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203. E. N. Meshalkin, I. I. Jevnina, E. I. Ivashkevitch, W. E. Davedova and N. N. Styshinskaja: On the functional state of the sympathetic-adrenal system and of the adrenal cortex in heart diseases.
204. A. Pekkarinen, K. Manninen and B. Thomasson: Effect of chlorprothixene, chlorpromazine and amitriptyline on the adrenaline and noradrenaline content in the adrenal vein plasma of dogs during irreversible haemorrhagic shock.
205. I. Suramo, S. Saarikoski and A. Pekkarinen: The effect of irreversible haemorrhagic shock on the fluorescence reaction of adrenergic nerve fibres and the content of noradrenaline and adrenaline in the heart, spleen and kidney of rabbits.

## INDUCTION OF OVULATION

206. G. Hellinga and H. J. M. Langedijk: Induction of menstruation, ovulation and pregnancy with Sexovit (F 6066).
207. M. Arnold, M. Berger, M. Keller, R. H. H. Richter and A. Uettwiller: Clinical and biochemical studies in patients treated with bis (p-acetoxyphenyl) cyclohexylidene methane (compound F 6066).
208. G. Bettendorf, M. Breckwoldt, P.-J. Czygan, C. Bordasch and K.-D. Schulz: Clinical studies with Clomid.
209. H. Schmidt-Elmendorff and E. Kaiser: Some observations on the induction of ovulation with gonadotrophins in women.
210. K.-D. Schulz, F. Hölzel and G. Bettendorf: The distribution of C<sup>14</sup>-Clomid (MRL-41) in various organs of immature female guinea pig.

## MAMMARY GLAND, etc.

211. M. Görlich and E. Heise: Biochemical parameters related to hormone therapy of chemically induced mammary carcinomas of the rat.
212. S. Sander: The uptake of oestradiol-17 in the normal mammary gland. An experimental study in rats.
213. N. Deshpande, V. Jensen and R. D. Bulbrook: Accumulation of <sup>3</sup>H-oestradiol by the human breast tissue.
214. R. M. Das and G. K. Benson: Uterine influences on the corpus luteum of the guinea-pig ovary.

## OESTROGENS

215. H.-G. Kraft and H. Kieser: Antifertility effects of anti-oestrogenic compounds in rats.
216. C. A. Michael and B. M. Schofield: The influence of hormones on myometrial development.
217. L. Martin: The oestrogenic activity of dimethylstilboestrol (DMS) and 16-oxo-oestradiol.
218. V. Sele: The effect of oestrogen-progestogen therapy on the development of secondary sex characters in women with severe primary hypogonadism.
219. J. van der Vies and H. Feenstra: The effects of ovarian hormones on the placenta of rats.

## READ BY TITLE

220. K. A. Ferguson, L. Lazarus, P. van Dooren and J. D. Young: The nature of the growth-promoting substances in human plasma.
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222. F. A. de la Balze, V. Goldberg, C. di Paola, J. C. De Paoli, E. Remy Sola, R. Dendukes and J. Cordero Funes: Urinary excretion of free, solvolizable and glucuronoconjugated 17-OHCS in normal, obese and carcinomatous females.
223. F. A. de la Balze, M. Janches, R. Dendukes, R. C. Socolsky and J. C. De Paoli: Excretion of urinary 17-HOCS in patients with pituitary pathology during administration of a bacterial pyrogen.
224. I. R. McDonald and M. Weiss: Turnover and excretion of cortisol in the Australian marsupial *Trichosurus vulpecula*.
225. A. C. Crooke and P. V. Bertrand: The outcome of pregnancy in infertile women after treatment with human gonadotrophins.
226. E. Z. Naugolnikh: 16-*epi*oestriol determination in urine during normal menstrual cycle.

II. Medizinische Klinik der Universität München, Germany

IN VITRO  $^{14}\text{C}$ -GLYCINE INCORPORATION INTO PROTEINS OF  
DENERVATED RAT MUSCLE, EFFECT OF NANDROLONE-DECANOATE

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In vitro incorporation of  $^{14}\text{C}$ -glycine into protein was studied in incubations of 1 ml 15 000 x g supernatant (SN) of rat gastrocnemius and extens. digit. long. muscle homogenates (200 mg per ml medium, consisting of 0.25 M sucrose, 0.08 M KCl, 0.01 M  $\text{MgCl}_2$  and 0.05 M tris-HCl buffer pH 7.6).  $^{14}\text{C}$ -glycine ( $0.053 \mu\text{mol} = 2 \mu\text{C}$ ), 2  $\mu\text{mol}$  ATP, 0.6  $\mu\text{mol}$  GTP, 6  $\mu\text{mol}$  phosphoenolpyruvate and 25 U pyruvic kinase were added (total volume = 2 ml).

Groups of rats were denervated (sciatic nerve), or sham operated (controls) 4 weeks prior to incubations. Incorporation of  $^{14}\text{C}$ -glycine was increased ( $p < 0.0005$ ) in denervated muscle:  $22.7 \pm 1.2$  cpm/mg protein (mean of 4 experiments with 3–5 incubations each  $\pm$  S.E.) against controls ( $11.2 \pm 0.04$  cpm/mg protein). In an attempt to determine whether increased  $^{14}\text{C}$ -glycine was due to factor(s) of soluble or microsomal cell fraction "criss-cross" incubations of denervated or sham operated 105 000 x g SN and microsomes were performed ( $\bar{x} \pm$  S.E.): microsomes (denerv.) + sol. cell fraction (denerv.) =  $46.0 \pm 7.5$ ; micr. (sham op.) + sol. cell fx. (denerv.) =  $29.6 \pm 4.3$ ; micr. (denerv.) + sol. cell fx. (sham op.) =  $25.4 \pm 3.8$ ; and micr. (sham op.) + sol. cell fx. (sham op.) =  $19.8 \pm 1.3$  cpm/mg protein. Stimulation of  $^{14}\text{C}$ -glycine incorporation appears to be due to the soluble cell fraction of denervated muscle rather than to denervated muscle microsomes, although some stimulation against control microsomes in incubations with sham operated muscle soluble cell fraction is apparent. Conceivably stimulatory material of soluble cell fraction was not completely removed upon preparation of microsomes.

No effect on  $^{14}\text{C}$ -glycine incorporation was obtained when normal rats were treated for 4 weeks with nandrolone-decanoate (10 mg/kg, once weekly): treated rats =  $13.1 \pm 0.8$ , placebo =  $12.5 \pm 0.3$  cpm/mg protein (4 experiments with 4 incubations each). This treatment produced slight, but so far (4 experiments) insignificant reduction of elevated  $^{14}\text{C}$ -glycine incorporation of denervated rat muscle ( $33.8 \pm 2.8$ ) against control ( $35.6 \pm 5.0$  cpm/mg protein).

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