Molecular Mechanisms of Membrane Traffic

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CONTENTS

CONTENTS
Preface
ENDOPLASMIC RETICULUM, STRUCTURE, FUNCTION AND TRANSPORT TO GOLGI COMPLEX
Cell-Free Analysis of ER to Golgi Apparatus Vesicular Traffic1-15 D J Morré and D M Morré
Identification of a Novel Post-ER, Pre-Golgi Compartment Where Unassembled Monomers of Oligomeric Proteins Accumulate17-34 T C Hobman, I Woodward, M Komuro and M G Farquhar
G Protein Regulation of Vesicular Transport Through the Exocytic Pathway
Fusion Rapidly Follows Vesicle Transport to the Target Membrane in Protein Transport Through the Golgi Apparatus <i>In Vitro</i> . A Reevaluation of Transport Kinetics Based on the Finding That the Glycosylation Used to Mark Transport, and Not Transport Itself is Rate Limiting in the Assay
Immunocytochemical Analysis of the Transfer of Vesicular Stomatitis Virus G Glycoprotein from the Intermediate Compartment to the Golgi Complex
Synthesis of Glycosyl-Phosphatidylinositol Anchor is Initiated in the Endoplasmic Reticulum
Protein Trafficking Along the Exocytotic Pathway57-58 W Hong
A Short N-Terminal Sequence is Responsible for the Retention of Invariant Chain (Ii) in the Endoplasmic Reticulum59-60 M-P Schutze, M R Jackson and P A Peterson
Isolation and Characterization of the Principal ATPase of Transitional Elements of the Endoplasmic Reticulum of Rat Liver61-62 L Zhang and D J Morré
A GTP Hydrolase Activity Purified from Transitional Endoplasmic Reticulum of Rat Liver Binds Retinol
A 38 kDa Protein Resident to Cis Golgi Apparatus Cisternae of Rat Liver is Recognized by an Antibody Directed Against α Subunits of Trimeric G-Proteins

Acyl Transfer Reactions Associated with Cis Golgi Apparatus67-68 J B Lawrence, T W Keenan and D J Morré
Hepatic Bile Formation: Intracellular Trafficking of Bile Salts69-70 J M Crawford, S Barnes, R C Stearns, D L Hastings, D C J Strahs and J J Godleski
Wheat Storage Proteins as a Model System to Study the Mechanism of Protein Sorting Within the Endoplasmic Reticulum71-72 G Galili and Y Altschuler
IMPORT INTO MITOCHONDRIA, PEROXISOMES, CHLOROPLASTS AND VACUOLES
Determinants of Outer Membrane Protein Sorting and Topology in Mitochondria73-79 G C Shore, H M McBride, D G Millar and J -M Li
Molecular Chaperones HSP70 and HSP60 in Protein Folding and Membrane Translocation81-90 J Martin and F -U Hartl
Cytosolic Reactions in Mitochondrial Protein Import91-94 D M Cyr and M G Douglas
Human Bifunctional Enzyme and Its Import Into Peroxisomes95-98 G L Chen and M C McGuinness
Transport of Microinjected Proteins into the Peroxisomes of Mammalian Cells
Isolation and Characterization of a Functionally Active Protein Translocation Apparatus from Chloroplast Envelopes101-104 K Waegemann and J Soll
Protein Insertion into the Outer Mitochondrial Membrane $\dots 105-106$ D Millar and GC Shore
The Membrane-Bound 95 kDa Subunit of the Yeast Vacuolar Proton-Pumping ATPase is Required for Enzyme Assembly and Activity107-108 M F Manolson, D Proteau, R A Preston, M E Colosimo, B T Roberts, M A Hoyt and E W Jones
The Traffic of Molecules Across the Tonoplast of Plant Cells109-110 B P Marin
Fusicoccin Binding and Internalization by Soybean Protoplasts111-112 M A Villanueva, R. Stout and L R Griffing
TRAFFIC WITHIN AND FROM THE GOLGI COMPLEX
TGN38 and Small GTP Binding Proteins are Part of a Macromolecular113-116 Complex in the Trans-Golgi Network K E Howell

β-Cop, a Coat Protein of Nonclathrin-Coated Vesicles of The Golgi Complex, is involved in Transport of Vesicular Stomatitis VirusGlycoprotein117-126
R Duden, B Storrie, R Pepperkok, J Scheel, B Joggerst-Thomalla, A Sawyer, H Horstmann, G Griffiths and T E Kreis
Defining the Retention Signal in a Model Golgi Membrane Protein127-133 C E Machamer, M G Grim, A Esquela, K Ryan and A M Swift
Organization of the Glycoprotein and Polysaccharide Synthetic Pathways in the Plant Golgi Apparatus135-144 L A Staehelin
CLIP-170, a Cytoplasmic Linker Protein Mediating Interaction of Endosomes with Microtubules
Maturation of Secretory Granules
Fatty Acylation in Membrane Trafficking and During Mitosis163-166 D I Mundy
Rates of Synthesis and Selective Loss Into the Bile of Four Rat Liver Proteins and the Polymeric IgA Receptor
Effect of Brefeldin A Treatment on the Resident Golgi Protein, MG160.171-172 P A Johnston and N K Gonatas
Inhibition of Cholera Toxin by Brefeldin A
Selective Association with Golgi of ADP-Ribosylation Factors, 20-kDa Guanine Nucleotide-Binding Protein Activators of Cholera Toxin
Intracellular Transport and Post-Translational Modifications of a Secretory Heat Shock Protein of Saccharomyces Cerevisiae177-178 M Simonen, M Wikström, B Walse and M Makarow
Transport to the Cell Surface of <i>Chlamydomonas</i> : Mastigonemes as a Marker for the Flagellar Membrane
LIPID TRAFFICKING PROTEINS OF THE PLASMA MEMBRANE AND THEIR INTERACTION
Lipid Transport From the Golgi to the Plasma Membrane of Epithelial Cells

Lipid Traffic to the Plasma Membrane of Leek Cells. Sorting Based on Fatty Acyl Chain Length
Hexadecylphosphocholine as a Useful Tool for Investigating Phosphatidylcholine Biosynthesis and Sorting197-198 C C Geilen, Th Wieder and W Reutter
Biosynthesis of the Scrapie Prion Protein in Scrapie-Infected Cells
Protein Phosphorylation Regulates the Cellular Trafficking and Processing of the Alzheimer Beta/A4 Amyloid Precursor Protein
Targeting of the Polymeric Immunoglobulin Receptor in Transfected PC12Cells203-204 F Bonzelius, G A Herman, M H Cardone, K E Mostov and R B Kelly
THE ENDOCYTIC PATHWAY AND SIGNALS
Signals for Receptor-Mediated Endocytosis
Regulation of Early Endosome Fusion <i>In Vitro</i>
Adaptins and Their Role in Clathrin-Mediated Vesicle Sorting229-236 M S Robinson, C L Ball and M N J Seaman
Endosomes and Cell Signalling237-244 B I Posner and J J M Bergeron
The Tubular Early Endosome
Direct Comparision of the Endocytic Routes of Fluorescently-Labelled Lipids and Transferrin
Linkage of Plasma Membrane Proteins with the Membrane Skeleton: Insights Into Functions in Polarized Epithelial Cells273-283 W J Nelson
Cell Specificity and Developmental Variation in the Targeting Pathways of FRT Cells285-288 C Zurzolo

Phosphoglucomutase is a Cytoplasmic Glycoprotein Implicated in the Regulated Secretory Pathway
Protein Targeting and the Control of Cl ⁻ Secretion in Colonic EpithelialCells293-296 A P Morris, S A Cunningham, D J Benos and R A Frizzell
Apical Membrane Protein Sorting is Affected by Brefeldin A297-299 C B Brewer and M G Roth
Structural Analysis of the Clathrin Triskelion301-305 I S Näthke and F M Brodsky
Polarity in Neurons and Epithelial Cells: Distribution of Endogenous and Exogenous Ion Pumps and Transporters
Biochemical Properties and Expression of Tau Proteins in the Endocytic Compartment During Liver Regeneration
Identification of Hepatic Endocytic Proteins Potentially Involved in Membrane Traffic
Studies on Sequence Requirements for Basolateral Targeting of the Polymeric Immunoglobulin Receptor in MDCK Cells313-314 B Aroeti and K Mostov
Endocytosis by <i>Trypanosoma Brucei</i> : Proteases and Their Possible Role in the Degradation of Anti-VSG Antibodies315-316 J D Lonsdale-Eccles, D J Grab, D C W Russo and P Webster
Receptor Mediated Endocytosis of EGF in a Cell Free System317-318 T E Redelmeier, C J Lamaze and S L Schmid
Transferrin on the Basolateral Surface Regulates Apical ⁵⁹ Fe Uptake and Transport Across Intestinal Epithelial Cells319-320 X Alvarez-Hernandez and J Glass
Intracellular Trafficking of Salmonella typhimurium Within HeLa EpithelialCells
Functional Analysis of Dynamin, a GTPase Meditating Early Endocytosis
Identification of a Regulatory Domain in Dynamin
Potassium Depletion Stimulates Clathrin-Independent Endocytosis in Rat Foetal Fibroblasts

Clathrin Interaction with NADH Dehydrogenases of Rat Liver Plasma Membrane329-330 P Navas, J M Villalba, J C Rodríguez-Aguilera, A Canalejo and M I Burón
Putative Adaptor Proteins of Clathrin Coated Vesicles from Developing Pea331-332 J M Butler and L Beevers
Uncoating of Plant Clathrin Vesicles by Uncoating ATPase from Peas333-334 T Kirsch and L Beevers
LYSOSOMES AND DEGRADATION
Selective Degradation of Cytosolic Proteins by Lysosomes335-338 J F Dice
Role of Calcium, Protein Phosphorylation and the Cytoskeleton in HepatocyticAutophagy339-349 P O Seglen, I Holen and P B Gordon
Signals for Transport From Endosomes to Lysosomes
A New Receptor for Lysosomal Proenzymes
A Protein Kinase/Lipid Kinase Complex Required for Yeast Vacuolar ProteinSorting363-366 J H Stack, P K Herman and S D Emr
The Morphology But Not the Function of Endosomes and Lysosomes is Affected by Brefeldin A
Purification of the N-Acetylglucosamine-1-Phosphodiester &-N-Acetylglucosamindase from Human Lymphoblasts
Altered Regulation of Protein Degradation in Tranformed Human Bronchial Epithelial Cells
An Examination of the Structure and Functions of the Bovine Vacuolar ATPase Using Antisense Oligonucleotides
MOLECULES IDENTIFIED IMPORTANT IN MEMBRANE TRAFFIC
Regulation of Endocytosis by the Small GTP-ase Rab5

The Nucleotide Cycle of Sec4 is Important for its Function in VesicularTransport
Activators of Trimeric G-Proteins Stimulate and Inhibit Inter- compartmental Golgi Transport <i>In Vitro</i> 391-394 P J Weidman
Evidence of a Role for Heterotrimeric GTP-Binding Proteins and ARF in Endosome Fusion
The Small GTPase Rab4 Controls an Early Endocytic Sorting Event399-402 P van der Sluijs, M Hull and I Mellman
Protein Modifications and Their Significance in rab5 Function403-404 J C Sanford and M Wessling-Resnick
Low Molecular Weight GTP-Binding Proteins in Rough Endoplasmic Reticulum Membranes From Rat Liver and Rat Hepatocellular Carcinomas
Low Molecular Weight GTP-Binding Proteins Associated with the Membranes Involved in Post-Golgi Transport of Rhodopsin407-408 D Deretic and D S Papermaster
AuthorIndex
Subject Index

Isolation And Characterization Of A Functionally Active
Protein Translocation Apparatus From Chloroplast Envelopes.

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Chloroplast structure and function depends vitally on the import of nuclear coded and cytosolically synthesized polypeptide constituents (de Boer and Weisbeek 1991). Proteins of the outer and inner envelope from chloroplasts collaborate to form an import machinery which is responsible for the specific recognition of chloroplast destined precursor proteins and their translocation through the two membrane barrier. Outer envelope membrane vesicles are purified from pea chloroplasts in a right side-out orientation, i.e. like in the intact organelle (Waegemann et al. 1992). Precursor proteins are bound to the membrane vesicles in an ATP, receptor and transitpeptide dependent manner (Waegemann and Soll 1991, Soll and Waegemann 1992). The translocation process of a precursor proceeds via distinct steps which can be detected in vitro as translocation intermediates, named deg 1-4 in chloroplasts (Fig 1). The outer envelope localized import apparatus yields deg 1 and 2 while deg 3 and 4 are translocation intermediates which occur in connection with the inner envelope import machinery (Fig 1).

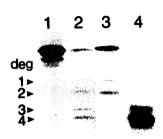


Fig 1) Localisation of pSSu translocation intermediates. pSSu binds to intact chloroplasts in the presence of 100 $\mu\rm M$ ATP (lane 1). Protease treatment results in the occurence of deg 1-4 (lane 2). Envelope membranes isolated from these plastids show that deg 3 and 4 are localized in the inner envelope (lane 4). Outer envelopes incubated with pSSu as above yield deg 1 and 2 after protease treatment (lane 3)

Using a solubilisation protocol of outer envelope membranes as outlined in Fig 2A (Kiebler et al. 1990) a protein complex could be enriched by sucrose density centrifugation which contained bound precursor protein (import complex I) (Fig 2B). Detection of this import complex I depended on the presence of ATP, a transit sequence and protease sensitive components, e.g. receptor molecules during the incubation of the membrane vesicle with the precursor (Waegemann and Soll 1991). The protein composition of import complex I is distinctively different from total outer envelope membrane proteins and shows an enrichment of a number of proteins as judged by gel electrophoresis followed by silver staining (Fig 2C). Using immunological technics we have identified four outer envelope proteins (OEP) of 86, 70, 75, and 34 kDa (Fig 2D). The 70 kDa polypeptide crossreacts with an antiserum against heatshock protein 70 (hsp 70). Furthermore it is possible to immunoprecipitate the precursor protein by antibodies against hsp 70 (Waegemann and Soll 1991) indicating a close interaction between these two components. Envelope localized hsc 70 might thus act as a chaperone to facilitate the translocation of a precursor protein by interacting with the transport competent, i.e. partially folded, conformation of the polypeptide chain (Marshall et al. 1990, von Heijne and Nishikawa 1991, Waegemann and Soll 1991). The role and function of other import complex I constituents remains to be established.

After solubilisation of outer envelope membranes and centrifugation through sucrose-gradients the fractions which contained import complex I were pooled and subsequently incubated with precursor protein (see Fig. 2A, right panel). This isolated membrane complex recognized specifically the precursor polypeptide in relation to the mature form and interacted with it in an ATP dependent manner. Following protease treatment translocation intermediates deg 1 and 2, identical to those described above (Fig 1) for the organellar system were detected. Transit peptide dependent interaction of the isolated import complex with the precursor points to the

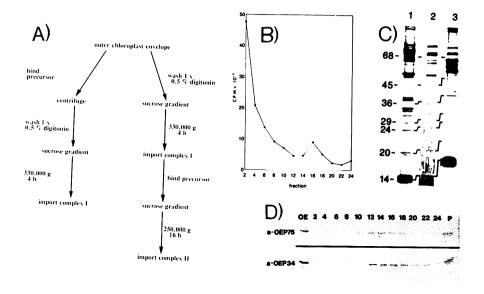


Fig 2) Characterization of the protein import complex from chloroplast outer envelopes. A) Solubilisation and isolation protocol of import complex I and II. B) Distribution of radioactivity in the sucrose density gradient as determined by liquid scintillation counting (line drawing). C) Polypeptide composition of outer chloroplast envelope (lane 1), complex I (lane 2) and complex II (lane 3). D) Immunoblot analysis of fractions obtained from a sucrose density gradient to enrich complex I using antisera against OEP 75 and OEP 34. Numbers on top indicate fraction numbers.

presence of a receptor in this complex. Protease sensitive components in import complex II are OEP 86 (Fig 3) and OEP 34 (see Fig 2C, D), however, direct evidence for their function as receptor is still missing. We were unable to detect a putative receptor of 30 kDa molecular (Pain et al. 1988, Schnell et al. 1990) in import complex II and conclude from this and from other results that the receptor for chloroplastic protein import is not yet conclusively identified (Flügge et al. 1991). The presence of hsc 70 in import complex II indicates again the involvement of this protein in the translocation mechanism (Fig 3). The protein composition of import complex I and II is similar (Fig 2C). To

our knowledge the results described above demonstrate for the first time that it is possible to isolate a protein translocation apparatus as a functional active unit from chloroplasts. This will enable us to study the mechanism of protein translocation in greater detail.

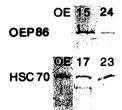


Fig 3) OEP 86 and a hsc 70 homologue are present in import complex II. Outer envelope membranes (OE) and samples of the sucrose density gradient (figures indicate fraction numbers) were tested using the respective antiserum.

Acknowledgement

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