleled by a similar assignment in the cation radical of bacteriochlorophyll *a* (**16**).⁵² The ESR spectrum of magnesium tetraphenylbacteriochlorin⁺, which was used as a model compound for **16** on the basis of spectroscopic similarities, shows a clearly resolved 7.5 G, (21 MHz) coupling with the eight equivalent "chlorin" protons of the reduced peripheral bonds.

2-Vinyl and 10-Protons. The vinyl group in chlorophylls has been shown to be partially conjugated, but not coplanar with the aromatic macrocycle by ¹H NMR⁴⁵ and UV-visible spectroscopy. The vinyl protons are thus intermediate in coupling between α - and β -protons. In a fully coplanar conformation, the only coupling mechanism would be spin polarization by spin transfer to the vinyl groups. With increasing dihedral angle of the 2-2a bond a hyperconjugative coupling can be expected to occur as well. For intermediate angles both effects are expected to be small, but as the protons in the 1-CH₃ group have a large coupling constant, a substantial coupling with the 2a proton could be possible by both mechanisms. Selective exchange of the vinyl protons was not possible in a straightforward way, but the spin density at the 2-position can be estimated from the meso compound (**12**). In **12**, the 2-vinyl group is converted to a 2-ethyl group by hydrogenation, and consequently two true β -protons are introduced into the molecule. This leads in the ENDOR spectrum to an additional shoulder with a smaller A value than the A₂ resonance, while the remainder of the spectrum is for the most part unchanged. We, therefore, suggest that the shoulder on A₂ arises from interactions with the 2-CH₂ protons in **12**. Although spin redistribution upon hydrogenation of the 2-vinyl group cannot be excluded, the negligible changes in other parts of the spectrum indicate that the new signal does indeed arise from a new group, rather than from change in the coupling constants giving rise to A₂. This assignment is reasonable, as a similar spin density is expected for both C-1 and C-2. In addition, it would suggest a similar coupling constant for the 4-CH₂ protons, which would likewise be concealed in the unresolved A₂ ENDOR peak. The only other significant difference in the ENDOR spectra of **9** and **12** is the coupling constant of the A₁ ENDOR line. This line is a superposition of several unresolved couplings, and A₁ thus represents a mean value only. The pronounced increase (0.38 vs. 0.65 MHz) on conversion of the 2-vinyl group in **1** to a 2-ethyl group in **12** indicates a dominant component with very small coupling in **1**. We take this as indication for a value of $A \leq 0.38$ MHz for coupling by the vinyl protons, although no distinction between the three vinyl protons can be made in this way.

The coupling constant of the 10-proton(s) is probably small, or its relaxation behavior is so unfavorable that the ENDOR signal is too small to be detected. This may be concluded from the similarity of the spectra of methylpyrochlorophyllide *a* (**9**) and of Chl *a* (**1**). In the former compound, C-10 is substituted with two protons, while Chl *a* itself bears only one at this position.

The cation radicals of both compounds have virtually identical ENDOR spectra, which excludes a major contribution of these protons to the ENDOR spectrum. The coupling constant of the 10-protons is more directly accessible from a mirror isotope experiment. In compound **9c**, the α -, β -, and 10-positions are the only ones bearing ¹H, while all other hydrogens in the molecule are ²H. The ENDOR spectrum shows two lines, a weak one with $A = 1.67$ MHz and a stronger one with $A = 0.5$ MHz. As discussed above, exchange occurs simultaneously in all three positions during the metalation procedure, and these three protons cannot be differentiated. An assignment of 10-H to the larger coupling constant with low intensity is probable, however, from the following con-

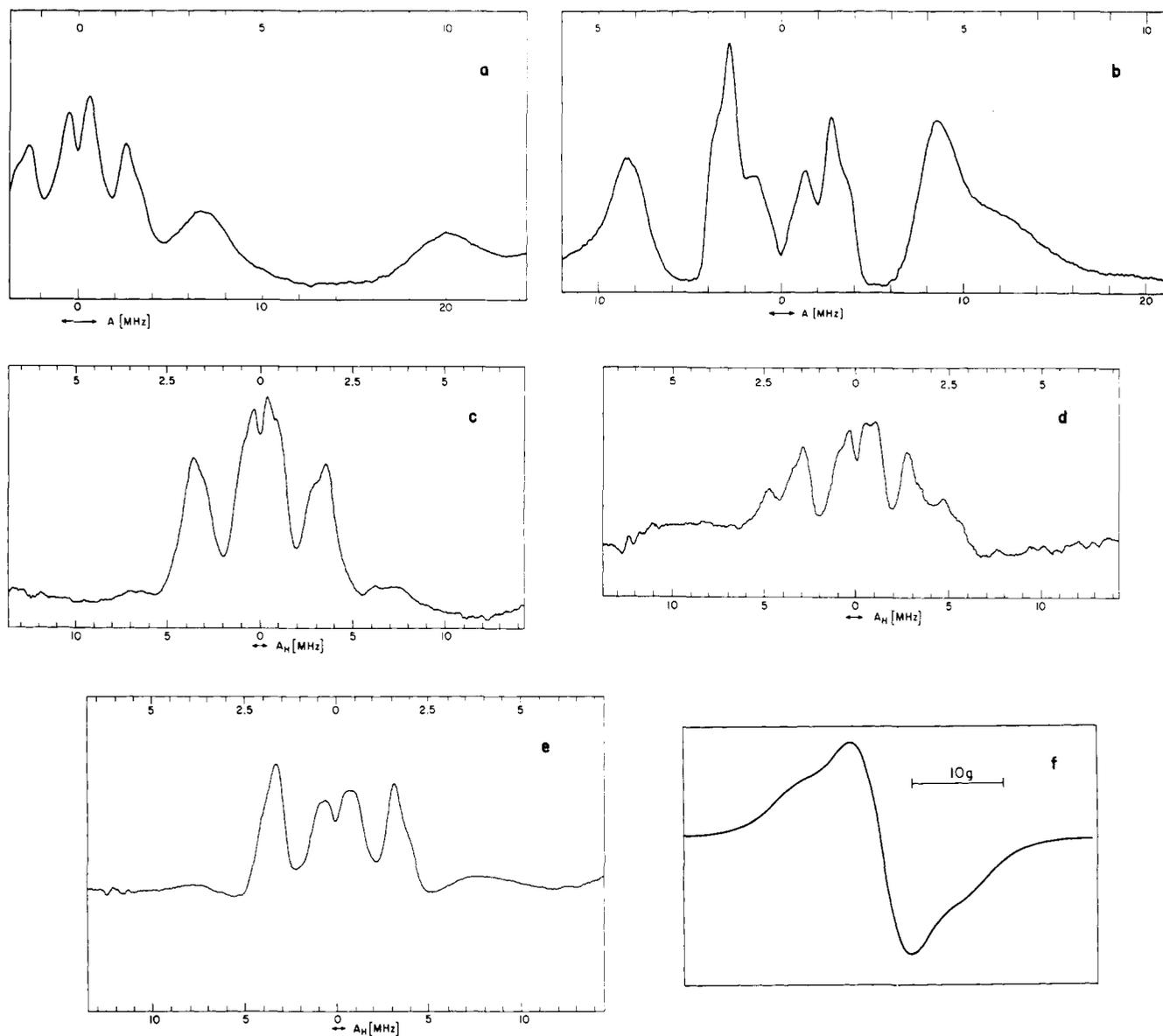


Figure 2. ENDOR spectra of (a) *rac*-7,8-*cis*-*meso*-methylpyrochlorophyllide *a* (**11**), (b) *rac*-9-deoxo-*meso*-methylpyrochlorophyllide *a* (**10**), (c) chlorin e_6 -TME-Mg (**14**), (d) rhodochlorin-DME-Mg (**13**), (e) isochlorin e_4 -DME-Mg (**15**), (f) ESR spectrum of rhodochlorin-DME-Mg (**13**). (See ENDOR spectrum in Figure 2d.) For convenience the lower scale of each ENDOR spectrum is labeled such that coupling constants of each peak A can be read directly in MHz. Upper scale of each spectrum is labeled in true units of MHz frequency shift from the free proton resonance value. In other words one unit of the upper scale corresponds to two units of the lower scale. Although these are typical ENDOR spectra for these compounds and are judged suitable for display purposes, it should be noted that the nonlinear aspects of ENDOR spectroscopy are important when comparing Figure 2 with Table I. Table I represents the results of many ENDOR spectra and is thus more complete.

siderations: The dihedral angle of the 10-H bond with the aromatic carbon is fixed and fairly large ($\sim 60^\circ$); the 10-H is in a β -position not only to the γ -methine C atom but also to C-10, which is conjugated to the aromatic system. The rigidity of the fused rings III and V may be held responsible for the poor relaxation and thus for the low intensity of the ENDOR resonance.

Asymmetry of the ENDOR Spectrum. The very different magnitude of the 5-CH₃ coupling constant and the 1,3-CH₃ coupling constants is one of the principal results of this study. It indicates a pronounced asymmetry of the spin density distribution in the π cation free radical. A series of compounds was investigated to evaluate the origin of this rather unexpected asymmetry.

The cation radical of BChl *a* (**16**) shows a similar difference for the aromatic CH₃ coupling constants. As compound **16** has conjugated carbonyl groups in both the 2- and 6-positions, the asymmetry in coupling constants does not arise from the 9-CO

group in the chlorophylls. Furthermore, it does not arise from the presence of a 2-vinyl group, as compound **12** that lacks this substituent shows the same basic ENDOR features.

These conclusions are supported by the ENDOR spectrum of the 9-deoxochlorophyllide (**10**) (Figure 2b), which lacks the 9-CO as well as the 2-vinyl group. The influence of the latter was shown to be small, thus any spectral changes must be due mostly to the 9-CO group. The ENDOR spectrum shows still the same general features as that of Chl a^+ , but two changes are noticeable: Both the A_2 and A_4 resonances are strongly increased in intensity. Although a detailed interpretation of these changes is not possible without examination of compounds with additional structural modifications, we can make two deductions. First, there is some redistribution in spin density, and second, the asymmetry of the spin distribution persists or is even rather enhanced. Thus, the coupling constant ratio of the two methyl resonances serving as labels for the asymmetry is increased from 2.5 to 2.85. It should be men-

tioned that reduction of the 9-CO group leads to a higher overall π -electron density in the macrocycle, as judged by the ^1H NMR ring current induced shifts. This increase seems to be true for the highest occupied MO likewise, as most discernible coupling constants in **10** are increased as compared to **1**, **9**, and **12**.⁵³

In the compounds without the isocyclic five-membered ring, the two methyl peaks in the ENDOR spectra gradually move together, as the steric hindrance between the substituents at C-6, C- γ , and C-7 is relieved. Chlorin e_6 (**14**) (ENDOR spectrum in Figure 2c) has an A_4/A_2 ratio of 1.2. The ratio is 1.3 in the rhodochlorin (**13**) (ENDOR spectrum in Figure 2d), and in the isochlorin e_4 (**15**) (ENDOR spectrum in Figure 2e) both methyl group signals have merged. As the coupling constants are in first order related by the same Q value (see below) to the spin densities at the neighboring C atom, these results clearly indicate that the strongest asymmetric perturbation of the spin density in the chlorophylls is due to the presence of the isocyclic ring. In addition to the effects on the methyl resonances, pronounced changes in other signals in the three ring V opened compounds can be observed. The A_5 resonance assigned to the 7,8-proton couplings is decreased,⁵⁴ and the A_1 ENDOR line shows resolved fine structure. All of our results confirm the major influence of ring V on the spin distribution. It is known that the presence of ring V gives rise to major structural distortions in the vicinity, especially in that of ring III.⁵⁵ While UV-visible and ^1H NMR are comparatively unaffected by this steric effect, the configuration of the highest occupied MO is obviously strongly affected by it. It has been argued that there are two such orbitals narrowly spaced in the porphyrin cations⁵⁶ (see below) and the observed changes are possibly due to a different degree of mixing between the two orbitals.

The ESR Line Width. The peak-to-peak ESR line width ΔH of an inhomogeneously broadened Gaussian line is related to the second moment $\langle\omega^2\rangle$ by $\Delta H^2 = 4\langle\omega^2\rangle$. In the case of chlorophyll, coupling with more than one kind of nucleus can occur, and the second moment can be split up into several components

$$\langle\omega^2\rangle = \langle\omega^2\rangle_{^1\text{H}} + \langle\omega^2\rangle_{^{13}\text{C}} + \langle\omega^2\rangle_{^{14}\text{N}} + \langle\omega^2\rangle_{^{25}\text{Mg}} + \langle\omega^2\rangle_{g\text{-anis}}$$

corresponding to the contributions of the nuclei present and a possible small contribution arising from g -factor anisotropy $\langle\omega^2\rangle_{g\text{-anis}}$. The magnitude of particular contributions can be determined by isotope substitution. In the case of hydrogen, the total line width of ^1H -chlorophyll a ($\Delta H_{^1\text{H-Chl}} = 9.3\text{ G}$)⁹ and ^2H -chlorophyll a ($\Delta H_{^2\text{H-Chl}} = 3.8\text{ G}$)⁹ is given by

$$\frac{1}{4}\Delta H_{^1\text{H-Chl}}^2 = \langle\omega^2\rangle_{^1\text{H-Chl}} = \langle\omega^2\rangle_{^1\text{H}} + \langle\omega^2\rangle_{\text{rest}}$$

$$\frac{1}{4}\Delta H_{^2\text{H-Chl}}^2 = \langle\omega^2\rangle_{^2\text{H-Chl}} = \langle\omega^2\rangle_{^2\text{H}} + \langle\omega^2\rangle_{\text{rest}}$$

With the relation

$$\langle\omega^2\rangle_{^1\text{H}} = 15.9\langle\omega^2\rangle_{^2\text{H}}$$

a ^1H contribution of 19.2 G^2 to the total second moment of 21.6 G^2 has been determined from fully ($\geq 99\%$) ^1H - or ^2H -chlorophylls.⁵⁷

In the case of protons, the second moment is related to the individual coupling constants A_i by

$$\langle\omega^2\rangle = \frac{1}{4} \sum_i I_i A_i^2$$

where I_i is the number of equivalent nuclei. With the coupling constants derived from the ENDOR spectra (Table II) a calculated second moment of $\langle\omega^2\rangle = 15.6\text{ G}^2$ is obtained. This value is in good agreement with experiment, especially when 80% of the ^1H second moment is due to coupling with just five

Table II. Comparison of Experimental Coupling Constants A with Predicted Spin Density Values ρ^e

	A, G	NA^2, G^2 ^b	$ \rho ^c$	ρ^d	ρ^e
1-CH ₃	1.0	3.0	0.030	0.046	0.040
2a-H	0.14	0.02	0.005		0.014
2b-H _{A,B}	0.14	0.04	0.005		-0.023
3-CH ₃	1.31	5.18	0.039	0.032	0.004
4-CH ₂	1.0	2.0	0.030	0.032	0.008
5-CH ₃	2.67	21.4	0.080	0.020	0.070
10-H	0.60	0.36	0.018		
7-H	3.89	15.11	0.116	0.114	0.035
8-H	3.89	15.11	0.116	0.114	0.037
α -H	0.18	0.03	0.007	-0.043	-0.067
β -H	0.18	0.03	0.007	-0.043	-0.073
δ -H	0.18	0.03	0.007	+0.017	-0.005

^a Experimental values from Table I using A (in G) = A (in MHz)/2.83. ^b $NA^2/4$ is the contribution of a particular group of protons to the total second moment where N is the degeneracy of the coupling constant A . This column sums to the value of 62.31 G^2 and thus accounts for 15.6 G^2 of the second moment. ^c Spin density ρ is obtained from the respective coupling constants by using the following Q values: $Q = 27\text{ G/spin}$ for α -protons, $Q = 33.5\text{ G}$ for β -protons in agreement with the interpolated value in text. In the case of fixed β -protons this corresponds to an approximate dihedral angle of 45° . ^d Private communication from R. Felton. ^e Private communication from P. S. Song. For ease of comparison Song's ρ values have been "normalized" by division by 5 before listing in Table II.

protons, i.e., the three 5-methyl protons and two at C-7 and C-8. The coupling with the 5-CH₃ gives rise to a well-defined ENDOR line, but the couplings with 7-H and 8-H give rise to broad and partially hidden lines. These two couplings contribute 44% of the line width,⁵⁹ and thus small errors in the magnitude of these coupling constants give rise to a considerable error in the second moment. Thus, an increase in each of the 7,8 coupling constants by 0.5 MHz (which may well be a possibility because of the weak, broad ENDOR lines from these protons) would be sufficient to fit the data completely. Although the assignment of the major coupling constants has been done in a straightforward way and is well-supported, the reader is cautioned that resonances not directly accessible may require additional data before the assignments can be considered conclusive.⁶⁰

Spin Density. Proton-electron coupling constants A_i are related to the spin density ρ at the carbon atom bearing the proton (α -type coupling) or on the neighboring carbon atom (β -type coupling) by a McConnell⁶¹ relation:

$$A_i = Q\rho_i$$

The spin densities in Table II were obtained from the respective coupling constants by using the following Q values: $Q = 27\text{ G/spin}$ for α -protons, $Q = 38\text{ G/spin}$ for rotating β -protons, and $Q = -9.2\text{ G} + 96.7\text{ G} \cos^2 \theta$ for fixed β -protons. The latter formula is derived from ESR measurements on alkylbenzene cation radicals,⁶² where θ is the dihedral angle between the π orbital at C_α and the C_β -H $_\beta$ bond. By averaging this relation over θ , the Q value of 38.6 G/spin for freely rotating β -protons was obtained. This value obviously represents an upper limit. Q shows a pronounced charge dependence⁶³ and decreases with increasing negative charge. The lower limit would then be defined by $Q = 25\text{ G/spin}$ found for anion radicals of alkylbenzenes.⁶⁴ In the alkylbenzenes, the charge is distributed over a much smaller number of π centers than in the porphyrins. Assuming an equal charge distribution, the average charge density in the chlorophyll macrocycle is about $1/4$ of that in benzene radical ions. From a linear interpolation between the two extremes, a Q value of about 33.5 G/spin can then be estimated for the Chl a^+ cation radical.

The results indicate a very low spin density at the methine bridge and intermediate spin density at the β -pyrrole carbon atoms of rings I, II, and III and the α -pyrrole carbons of ring IV. Systematic studies of porphyrin π -cation radicals have led to the characterization of two types of radicals which show pronounced differences both in their uv-visible and ESR spectra.⁵⁶ The first group of which ZnTPP^{•+} is a typical example has a well-resolved ESR spectrum and a series of overlapping visible absorption bands in the region of 500–700 nm. The second group of which MgOEP^{•+} is representative has an unresolved, inhomogeneously broadened ESR spectrum at room temperature⁵⁵ and shows a strong absorption band at about 700 nm. Both classes of compounds have been investigated in detail by Fajer et al.,⁶⁵ and the experimental findings have been interpreted in terms of two closely spaced highest occupied molecular orbitals, which have different relative energies in the two classes of compounds. In ZnTPP^{•+} the free electron occupies the a_{2u} orbital, which has high density at the nitrogen and meso positions. In ZnOEP^{•+} the a_{1u} orbital becomes the highest occupied molecular orbital, which is rotated by $\pi/4$ with respect to the a_{2u} orbital and has high density at the π carbons. Although there is probably some mixing of the two states, the model is a good guide in interpreting the spectral properties of π -cation radicals of porphyrins.

To first order, the results obtained for porphyrin cations can be applied to chlorin cation radicals as well. Both the observed low-spin density at the methine positions of Chl $a^{\cdot+}$ and the earlier demonstrated low-spin density at the inner nitrogens are against the free electron residing in an a_{2u} type orbital. This is also indicated from the UV-visible spectrum of Chl $a^{\cdot+}$, which shows a strong absorption at 662 nm. However, it is important to note that Chl $a^{\cdot+}$ can only be crudely approximated by the symmetric MgOEP^{•+}.

Two open shell calculations for the chlorophyll a cation radical are compared in Table II. In the first study (Table II) the Chl $a^{\cdot+}$ is approximated by a Mg chlorin.⁶⁶ The C_2 symmetry of the model is reflected in the equal spin density at symmetric positions. The orbital occupied by the free electron is basically of the a_{2u} type, but there are pronounced differences in the spin density distribution between rings I and III and ring II, respectively. The net spin density of the rings increases monotonically with increasing distance from the reduced peripheral double bond (IV < I, III < II) and four of the β -pyrrole carbons (1, 6, 3, 4) gain considerable spin density from the neighboring α -pyrrole carbons. A (small) negative spin density is only obtained at the α - and β -methine positions. The more detailed SCF-CI calculation of Song⁶⁷ includes both the 2-vinyl and the 6-carbonyl group (Table II). Two pronounced differences in this model are immediately obvious. It yields a much less symmetric spin density distribution, and it gives rise to both large positive and negative spin densities. The total spin in this model is represented by five rather than one spin, which renders an assessment of the spin density from proton ENDOR data alone difficult. The model correctly predicts a much larger (negative) spin density on C-5 than on C-1, but the small spin density on C-3 and the predicted large spin density at the methine positions seems to be incompatible with the ENDOR results. Moreover, the ENDOR results do indicate the asymmetric spin density is due neither to the 2-vinyl nor to the 6-CO substituent (the 9-carbonyl group), but rather to the presence of the isocyclic ring. It is the influence of this ring, however, which is most difficult to take into account in calculations of the spin density in the π cation radical. If the sign for the spin densities predicted by the latter calculation (Song) (Table II) is used the total spin density in the β -pyrrole, the methine, and the vinyl C atoms is about 31% of one electron. As the four inner nitrogen atoms, as well as the central magnesium atom, are essentially spin free, we are forced to conclude that the remaining spin does reside in the α -pyrrole carbons as well as

in the 6-carbonyl group.

Conclusions

Our ENDOR results show protons 7-H and 8-H to have the largest proton–electron coupling constants. As these are fixed β -protons, small changes in the dihedral angle between them can make a significant change in their coupling constants, and thus in both the ENDOR and ESR line widths. The next largest coupling constant arises from the 5-CH₃ group, followed by the 1-CH₃, 3-CH₃, and 4-CH₂ groups. We have tentatively assigned small coupling constants to all macrocycle α -protons, i.e., α , β , δ , the 2-vinyl protons, and the β -10-H. In the ENDOR experiments described here, α -protons are expected to give rise to broad and very weak ENDOR signals, and thus can easily go undetected. Consequently, it is possible that some of these α -protons may have a somewhat larger coupling constant than we have indicated. We again caution the reader that we still do not account for a small amount ($\sim 3 G^2$) in the second moment of the proton–electron hyperfine interactions. As previously mentioned, this missing portion of the second moment may be attributed to inaccuracies in the measurement of the coupling constants of the 7- and 8- (fixed) β -protons. These have wide ENDOR resonances that make accurate determination of coupling constants difficult. However, we cannot rule out at this time the possibility that some of the missing second moment arises from unobserved α -protons that have higher spin density than we have tentatively assigned.

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Advantages of ^{15}N NMR in Studying Delocalization and Evaluating the Energy Barriers of C-N Rotation Process in Amides, Thioamides, and Related Compounds

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Abstract: Linear correlations are shown to exist between the activation energy E_a of the C-N rotation process and $\delta^{15}\text{N}$ in *N,N*-dimethylamino derivatives. The relations are applied to the prediction of E_a values related to high or low energy barriers, where the direct NMR methods of measurement are not applicable. The validity of the correlations is discussed on the basis of semiempirical calculations of the nitrogen chemical shifts.

The variation of ^{14}N chemical shifts in amides and thioamides was qualitatively explained by Hampson and Mathias¹ in terms of the delocalization of the nitrogen lone pair. Siddall et al.,² however, failed to observe the predicted correlation between ^{14}N downfield shifts and the increase of barrier heights to rotation in *N*-alkyl substituted amides and explained their results by the prominent role of steric effects. We wish to present here ^{15}N data which provide a good means of evaluating C-N bond rotation energy barriers in cases where

conventional dynamic NMR methods (line shape analysis or Forsen-Hoffmann double resonance studies) are not applicable. Our results also indicate a striking difference in the behavior of ^{13}C and ^{15}N chemical shifts in a $>\text{N}=\text{C}<$ fragment, insofar as electronic delocalization is concerned.

Table I collects $\delta^{15}\text{N}$ and $\delta^{13}\text{C}$ values for the $>\text{N}=\text{C}<$ fragment of selected compounds for which good activation energy values are available. Amides, thioamides, and related compounds were considered and good correlations between E_a