Therapeutic potential of targeting interleukin-1 family cytokines in chronic inflammatory skin diseases*

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Summary

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Introduction

Cytokines of the interleukin-1 family

The interleukin (IL)-1 family comprises a large group of cytokines, partially sharing a conformational structure, a common receptor-binding mode, and a similar signalling pathway.¹ All members, albeit to different degrees, are key molecules involved in a myriad of immunological responses, primarily orchestrating innate immunity and bridging innate and adaptive immune responses. The IL-1 family encompasses 11 cytokines, seven with agonistic effects on their receptors and four with antagonistic effects (Table 1).^{2,3} IL-1 α , IL-1 β , IL-36 α , IL-36 β , IL-36 γ , IL-33 and IL-18 bind to a cognate membrane receptor to form a binary complex, and thereupon a coreceptor is recruited so as to form a signal-competent ternary complex (Figure 1).²

IL-1 α and IL-1 $\beta^{2,4}$ bind to a common transmembrane receptor IL-1R1, with subsequent recruitment of the coreceptor IL-1 receptor accessory protein (IL-1RAP, also named IL-1R3).

The interleukin (IL)-1 family of cytokines is a central regulator of a myriad of immunological responses. It comprises several cytokines, including those belonging to the IL-1, IL-36 and IL-18 subfamilies, as well as IL-33. The IL-1 family primarily plays a role in orchestrating innate immune responses, but is also involved in adaptive immunity. Increased interest in the IL-1 family occurred following the discovery that dysregulation of IL-1 signalling underlies the pathogenesis of several monogenic autoinflammatory diseases, characterized by sterile inflammation involving the skin and other organs. This also provided increased understanding of the role of innate immunity and the IL-1 family in polygenic autoinflammatory skin conditions, such as neutrophilic dermatoses, as well as in some of the most common chronic inflammatory skin diseases, such as psoriasis and hidradenitis suppurativa. Several therapeutic agents have been developed to inhibit the IL-1 family members and their signalling pathways. These have shown therapeutic efficacy in several chronic inflammatory skin disorders. The aim of this review is to thoroughly describe the consequences of pathological dysregulation of the IL-1, IL-33, IL-36 and IL-18 pathways in dermatological conditions and to provide a forward-looking update on therapeutic strategies targeting signalling by IL-1 family cytokines.

Conversely, the antagonistic cytokine IL-1 receptor antagonisti (IL-1Ra) engages the binding site of IL-R1 without recruiting the coreceptor IL-1R3, thus blocking signal transmission.²

The IL-36 subfamily comprises IL-36 α , IL-36 β , IL-36 γ , IL-36Ra and IL-38, with the latter two exerting an antagonistic function.⁵ The IL-36 receptor (IL-36R, IL-1R6) is the common receptor of all five members, which upon binding of IL-36 $\alpha/\beta/\gamma$ recruits the coreceptor IL-1R3, forming a ternary complex.⁶

IL-33 binds to its receptor IL-33R (ST2 or IL-1R4) with subsequent recruitment of IL-1R3.^{3,7} Finally, IL-18 and IL-37 belong to the IL-18 subfamily and exert pro- and antiinflammatory effects, respectively.⁸ IL-18 binds to the receptor IL-18R α (IL-1R5, IL-1Rrp1), recognized by the coreceptor IL-18R β (IL-1R7), thus also forming a ternary complex.^{9,10}

Receptors of the interleukin-1 family

The IL-1 receptor family includes 10 members, numbered from IL-1R1 to IL-1R10 (Table 2). IL-1R1 (IL-1RI, CD121a),

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Cytokine	Gene name	Other gene names	Chromosomal location	Property	Receptor (alternative names)	Coreceptors (alternative names)
IL-1 α	IL1A	IL1F1	2q14	Proinflammatory	IL-1R1 IL-1R2 (decoy receptor)	IL1-R3 (IL-1RacP, IL1RAP)
IL-1 β	IL1B	IL1F2	2q14	Proinflammatory	IL-1R1 IL-1R2 (decoy receptor)	IL1-R3 (IL-1RacP, IL1RAP)
IL-1Ra	IL1RN	IL1F3, IL1RA	2q14.2	Antagonist for IL-1α, IL-1β	IL-1R1 IL-1R2 (decoy receptor)	
IL-36a	IL36A	IL1F6, FIL1E, IL1E	2q12-q14.1	Proinflammatory	IL-R6 (IL-36R, IL-1Rrp2)	IL1-R3 (IL-1RacP, IL1RAP)
IL-36β	IL36B	IL1F8, IL1H2	2q14	Proinflammatory	IL-R6 (IL-36R, IL-1Rrp2)	IL1-R3 (IL-1RAcP, IL1RAP)
IL-36γ	IL36G	IL1F9, IL1E, IL1H1, IL1RP2, UNQ2456/PRO5737		Proinflammatory	IL-R6 (IL-36R, IL-1Rrp2)	IL1-R3 (IL-1RAcP, IL1RAP)
IL-36Ra	IL36RN	L1F5, FIL1D, L1HY1, L111, L1RP3, UNQ1896/PRO4342	2q14	Antagonist for IL-36α, IL-36β, IL-36γ	IL-R6 (IL-36R, IL-1Rrp2)	
IL-38	IL1F10	FIL1T, IL1HY2, IL38, FKSG75, UNQ6119/PRO20041	2q13	Anti-inflammatory	IL-R6, IL-36R, IL-1Rrp2 IL-R9	Unknown
IL-18	IL18	IL1F4, IGIF	11q22.2–q22.3	Proinflammatory	IL-1R5	IL-R7 (IL-18Rβ, IL18RacP)
IL-33	IL33	IL1F11, C9orf26, IL1F11, NFHEV	9p24.1	Proinflammatory and T helper 2 responses	IL-1R4 (IL-33R, ST2)	IL1-R3 (IL-1RAcP, IL1RAP)
IL-37	IL37	FIL1Z, IL1F7, IL1H4, IL1RP1	2q12-q14.1	Anti-inflammatory	IL-1R5 (IL-18R-1, IL-18Ra, IL-1Rp)	IL-R8 (TIR8, SIGIRR)

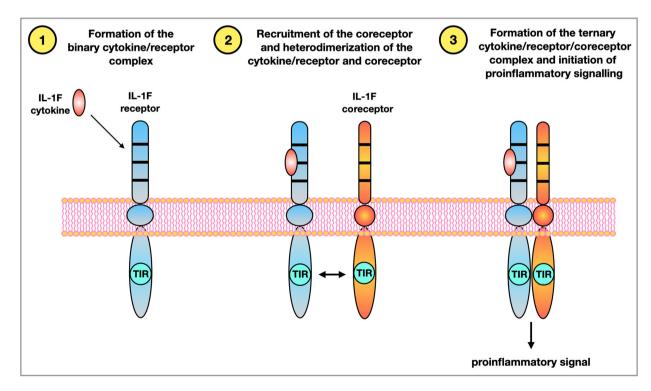


Figure 1 Most interleukin (IL)-1 family cytokines bind the cognate membrane receptor to form a so-called binary complex. Thereupon, the coreceptor, necessary for proinflammatory signal transmission, is recruited (ternary complex).

Human chromosome 2q12 2q12 3q28 2q12 2q12

2q12

2q12 2q12

11p15.5

SIGIRR

able 2 Rec	eptors of the interleukin (IL)-1R	. family		
Name	Alternative name(s)	Ligands	Coreceptor (alternative names)	Human gene
IL-1R1	IL-1RI, CD121a	IL-1α, IL-1β, IL-1Ra, IL-38	IL1-R3 (IL-1RacP, IL1RAP)	IL1R1
IL-1R2	IL-1RII, CD121b	IL-1α, IL-1β, IL-1Ra, pro-IL-1α		IL1R2
IL-1R3	IL-1RacP	No ligand known, accessory chain		IL1RAP
IL-1R3b	IL-1RacPb	No ligand known, accessory chain		IL1RAP
IL-1R4	T1, ST2, ST2L, DER4, Fit-1, IL-33Rα	IL-33	IL1-R3 (IL-1RacP, IL1RAP)	IL1RL1
IL-1R5	IL-18Rα, IL-1Rrp, IL-1Rrp1, CD218a	IL-18, IL-37	IL-R8 (TIR8, SIGIRR)	IL18R1
IL-1R6	IL-1Rrp2, IL-1RL2, IL-36R	IL-36α, IL-36β, IL-36γ, IL-36Ra, IL-38	IL1-R3 (IL-1RacP, IL1RAP)	IL1RL2
IL-1R7	IL-18Rβ, IL-18RAcP, AcPL, CD218b	No ligand known, accessory chain		IL18RAP

No ligand known, inhibitory receptor

IL-1R4 (ST2, IL-33R α) and IL-1R6 (IL-36R, IL-1Rrp2) are the ligand-binding chains for IL-1, IL-33 and IL-36, respectively.¹¹ They all use IL-1R3 as an accessory protein to form the ternary complex and induce intracellular signalling. In contrast, the ternary receptor complex of IL-18 consists of the main receptor IL-18R α and the accessory protein IL-18R β .¹² Soluble forms of IL-1 family receptors also exist, mostly acting as negative regulators (Table 3).¹³

Interleukin-1 receptor accessory protein

TIR8, SIGIRR

TIGIRR-2

TIGIRR-1

IL-1R8

IL-1R9

IL-1R10

First described in 1995,¹⁴ IL-1R3 is an accessory receptor of the IL-1 family. It is not directly involved in ligand binding, although it is crucial for the constitution of the three highaffinity ternary complexes necessary for IL-1, IL-33 and IL-36 signal transmission. IL-1R3 interacts with a composite surface created by IL-1R bound to the ligand, allowing the formation of a ternary complex and the initiation of an intracellular signalling pathway.¹⁵ Subsequent activation of several kinases,

Table 3 Soluble	receptors	of the	interleukin	(IL)-1R family
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Name	Other names	Ligands
sIL-1R1	sIL-1RI	IL-1α, IL-1β, IL-1Ra, IL-38
sIL-1R2	sIL-1RII	IL-1α, IL-1β, (IL-1Ra),
		pro-IL-1β
sIL-1R3	sIL-1RacP, sIL-1RAP	None (accessory chain)
sIL-1R3β	sIL-1RacP, sIL-1RAP-b	None (accessory chain)
sIL-1R4	sST2	IL-33
sIL-1R5		IL-18 (IL-37)
sIL-1R6		IL-36α, IL-36β, IL-36γ,
		IL-36Ra, IL-38
sIL-1R7		None (accessory chain)
sIL-1R8		No ligand known
sIL-1R9		IL-38
sIL-1R10		No ligand known
IL-18BP		IL-18, IL-37

especially nuclear factor-κB and mitogen-activated protein kinase, promotes the transmission of a strong proinflammatory signal to the cell nucleus.

Recent research has demonstrated a major role of IL-1 family cytokines in certain chronic inflammatory skin diseases (Figure 2), and the potential of blocking one or more of the family members is being explored in depth (Figure 3).^{16,17} The aim of this review is to thoroughly describe the consequences of pathological dysregulation of IL-1 family signalling in chronic inflammatory skin disorders and to provide an update on the therapeutic strategies targeting these pathways.

Interleukin-1 in skin diseases

IL-1 α is expressed constitutively in epithelial and mesenchymal cells of healthy individuals. This cytokine is active in its pro-form and can perform a dual function, either in the nucleus as a transcription factor or in the extracellular environment.¹⁸ Conversely, activation of IL-1β requires proteolytic cleavage mediated by the NLRP3 inflammasome, a cytoplasmic innate immune protein complex.¹⁹ This cytokine is produced mainly by activated macrophages, but also by other cell types including keratinocytes.²⁰ Activation of the IL-1 pathway promotes a proinflammatory signal, well described in the pathogenesis of various chronic skin diseases (Table 4).²¹

Interleukin-1 in monogenic autoinflammatory skin disorders

Autoinflammatory diseases (AIDs) are primarily characterized by sterile inflammation, with innate immunity playing the primary pathophysiological role. The stigmata of classic autoimmune diseases are typically absent.²² Among AIDs, the socalled inflammasomopathies are directly caused by gain-offunction mutations in components of the inflammasome.

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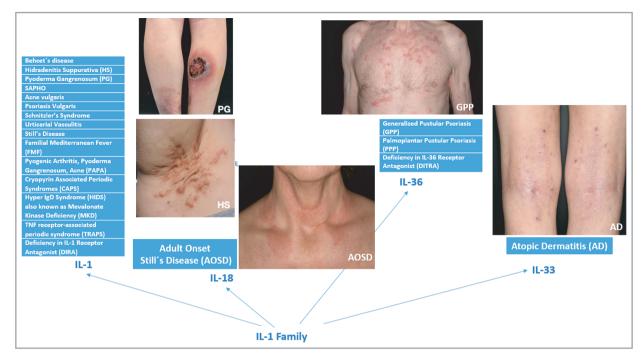


Figure 2 The main dermatological diseases with solid evidence of involvement of interleukin (IL)-1, IL-36, IL-33 and IL-18 in their pathogenesis. SAPHO, synovitis, acne, pustulosis, hyperostosis, osteitis; TNF, tumour necrosis factor.

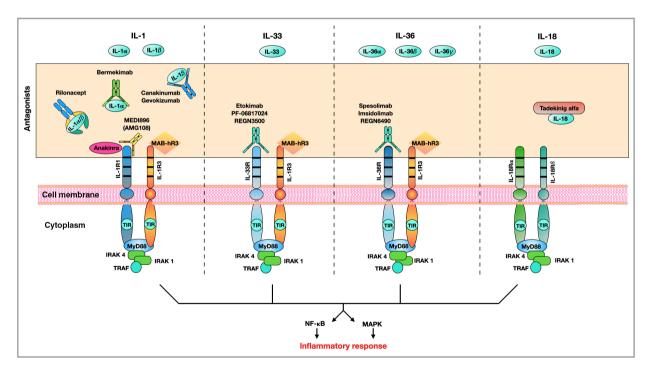


Figure 3 Schematic representation of seven members of the interleukin (IL)-1 family (IL-1 α , IL-1 β , IL-36 α , IL-36 α , IL-36 β , IL-36 γ and IL-18) acting on their receptor complex with transmission of a proinflammatory signal to the nucleus. Multiple inhibitors of the inflammatory cascade at different levels are also shown. IRAK, interleukin-1 receptor-associated kinase; MAB-hR3, monoclonal antibody targeting IL-1R3; MAPK, mitogenactivated protein kinase; NF, nuclear factor; TIR, Toll–interleukin-1 receptor; TRAF, tumour necrosis factor receptor-associated factor.

These mutations lead to dysregulation of IL-1 β signalling, as is the case in two prototypical AIDs: familial Mediterranean fever and cryopyrin-associated periodic syndromes.^{23–25}

Additionally, the IL-1 pathway can be imbalanced by the lack of counter-regulatory mechanisms, as in deficiency of IL-1Ra (DIRA), first described in 2009.²⁶ DIRA is caused by

Table 4 Current evidence for a pa	athogenic role of interleukin-1 in cutaneous diseases
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Strong ^a	Possible ^b	Uncertain ^c
Adult-onset Still disease ^{123,153,154}	Sweet syndrome ^{125,155,156}	PAPASH ¹²⁸
Behçet disease ^{121,157–160}	PASH ¹⁶¹	Rosacea ¹⁶²
Hidradenitis suppurativa ^{135,136,140}	PFAPA ¹⁶³	Acute generalized exanthematous
Pyoderma gangrenosum ³⁵	Generalized pustular psoriasis (GPP) ^{132–134}	pustulosis (AGEP) ⁹⁰
SAPHO ¹³⁰	Palmoplantar pustular psoriasis (PPP) ¹³⁴	Atopic dermatitis ^{164,165}
Acne vulgaris ¹⁴²	Dermatomyositis ¹⁶⁸	Allergic contact dermatitis ^{166,167}
Psoriasis vulgaris ¹⁴²	Panniculitis ¹²⁸	Irritant contact dermatitis ^{169,170}
Schnitzler syndrome ^{119–121,172}	Erdheim–Chester syndrome ^{173–175}	Mastocytosis ¹⁷¹
Urticarial vasculitis ¹⁷⁸	Deficiency of adenosine deaminase (DADA2) ¹⁷⁹	Systemic sclerosis ^{176,177}
Familial Mediterranean fever (FMF) ¹⁸¹	Majeed syndrome ¹⁸²	Chronic spontaneous urticaria ¹⁸⁰
Pyogenic arthritis, pyoderma	Deficiency of IL-36 receptor antagonist (DITRA) ¹³²	Autoimmune blistering diseases ¹⁸³
gangrenosum, acne (PAPA) ¹⁸⁴	Haploinsufficiency of A20 (HA20) ¹⁸⁷	Vitiligo ¹⁸⁵
Cryopyrin-associated periodic		CARD-14 mediated pustular
syndromes (CAPS) ¹⁸⁶		psoriasis (CAMPS) ²²
Hyper-IgD syndrome (HIDS),		Familiar keratosis lichenoides chronica
also known as mevalonate kinase		(FKLC), multiple self-healing
deficiency (MKD) ¹⁸⁸		palmoplantar carcinoma (MSPC) ²³
TNF receptor-associated periodic		Pyrin-associated autoinflammation with
syndrome (TRAPS) ¹⁸⁸		neutrophilic dermatosis (PAAND) ¹⁸⁹
Deficiency of IL-1 receptor		NLRC4-related macrophage activation
antagonist (DIRA) ²⁶		syndrome (NLRC4-MAS) ²³

PASH, pyoderma gangrenosum, acne, hidradenitis suppurativa; PAPASH, pyogenic arthritis, pyoderma gangrenosum, acne, hidradenitis suppurativa; PFAPA, periodic fever, aphthous stomatitis, pharyngitis, adenitis; SAPHO, synovitis, acne, pustulosis, hyperostosis, osteitis. ^aStrong evidence: clinical trials showing efficacy of selective cytokine antagonism in a cohort of patients. ^bPossible: case reports or series reporting efficacy of selective cytokine antagonism in a single patient or a small number of patients. ^cUncertain: biological evidence concerning the role of interest of the cytokine in the disease.

mutations in the IL1RN gene and presents clinically as neonatal onset of generalized cutaneous pustulosis, multifocal osteomyelitis, and high levels of acute-phase reactants.

Interleukin-1 in neutrophilic dermatoses

Neutrophilic dermatoses (NDs) are chronic inflammatory skin disorders characterized by neutrophil-driven sterile cutaneous inflammation. Two of the most prototypical NDs include Sweet syndrome and pyoderma gangrenosum (PG). Sweet syndrome typically occurs in individuals aged 47-57 years, with a slight female predominance, and is characterized by the sudden appearance of painful, oedematous and erythematous papules, plaques or nodules on the skin associated with fever and leucocytosis. PG presents, in its classic form, with rapidly developing, painful skin ulcers with undermined borders and violaceous peripheral erythema. The incidence of PG is approximately six cases per million person-years, with an average age of onset between 40 and 60 years.²⁷ The pathogenesis of ND is so far not completely elucidated. A complex interplay between an imbalanced expression of inflammatory molecules, abnormal neutrophil function, and genetic predisposition contributes to the onset of ND. In the end, extravasation of activated neutrophils and migration towards the source of the inflammatory chemoattractant cause inflammation and tissue damage.²⁸

Dysregulation of innate immune pathways is regarded as one of the predominant mechanisms underlying the pathophysiology of ND.²⁹ Indeed, IL-1 β has been shown to promote the generation of T helper (Th)17 cells, which can amplify the recruitment of neutrophils.^{30,31} This cytokine both acts on neutrophils, exerting antiapoptotic effects and thus promoting their survival,³² and is produced by neutrophils, mainly in an inflammasome-dependent manner.³³ Because of numerous clinical and pathogenic similarities and the frequent response to IL-1-targeted therapy, NDs are now considered to be predominantly autoinflammatory in nature.²² Indeed, IL-1 β gene expression and protein levels were found to be elevated in two prototypical NDs, Sweet syndrome³⁴ and PG.³⁵ Similarly, elevated serum levels of IL-1 β have been described in Behçet disease,³⁶ and IL-1 α was found to be upregulated in skin from a patient with amicrobial pustulosis of the folds.³⁷

Interleukin-1 in hidradenitis suppurativa

Hidradenitis suppurativa (HS) is a chronic inflammatory skin disorder that affects approximately 1% of the general population, with the highest prevalence reported in young adults. The disease manifests clinically as recurrent episodes of neutrophilic inflammation mostly involving skin bearing pilosebaceous and apocrine units (predominantly the axillary and inguinal folds and perianal area). The pathogenesis of HS has not been completely elucidated, although it is known that genetic, hormonal, immunological and microbial factors, together with tobacco smoking and obesity, contribute to

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disease occurrence and/or severity. The main events leading to the development of HS lesions encompass aberrant infundibular keratinization with consequent hyperkeratosis and occlusion, and the aberrant activation of innate immune pathways with a massive neutrophil-rich inflammatory infiltrate.³⁸ The pathogenic role of IL-1 β has recently been investigated in HS.³⁹

van der Zee et al. found a significant increase in IL-1 β , tumour necrosis factor (TNF)- α and IL-10 in the supernatants of ex vivo cultured HS lesional skin, compared with healthy controls and with psoriatic skin.⁴⁰ Kelly et al. described augmented protein levels of IL-1 β , IL-17 and TNF- α and enhanced NLRP3 and IL18 gene expression in lesional HS skin, supporting the pathogenic involvement of the inflammasome and IL-1B.41 Furthermore, Witte-Händel et al. showed that the IL-1 β pathway is clearly hyperactive in HS lesions, compared with psoriasis lesions and healthy skin, thus likely contributing to local and systemic inflammation.³⁹ In HS skin, IL-1 β was found to be produced mainly by monocytes and macrophages, whereas fibroblasts were the most potent producers of IL-1 β target molecules. Interestingly, this strong IL-1 β signature with downstream upregulation of matrix metalloproteinases, chemokines (including CXCL1, CXCL6, CXCL10 and CCL7) and several cytokines (IL-1 β , IL-6, IL-32 and IL-36), could be specifically reversed ex vivo by inhibition of IL-1β signalling with IL-1Ra.³⁹

Given the above experimental data, it would be expected that IL-1 pathway blockade could be of therapeutic benefit in patients with HS, and indeed, as discussed later in this review, some evidence for this exists.

Interleukin-1 in psoriasis

Psoriasis is an immune-mediated inflammatory disease with a chronic course and a multifactorial pathogenesis, which manifests as scaly itchy and/or painful patches and plaques on the skin. The prevalence of the disease varies among populations and ages within a range from 0.09% to 5.1%.⁴² The pathogenesis of psoriasis is based on a complex interaction between innate and adaptive immune compartments and relies on a predominant Th1/Th17 signature.⁴³

In psoriasis, the IL-1 pathway has a well-documented pathogenic role, albeit early in the pathogenesis.⁴⁴ IL-1 α is essential for the development of neutrophilic abscesses in the imiquimod-induced murine psoriasis-like model.⁴⁵ IL-1 β , produced by macrophages, dendritic cells and keratinocytes, is critical in Th17-cell differentiation and activation.^{46–48}

Interleukin-33 in dermatological diseases

IL-33 primarily plays a defensive role at barrier sites, being constitutionally expressed by keratinocytes.⁴⁹ Like IL-1 α , IL-33 can transmit its signal by acting as a transcription factor in the cell nucleus or in the extracellular environment.⁵⁰ This cytokine can stimulate group 2 innate lymphoid cells⁵¹ and predominantly drives Th2 polarization, thus playing a role in

allergic diseases and eosinophilic inflammation.⁵² IL-33 dysfunction has been investigated in a wide range of inflammatory skin diseases, including AD and psoriasis (Table 5).⁵³

Interleukin-33 in atopic dermatitis

Atopic dermatitis (AD) is the most common chronic inflammatory skin disorder, affecting an increasing number of patients globally, with a prevalence of up to 7% in adults and up to 25% among children.⁵⁴ The disease clinically manifests as itchy eczematous lesions predominantly localized on flexural areas and the face, neck and distal extremities. Classically considered to be mediated by a Th2-skewed adaptive immune response,⁵⁵ the immune map of the disease is actually far more complex and involves multiple inflammatory pathways guided by Th22, Th17/IL-23 and Th1 cytokines.⁵⁴

Transgenic mice with enhanced skin-selective expression of the IL33 gene have been shown to have AD-like dermatitis.⁵⁶ Also, serum IL-33 in AD skin, as well as IL-33 mRNA and protein levels, were found to be elevated.^{57,58} Current knowl-edge also indicates that IL-33 is able to induce Th2 cell differentiation and to promote IL-31 expression by Th2 cells, alone or in combination with IL-4.⁵⁹ IL-31 further stimulates the onset and persistence of itching, and directly downregulates the expression of claudin-1 and filaggrin, thus contributing to skin barrier impairment.^{60,61}

Interleukin-33 in psoriasis

Despite its original description as a Th2-driving cytokine, IL-33 also plays a role in psoriasis, classically considered a Th1/ Th17-mediated disease. IL-33 has been shown to be produced primarily by keratinocytes following psoriatic inflammatory stimuli and to induce the transcription of inflammation-related

 Table 5
 Current evidence for a pathogenic role of interleukin-33 in cutaneous diseases

Strong ^a	
Atopic dermatitis ^{149,150}	
Uncertain ^b	
Allergic contact dermatitis ¹⁶⁶	
Irritant contact dermatitis ¹⁹⁰	
Rosacea ¹⁹¹	
Psoriasis vulgaris ^{65,192,193}	
Pustular psoriasis ⁶⁷	
Mastocytosis ¹⁷¹	
Systemic lupus erythematosus ¹⁹⁴	
Systemic sclerosis ^{195,196}	
Chronic spontaneous urticaria ¹⁹⁷	
Autoimmune blistering diseases ¹⁹⁸	
Behçet disease ¹⁹⁹	
Vitiligo ²⁰⁰	

^aStrong evidence: clinical trials showing efficacy of selective cytokine antagonism in a cohort of patients. ^bUncertain: biological evidence concerning the role of interest of the cytokine in the disease. genes (such as CCL2, CXCL1, CXCL2, Cxcl15 and vascular endothelial growth factor genes) in keratinocytes, in an autocrine manner. 62

Also, in a murine model, IL-33 induced psoriasis-like lesions through interaction with mastocytes and neutrophils.⁶³ Additionally, several studies have found higher IL-33 expression levels in psoriatic lesions than in healthy skin,^{64–66} and some data are suggestive of an increase in IL-33 levels in the serum of patients with psoriasis.^{67,68}

Interleukin-36 in skin diseases

The three splice variants of IL-36 (IL-36 α , IL-36 β and IL-36 γ) are potent proinflammatory cytokines, mainly produced in barrier sites of the body (cutaneous, bronchial and intestinal epithelium).⁶⁹ In the skin, IL-36 γ is predominantly expressed in epidermal keratinocytes.^{70,71} These cytokines play a first-line defensive role against exogenous insults and contribute to maintaining cutaneous homeostasis. Furthermore, they contribute to crosstalk between the innate and adaptive immune responses, for example by stimulating Th-cell activation and Th1 polarization.⁷² IL-36 pathway dysfunction is associated with selected inflammatory skin diseases (Table 6).⁷³

Interleukin-36 in psoriasis

The role of IL-36 in skin diseases received great attention when loss-of-function mutations in the gene IL36RN, encoding IL-36Ra, were discovered as a cause of a severe recessive autoinflammatory syndrome named deficiency of IL-36 receptor antagonist (DITRA).⁷⁴ The reported mutations influence

Table 6 Current evidence for a pathogenic role of interleukin (IL)-36 in cutaneous diseases

Strong ^a	
Generalized pustular psoriasis ¹⁴⁵	
Palmoplantar pustular psoriasis ¹⁴⁶	
Deficiency of IL-36 receptor antagonist (DITRA) ⁷⁴	
Uncertain ^b	
Psoriasis vulgaris ^{86,194}	
CARD14-mediated psoriasis ²²	
Acute generalized exanthematous pustulosis ⁹¹	
Hidradenitis suppurativa ^{92–94}	
Pyoderma gangrenosum ³⁵	
Sweet's syndrome ²²	
Systemic lupus erythematosus ²⁰¹	
Systemic sclerosis ²⁰²	
Autoimmune blistering diseases ²⁰³	
Acne ⁹⁵	
Atopic dermatitis ²⁰⁴	
Allergic contact dermatitis ¹⁶⁶	
Folliculitis and eosinophilic pustular folliculitis ²⁰⁵	

^aStrong evidence: clinical trials showing efficacy of selective cytokine antagonism in a cohort of patients. ^bUncertain: biological evidence concerning the role of interest of the cytokine in the disease. the stability of IL-36Ra and its ability to bind to IL-1R6, thus limiting its ability to inhibit IL-36 signalling. DITRA presents clinically with episodic fever and generalized pustular psoriasis (GPP). Loss-of-function mutations or single-nucleotide polymorphisms of IL36RN have also been identified in 23–37% of sporadic forms of GPP.^{75–77} Although the frequency of IL36RN gene mutations in palmoplantar pustular psoriasis (PPP) is < 5%,⁷⁸ gene expression levels of IL36G in PPP are higher than in normal skin, suggesting that IL-36 pathway signalling is also upregulated in PPP.⁷⁹

Furthermore, a severe pustular or erythrodermic psoriasis phenotype named CAMPS (CARD-14-mediated pustular psoriasis) has been described, caused by autosomal dominant gain-of-function mutations in CARD-14.⁸⁰ CARD-14 is strongly expressed in keratinocytes and drives IL-1 β production and subsequent increased transcription of IL-8 and IL-36 γ .⁸¹

IL-36 cytokines are also relevant in the most frequent form of psoriasis, namely psoriasis vulgaris.⁸² These cytokines are released mainly by keratinocytes upon stimulation by Toll-like receptor agonists or proinflammatory cytokines (TNF- α , IL-17 and IL-22), but are also produced by endothelial and immune cells.⁸³ In psoriasis, IL-36 cytokines influence the cornification processes in the epidermis by acting on keratinocytes, induce the production of Th17- and Th1-polarizing cytokines by myeloid dendritic cells and macrophages, and potently sustain neutrophil recruitment. In lesional psoriatic skin, IL-36 α , IL-36 γ and IL-36Ra are indeed highly expressed.^{70,84} Conversely, the antagonist IL-38 is downregulated.⁸⁵ Particularly, the IL-36 γ isoform, which is not normally expressed in healthy skin, appears to be crucial in psoriasis and has been suggested as a biomarker for disease activity.⁸⁶

Interleukin-36 in neutrophilic dermatoses

IL-36 cytokines have a strong ability to recruit neutrophils to the skin⁸⁷ and, in turn, neutrophil-derived proteases process IL-36 cytokines, enhancing their biological activity.⁸⁸ This suggests that neutrophils are key players in escalating IL-36driven inflammation, and that IL-36 may be central to the pathogenesis of NDs.¹⁶ Indeed, in lesional PG skin, Kolios et al. reported selective upregulation of IL36A mRNA, whereas IL36G mRNA was not elevated.³⁵

IL-36 has also been shown to play a role in acute generalized exanthematous pustulosis (AGEP), a severe adverse cutaneous drug reaction that shares certain phenotypical and histological features with GPP.^{89,90} IL-36 γ expression is strongly increased in the epidermis during AGEP, and culprit drugs can specifically stimulate keratinocytes to secrete IL-36 γ with subsequent IL-8 production by macrophages and T cells, thus driving neutrophil recruitment and survival in lesional AGEP skin.⁹¹

Furthermore, IL-36 has been shown to be a driver in the pathogenesis of the acneiform eruption induced by epidermal growth factor receptor and MEK (MAPK kinase) inhibitors. In fact, these targeted drugs are able, by synergizing with the commensal Cutibacterium acnes, to potently induce keratinocyte

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IL-36 γ expression and drive IL-8-mediated neutrophil-rich inflammation. 92

Interleukin-36 in hidradenitis suppurativa

As in psoriasis, in HS skin, keratinocytes are primarily responsible for the secretion of IL-36 cytokines. These cytokines can target different types of cells, from keratinocytes to dendritic cells, promoting the production of proinflammatory cytokines, such as IL-12 and IL-23, which selectively provoke adaptive immunity (Th1 and Th17 responses). In turn, dendritic cells also produce IL-36, thus establishing an autocrine loop that further amplifies inflammation.93 Several studies have shown significantly high protein and gene expression levels of IL-36 α , IL-36 β and IL-36 γ in lesional HS skin, as well as serum levels in patients with HS compared with healthy controls.^{93–95} Furthermore, Wolk et al. demonstrated in HS skin high levels of granulocyte colony-stimulating factor, a major driver of neutrophil recruitment and survival, which were induced by IL-36.96 Moreover, currently available data are supportive of a pivotal role of the IL-36 cytokine family in orchestrating the crosstalk between keratinocytes and immune cells in HS, thus likely contributing to the chronic inflammation.93

Interleukin-18 in dermatological diseases

Originally described as IGIF (interferon- γ -inducing factor), IL-18 can exert pleiotropic functions, mainly depending on the surrounding cytokine milieu.⁹⁷ IL-18 is constitutively expressed by human keratinocytes⁹⁸ and exerts strong proinflammatory activity. The biological action of IL-18 is neutralized by IL-18BP and IL-37. IL-18BP is an endogenous soluble factor that prevents IL-18 from binding to IL-18R, thus suppressing interferon (IFN)- γ production and inhibiting Th1 immune responses.¹¹ Dysregulation of the IL-18 pathway has been reported in several cutaneous diseases (Table 7).

Interleukin-18 in adult-onset Still disease

Adult-onset Still disease (AoSD) is a systemic inflammatory condition typically affecting young adults and traditionally characterized by four symptoms: fever, arthralgia, cutaneous eruption and leucocytosis. The estimated annual incidence is approximately 0.16 cases per 100 000 people.⁹⁹ Crucial for the pathogenesis of AoSD is an intense activation of the innate immune system, with several proinflammatory cytokines suggested to be involved, including IL-1 β , IL-6, TNF- α , IFN- γ and IL-18.¹⁰⁰ Particularly, IL-18 and IL-1 β appear to be crucial in initiating the proinflammatory cascade in AoSD.¹⁰¹ IL-18 is produced mainly by macrophages in an NLRP3 inflammasome-dependent manner and further promotes immune cells to produce a large amount of proinflammatory cytokines, including IL-6, IL-8, IL-17 and TNF- α , thus contributing to the so-called 'cytokine storm' in AoSD.¹⁰⁰ Indeed, several studies have shown high levels of serum IL-18 in systemic forms of AoSD, so this Table 7 Current evidence for a pathogenic role of interleukin-18 in cutaneous diseases

Strong ^a	
Adult-onset	Still disease ¹⁵¹
Uncertain ^b	
Cutaneous	lupus erythematosus ^{107,206}
Psoriasis ²⁰⁷	,208
Atopic derr	natitis ^{209,210}
Chronic spo	ontaneous urticaria ²¹¹
Contact der	rmatitis ²¹²
Alopecia ar	eata ²¹³
	drug eruptions ²¹⁴
Graft-versu	s-host disease ^{215,216}
Cryopyrin-a	associated periodic syndromes ²¹⁷
Granuloma	tosis with polyangitis ²¹⁸
Systemic sc	
Hidradeniti	s suppurativa ⁴¹
Pyogenic a	thritis, pyoderma gangrenosum and acne $(PAPA)^{22}$
Familial Me	editerranean fever ^{221,222}
Rosacea ²²³	
Synovitis, a	cne, pustulosis, hyperostosis, osteitis (SAPHO) ²²⁴
Bullous per	nphigoid ²²⁵
Pemphigus	vulgaris ²²⁶
Behçet dise	
Schnitzler s	yndrome ²²⁸

^aStrong evidence: clinical trials showing efficacy of selective cytokine antagonism in a cohort of patients. ^bUncertain: biological evidence concerning the role of interest of the cytokine in the disease.

cytokine has been proposed as a promising biomarker for the diagnosis of AoSD. $^{\rm 102-105}$

Interleukin-18 in lupus erythematosus

Recent evidence has highlighted a relevant role of IL-18 pathway dysfunction in the pathogenesis of systemic lupus erythematosus. In particular, single-nucleotide polymorphisms in the IL18 gene have been shown to be associated with predisposition to systemic lupus erythematosus,¹⁰⁶ and IL-18 was found to be overexpressed in skin samples¹⁰⁷ and serum¹⁰⁸ of patients with cutaneous lupus. Indeed, IL-18 has been proposed as a predictive marker of disease activity.¹⁰⁹ In cutaneous lupus, IL-18 has been shown to induce apoptosis of keratinocytes by stimulating TNF- α expression in these cells and reducing the expression of IL-12, which instead is able to protect keratinocytes from TNF- α - and ultraviolet-induced apoptosis.¹¹⁰

Interleukin-18 in psoriasis

In psoriasis, cathelicidin LL-37-stimulated keratinocytes are able to produce IL-18, via activation of the NLRP3 inflammasome.¹¹¹ Subsequently, IL-18 can enhance IFN- γ production by Th1 cells and IL-17 secretion by Th17 lymphocytes, thus maintaining the inflammatory circuit underlying disease pathogenesis. Elevated skin and serum levels of IL-18 have been reported in patients with psoriasis and these correlate with disease severity.^{112,113}

Table 8 Current interleukir	(IL)-1 family	antagonists targeting	chronic inflammatory	v skin diseases
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			Published clinical	Ongoing clinical trial		
Drug	Mechanism of action	Approved indications (FDA and/or EMA)	trials beyond label indications	Disease(s)	Phase	Clinical trial number
Anakinra (Kineret®)	Competitive binding of IL-1 α and IL-1 β to the IL-1 receptor	FDA: RA and CAPS. EMA: FMF, SJIA, AoSD, rheumatoid arthritis and CAPS	HS: Leslie 2014, ¹³⁶ Tzanetakou 2016 ¹³⁷	Inflammatory pustular skin diseases AD (severe)	II	NCT01794117 NCT01122914
Rilonacept (Arcalyst®)	Soluble decoy receptor that binds	FDA: CAPS, recurrent idiopathic	FMF: Hashkes 2012. ²²⁹ SSc:	Cold contact urticaria	II	NCT02171416
	both IL-1 α and IL-1 β	pericarditis, maintenance of remission of DIRA	Mantero 2018. ²³⁰ Schnitzler syndrome: Krause 2012 ¹²¹	CAPS (MWS) or Schnitzler syndrome	Π	NCT01045772
Canakinumab	Human monoclonal	FDA: CAPS, TRAPS,	PG: Kolios 2015 ³⁵	Urticaria	II	NCT01635127
(Ilaris®)	antibody specific for IL-1β	HIDS/MKD, SJIA, FMF. EMA: CAPS, TRAPS, HIDS/MKD, SJIA, FMF, AoSD, gouty arthritis		Urticarial vasculitis Schnitzler syndrome	II II	NCT01170936 NCT01245127
Gevokizumab	Humanized	None	None	PG	II	NCT01882504
	monoclonal			PG	III	NCT02318914
	antibody specific			PG	III	NCT02326740
	for IL-1β			PG	III	NCT02315417
				Acne vulgaris	II I/II	NCT01498874
				CAPS (FCAS/MWS) and Behçet disease		NCT01211977
Bermekimab	Human monoclonal	None	HS: Gottlieb	AD	II	NCT04990440
	antibody specific for IL-1α		2020, ¹⁴¹ Kanni 2018. ¹⁴⁰ Acne vulgaris: Carrasco 2015. ¹⁴² Psoriasis: Coleman 2015 ¹⁴³	AD	II	NCT04791319
				HS SSc	II II	NCT04045742
				AD	II	NCT04045743 NCT04021862
				HS	II	NCT04988308
			Coleman 2015	AD	II	NCT03496974
ИЕDI8968 (AMG 108)	Human monoclonal antibody targeting IL-1R1: inhibition of its activation by IL-1α and IL-1β	None	None	HS	II, lack of	efficacy
NCT01838499	RPH-104	Macromolecular compound binding human IL-1β	None	None		Schnitzler syndrome
П	NCT04213274	ituillali IL-1p				
Spesolimab (BI	Humanized	None	GPP: Bachelez	HS	II	NCT04762277
655130)	monoclonal		2019 ¹⁴⁵	HS	II	NCT04876391
	antibody targeting			PPP	II	NCT04493424
	IL-36R			GPP	II	NCT04399837
				GPP	II	NCT03886246
				GPP	II	NCT03782792
				AD	II	NCT03822832
				AD	II	NCT04086121
				PPP	II	NCT03135548
	TT · 1	N),	PPP	II	NCT04015518
(ANIPO10)	Humanized	None	None	Acne vulgaris	II	NCT04856917
(ANB019)	monoclonal			HS	II	NCT04856930
	antibody targeting IL-36R			Acneiform rash Ichthyosis	II II	NCT04697069 NCT04697056
	IL-30I			,		
				GPP	II	NCT03619902

Table 8	(continued)
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Drug	Mechanism of action	Approved indications (FDA and/or EMA)	Published clinical trials beyond label indications	Ongoing clinical trial		
				Disease(s)	Phase	Clinical trial number
REGN6490	Monoclonal antibody targeting IL-36R	None	None	Healthy volunteers Healthy volunteers	I I	NCT04616105 NCT04616079
Etokimab	Monoclonal antibody targeting IL-33	None	AD: Chen 2019 ¹⁴⁹	AD	Π	NCT03533751
PF-06817024	Monoclonal antibody targeting IL-33	None	None	Chronic rhinosinusitis with nasal polyps and AD	Ι	NCT02743871
	Human monoclonal		None	AD	II	NCT03736967
	antibody targeting IL-33			AD	Π	NCT03738423
CNTO 7160	Monoclonal antibody targeting IL-33R	None	None	Asthma or AD	Π	NCT02345928
Tadekinig alfa	Recombinant human	None	None	AoSD NLRC4 mutation	II III	NCT02398435 NCT03512314
	interleukin-18- binding protein			and XIAP deficiency		

AD, atopic dermatitis; AoSD, adult-onset Still disease; CAPS, cryopyrin-associated periodic syndromes; DIRA, deficiency of interleukin-1 receptor antagonist; EMA, European Medicines Agency; FCAS, familial cold autoinflammatory syndrome; FDA, US Food and Drug Administration; FMF, familial Mediterranean fever; HIDS/MKD, hyperimmunoglobulin D syndrome/mevalonate kinase deficiency; GPP, generalized pustular psoriasis; HS, hidradenitis suppurativa; MWS, Muckle–Wells syndrome; PG, pyoderma gangrenosum; PPP, palmoplantar pustulosis; RA, rheumatoid arthritis; SJIA, systemic juvenile idiopathic arthritis; SSc, systemic sclerosis; TRAPS, tumour necrosis factor receptor-associated periodic syndrome.

Registered and emerging therapies specifically targeting interleukin-1 family members

Interleukin-1 antagonists

To date, several different therapeutic approaches have been developed to antagonize the IL-1 pathway. Among these are anakinra, 114,115 a recombinant IL-1Ra that simultaneously inhibits IL-1 α and IL-1 β ; rilonacept, 116 a recombinant soluble decoy receptor of IL-1 β that binds IL-1 β and, with lower affinity IL-1 α and IL-1Ra; and canakinumab, 117,118 a monoclonal antibody targeting IL-1 β . The above-mentioned drugs have received approval by the US Food and Drug Administration and/or the European Medicines Agency for certain inflammatory skin diseases and have been investigated in randomized clinical trials (Tables 4 and 8).

In detail, IL-1-blocking agents have shown therapeutic effects, albeit of differing levels, in Schnitzler syndrome,^{119–121} Behçet disease,¹²² Still disease,^{123–125} Sweet syndrome,¹²⁶ PG,^{35,127} neutrophilic panniculitis,¹²⁸ PG-associated autoin-flammatory syndromes [PASH (PG, acne, HS) and PAPASH (pyogenic arthritis, PG, acne, HS)],¹²⁹ SAPHO syndrome (synovitis, acne, pustulosis, hyperostosis, osteitis)¹³⁰ and PFAPA syndrome (periodic fever, aphthous stomatitis, pharyngitis and adenitis).¹³¹ In GPP, several reports have documented

therapeutic efficacy of anakinra,^{132,133} and also canakinumab,¹³⁴ whereas only partial and transient clinical remission was observed in patients with PPP treated with anakinra.¹³⁵ Anakinra has shown clinical efficacy also in more common diseases, such as HS, as demonstrated in a small open-label study¹³⁶ and in a randomized clinical trial.¹³⁷ Conversely, data concerning the efficacy of canakinumab in HS, from case reports and series, showed contrasting results, raising the possibility that IL-1 α may, in addition to IL-1 β , be an important inflammatory mediator in this disease.^{138,139}

Three additional investigational mAbs targeting the IL-1 pathway have been investigated in clinical trials: bermekimab (anti-IL-1 α), gevokizumab (anti-IL-1 β) and MEDI8968 (AMG 108; anti-IL-1R1). Also reported is RPH-104, a heterodimeric fusion protein that inhibits IL-1 β (Table 8). Bermekimab has already shown clinical efficacy in patients with HS, both in a randomized clinical trial¹⁴⁰ and in an open-label study.¹⁴¹ Additionally, the drug has shown encouraging results in open-label studies in acne¹⁴² and psoriasis.¹⁴³

Interleukin-36 antagonists

Three IL-36 antagonists, directly inhibiting IL-36R, are currently in the clinical development phase, namely spesolimab (BI 655130), imsidolimab (ANB019) and REGN6490. Spesolimab has shown efficacy in GPP treatment in a phase I clinical trial^{144,145} and is currently being investigated in phase II studies (Table 8). In a randomized clinical trial conducted on patients with PPP treated with spesolimab, improvement of disease severity was reported in all treatment groups, although the primary endpoint of the trial was not met.^{146,147} Spesolimab is currently being investigated in clinical trials for other skin disorders. Imsidolimab has shown a favourable safety profile in a phase I clinical trial in healthy volunteers.¹⁴⁸ Phase II clinical trials are currently ongoing. REGN6490 is currently being investigated in two phase I clinical trials on healthy volunteers (Table 8).

Interleukin-33 antagonists

Etokimab (ANB020) is a humanized IgG1 monoclonal antibody targeting IL-33, and encouraging data have been reported in a phase IIa proof-of-concept clinical trial conducted on patients with AD.¹⁴⁹ Currently etokimab is being investigated in a phase IIb trial (NCT03533751) in patients with AD. Two other monoclonal antibodies targeting IL-33 are currently under investigation in patients with AD: PF-06817024 and REGN3500 (Table 8).

Furthermore, blockade of IL-33 has been explored in AD using a monoclonal antibody targeting IL-33R (CNTO 7160). The safety and efficacy of this drug have been investigated in a phase I trial in patients with asthma, patients with AD and healthy individuals. While laboratory evidence showed inhibition of the IL-33 pathway, no significant clinical improvement was achieved.¹⁵⁰

Interleukin-18 antagonist

A human recombinant IL-18-binding protein (tadekinig alfa) has been developed and investigated in a phase II open-label clinical trial on 23 patients with AoSD, randomized to receive tadekinig alfa 80 mg or 160 mg subcutaneously, three times per week for 12 weeks. Tadekinig alfa demonstrated a favourable safety profile at both doses and early signs of efficacy, with a response rate of 50%, as endorsed by clinical and laboratory assessment.¹⁵¹

Interleukin-1 receptor 3 antagonism

IL-1R3¹⁴ is an accessory receptor of the IL-1 family, crucial for the constitution of the three high-affinity ternary complexes necessary for IL-1, IL-33 and IL-36 signal transmission.¹⁵ IL-1R3 blockade was investigated by Højen et al., by developing a humanized monoclonal antibody targeting IL-1R3 (MAB-hR3).¹⁵² In vitro, MAB-hR3 was able to inhibit signalling by IL-1R1, IL-1R4 and IL-1R6. Blocking IL-1R3 had a greater impact in lowering the production of proinflammatory cytokines compared with the inhibition of individual receptors in vitro (IL-1R1, IL-1R4 and IL-1R6).The effects of IL-1R3 blockade were also explored in vivo using a chimeric mouse monoclonal antibody (MAB-mR3) in murine models of inflammatory diseases driven by IL-1 β , IL-33 and IL-36. Treatment resulted in significant disease improvement, ¹⁵² suggesting that IL-1R3 blockade may have potential for the treatment of several types of inflammatory diseases in humans.

Conclusions and future directions

Thanks to recent advances in research, the complex pathophysiology of chronic inflammatory skin disorders, and their similarities, differences and driving pathways are being increasingly unravelled, thus identifying eligible targets for therapeutic intervention. Evidence that IL-1 family cytokines play a central role not only in rare monogenic AIDs, but also in some of the most common inflammatory skin diseases is rapidly growing. Further clinical and preclinical data, as well as further clinical trials, will hopefully lead to an extension of future indications and wider use of IL-1 family cytokine antagonists in daily clinical practice. The development of new drugs antagonizing IL-1 family pathways at multiple levels, such as via IL-1R3 antagonism, may have great potential, given that single-cytokine blockade in inflammatory diseases sometimes has limited efficacy.

References

- 1 Dinarello CA. Immunological and inflammatory functions of the interleukin-1 family. Annu Rev Immunol 2009; **27**:519–50.
- 2 Boraschi D, Italiani P, Weil S et al. The family of the interleukin-1 receptors. Immunol Rev 2018; **281**:197-232.
- 3 Fields JK, Günther S, Sundberg EJ. Structural basis of IL-1 family cytokine signaling. Front Immunol 2019; **10**:1412.
- 4 Dinarello CA. Overview of the IL-1 family in innate inflammation and acquired immunity. Immunol Rev 2018; 281:8–27.
- 5 van de Veerdonk FL, Stoeckman AK, Wu G et al. IL-38 binds to the IL-36 receptor and has biological effects on immune cells similar to IL-36 receptor antagonist. Proc Natl Acad Sci U S A 2012; 109:3001–5.
- 6 Zhou L, Todorovic V. Interleukin-36: structure, signaling and function. Adv Exp Med Biol 2021; 21:191-210.
- 7 Liew FY, Girard JP, Turnquist HR. Interleukin-33 in health and disease. Nat Rev Immunol 2016; 16:676-89.
- 8 Yasuda K, Nakanishi K, Tsutsui H. Interleukin-18 in health and disease. Int J Mol Sci 2019; 20:649.
- 9 Bufler P, Azam T, Gamboni-Robertson F et al. A complex of the IL-1 homologue IL-1F7b and IL-18-binding protein reduces IL-18 activity. Proc Natl Acad Sci U S A 2002; 99:13723-8.
- 10 Pan G, Risser P, Mao W et al. IL-1H, an interleukin 1-related protein that binds IL-18 receptor/IL-1Rrp. Cytokine 2001; 13:1–7.
- 11 Migliorini P, Italiani P, Pratesi F et al. The IL-1 family cytokines and receptors in autoimmune diseases. *Autoimmun Rev* 2020; 19:102617.
- 12 Novick D, Kim SH, Fantuzzi G et al. Interleukin-18 binding protein: a novel modulator of the Th1 cytokine response. Immunity 1999; 10:127–36.
- 13 Boraschi D, Tagliabue A. The interleukin-1 receptor family. Semin Immunol 2013; 25:394–407.
- 14 Greenfeder SA, Nunes P, Kwee L et al. Molecular cloning and characterization of a second subunit of the interleukin 1 receptor complex. J Biol Chem 1995; 270:13757–65.

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- 15 Wang D, Zhang S, Li L et al. Structural insights into the assembly and activation of IL-1 β with its receptors. Nat Immunol 2010; 11:905–11.
- 16 Bou-Dargham MJ, Khamis ZI, Cognetta AB et al. The role of interleukin-1 in inflammatory and malignant human skin diseases and the rationale for targeting interleukin-1 alpha. Med Res Rev 2017; 37:180–216.
- 17 Feldmeyer L, Werner S, French LE et al. Interleukin-1, inflammasomes and the skin. Eur J Cell Biol 2010; 89:638–44.
- 18 Di Paolo NC, Shayakhmetov DM. Interleukin 1α and the inflammatory process. Nat Immunol 2016; **17**:906–13.
- 19 Satoh T, Otsuka A, Contassot E et al. The inflammasome and IL- 1β : implications for the treatment of inflammatory diseases. Immunotherapy 2015; **7**:243–54.
- 20 England H, Summersgill HR, Edye ME et al. Release of interleukin-1 α or interleukin-1 β depends on mechanism of cell death. J Biol Chem 2014; **289**:15942–50.
- 21 Garlanda C, Dinarello CA, Mantovani A. The interleukin-1 family: back to the future. Immunity 2013; **39**:1003–18.
- 22 Satoh TK, Mellett M, Contassot E et al. Are neutrophilic dermatoses autoinflammatory disorders? Br J Dermatol 2018; **178**:603–13.
- 23 Manthiram K, Zhou Q, Aksentijevich I et al. The monogenic autoinflammatory diseases define new pathways in human innate immunity and inflammation. Nat Immunol 2017; 18:832–42.
- 24 Booshehri LM, Hoffman HM. CAPS and NLRP3. J Clin Immunol 2019; **39**:277-86.
- 25 Chae JJ, Cho YH, Lee GS et al. Gain-of-function pyrin mutations induce NLRP3 protein-independent interleukin-1 β activation and severe autoinflammation in mice. Immunity 2011; **34**:755–68.
- 26 Aksentijevich I, Masters SL, Ferguson PJ et al. An autoinflammatory disease with deficiency of the interleukin-1-receptor antagonist. N Engl J Med 2009; 360:2426–37.
- 27 Maverakis E, Ma C, Shinkai K et al. Diagnostic criteria of ulcerative pyoderma gangrenosum: a Delphi consensus of international experts. JAMA Dermatol 2018; **154**:461–6.
- 28 Nelson CA, Stephen S, Ashchyan HJ et al. Neutrophilic dermatoses: pathogenesis, Sweet syndrome, neutrophilic eccrine hidradenitis, and Behçet disease. J Am Acad Dermatol 2018; 79:987–1006.
- 29 Marzano AV, Ortega-Loayza AG, Heath M et al. Mechanisms of inflammation in neutrophil-mediated skin diseases. Front Immunol 2019; 10:1059.
- 30 Ikeda S, Saijo S, Murayama MA et al. Excess IL-1 signaling enhances the development of Th17 cells by downregulating TGF- β -induced Foxp3 expression. J Immunol 2014; **192**:1449–58.
- 31 Pelletier M, Maggi L, Micheletti A et al. Evidence for a cross-talk between human neutrophils and Th17 cells. Blood 2010; 115:335–43.
- 32 Watson RW, Rotstein OD, Parodo J et al. The IL-1 β -converting enzyme (caspase-1) inhibits apoptosis of inflammatory neutrophils through activation of IL-1 β . J Immunol 1998; **161**:957–62.
- 33 Mankan AK, Dau T, Jenne D et al. The NLRP3/ASC/caspase-1 axis regulates IL-1 β processing in neutrophils. Eur J Immunol 2012; **42**:710–15.
- 34 Imhof L, Meier B, Frei P et al. Severe Sweet's syndrome with elevated cutaneous interleukin-1 β after azathioprine exposure: case report and review of the literature. *Dermatology* 2015; **230**:293–8.
- 35 Kolios AG, Maul JT, Meier B et al. Canakinumab in adults with steroid-refractory pyoderma gangrenosum. Br J Dermatol 2015; 173:1216-23.
- 36 Düzgün N, Ayaşlioğlu E, Tutkak H et al. Cytokine inhibitors: soluble tumor necrosis factor receptor 1 and interleukin-1 receptor antagonist in Behçet's disease. Rheumatol Int 2005; 25:1–5.

- 37 Amazan E, Ezzedine K, Mossalayi MD et al. Expression of interleukin-1α in amicrobial pustulosis of the skin folds with complete response to anakinra. J Am Acad Dermatol 2014; 71:e53– 6.
- 38 Sabat R, Jemec GBE, Matusiak Ł et al. Hidradenitis suppurativa. Nat Rev Dis Primers 2020; 6:18.
- 39 Witte-Händel E, Wolk K, Tsaousi A et al. The IL-1 pathway is hyperactive in hidradenitis suppurativa and contributes to skin infiltration and destruction. J Invest Dermatol 2019; 139:1294–305.
- 40 van der Zee HH, de Ruiter L, van den Broecke DG et al. Elevated levels of tumour necrosis factor (TNF)- α , interleukin (IL)-1 β and IL-10 in hidradenitis suppurativa skin: a rationale for targeting TNF- α and IL-1 β . Br J Dermatol 2011; **164**:1292–8.
- 41 Kelly G, Hughes R, McGarry T et al. Dysregulated cytokine expression in lesional and nonlesional skin in hidradenitis suppurativa. Br J Dermatol 2015; 173:1431–9.
- 42 Michalek IM, Loring B, John SM. A systematic review of worldwide epidemiology of psoriasis. J Eur Acad Dermatol Venereol 2017; 31:205–12.
- 43 Boehncke WH, Schön MP. Psoriasis. Lancet 2015; 386:983-94.
- 44 Cai Y, Xue F, Quan C et al. A critical role of the IL-1β–IL-1R signaling pathway in skin inflammation and psoriasis pathogenesis. J Invest Dermatol 2019; 139:146–56.
- 45 Uribe-Herranz M, Lian LH, Hooper KM et al. IL-1R1 signaling facilitates Munro's microabscess formation in psoriasiform imiquimod-induced skin inflammation. J Invest Dermatol 2013; 133:1541–9.
- 46 Lowes MA, Suárez-Fariñas M, Krueger JG. Immunology of psoriasis. Annu Rev Immunol 2014; 32:227–55.
- 47 Cai Y, Xue F, Fleming C et al. Differential developmental requirement and peripheral regulation for dermal V γ 4 and V γ 6T17 cells in health and inflammation. Nat Commun 2014; **5**:3986.
- 48 Ghoreschi K, Laurence A, Yang XP et al. Generation of pathogenic $T_{\rm H}17$ cells in the absence of TGF- β signalling. Nature 2010; **467**:967–71.
- 49 Moussion C, Ortega N, Girard JP. The IL-1-like cytokine IL-33 is constitutively expressed in the nucleus of endothelial cells and epithelial cells in vivo: a novel 'alarmin'? PLOS ONE 2008; 3: e3331.
- 50 Ali S, Mohs A, Thomas M et al. The dual function cytokine IL-33 interacts with the transcription factor NF- κ B to dampen NF- κ B-stimulated gene transcription. J Immunol 2011; **187**:1609–16.
- 51 Lefrançais E, Duval A, Mirey E et al. Central domain of IL-33 is cleaved by mast cell proteases for potent activation of group-2 innate lymphoid cells. Proc Natl Acad Sci U S A 2014; 111:15502–7.
- 52 Martin NT, Martin MU. Interleukin 33 is a guardian of barriers and a local alarmin. Nat Immunol 2016; 17:122–31.
- 53 Balato A, Raimondo A, Balato N et al. Interleukin-33: increasing role in dermatological conditions. Arch Dermatol Res 2016; 308:287–96.
- 54 Brunner PM, Guttman-Yassky E, Leung DY. The immunology of atopic dermatitis and its reversibility with broad-spectrum and targeted therapies. J Allergy Clin Immunol 2017; 139 (4 Suppl.): S65–76.
- 55 Renert-Yuval Y, Guttman-Yassky E. What is new in atopic dermatitis. Dermatol Clin 2019; 37:205–13.
- 56 Imai Y, Yasuda K, Sakaguchi Y et al. Skin-specific expression of IL-33 activates group 2 innate lymphoid cells and elicits atopic dermatitis-like inflammation in mice. Proc Natl Acad Sci U S A 2013; 110:13921–6.
- 57 Tamagawa-Mineoka R, Okuzawa Y, Masuda K et al. Increased serum levels of interleukin 33 in patients with atopic dermatitis. J Am Acad Dermatol 2014; 70:882–8.

- 58 Savinko T, Matikainen S, Saarialho-Kere U et al. IL-33 and ST2 in atopic dermatitis: expression profiles and modulation by triggering factors. J Invest Dermatol 2012; 132:1392–400.
- 59 Stott B, Lavender P, Lehmann S et al. Human IL-31 is induced by IL-4 and promotes TH2-driven inflammation. J Allergy Clin Immunol 2013; 132:446–54.
- 60 Nygaard U, van den Bogaard EH, Niehues H et al. The 'alarmins' HMBG1 and IL-33 downregulate structural skin barrier proteins and impair epidermal growth. Acta Derm Venereol 2017; 97:305–12.
- 61 Imai Y. Interleukin-33 in atopic dermatitis. J Dermatol Sci 2019; 96:2–7.
- 62 Zeng F, Chen H, Chen L et al. An autocrine circuit of IL-33 in keratinocytes is involved in the progression of psoriasis. J Invest Dermatol 2021; 141:596–606.
- 63 Hueber AJ, Alves-Filho JC, Asquith DL et al. IL-33 induces skin inflammation with mast cell and neutrophil activation. Eur J Immunol 2011; 41:2229–37.
- 64 Balato A, Di Caprio R, Canta L et al. IL-33 is regulated by TNF-α in normal and psoriatic skin. Arch Dermatol Res 2014; 306:299–304.
- 65 Theoharides TC, Zhang B, Kempuraj D et al. IL-33 augments substance P-induced VEGF secretion from human mast cells and is increased in psoriatic skin. Proc Natl Acad Sci U S A 2010; 107:4448–53.
- 66 Balato A, Lembo S, Mattii M et al. IL-33 is secreted by psoriatic keratinocytes and induces pro-inflammatory cytokines via keratinocyte and mast cell activation. Exp Dermatol 2012; 21:892–4.
- 67 Mitsui A, Tada Y, Takahashi T et al. Serum IL-33 levels are increased in patients with psoriasis. Clin Exp Dermatol 2016; 41:183–9.
- 68 Cannavò SP, Bertino L, Di Salvo E et al. Possible roles of IL-33 in the innate-adaptive immune crosstalk of psoriasis pathogenesis. Mediators Inflamm 2019; 2019:7158014.
- 69 Gresnigt MS, van de Veerdonk FL. Biology of IL-36 cytokines and their role in disease. Semin Immunol 2013; 25:458–65.
- 70 Boutet MA, Bart G, Penhoat M et al. Distinct expression of interleukin (IL)-36 α , β and γ , their antagonist IL-36Ra and IL-38 in psoriasis, rheumatoid arthritis and Crohn's disease. Clin Exp Immunol 2016; **184**:159–73.
- 71 Dyring-Andersen B, Løvendorf MB, Coscia F et al. Spatially and cell-type resolved quantitative proteomic atlas of healthy human skin. Nat Commun 2020; 11:5587.
- 72 Foster AM, Baliwag J, Chen CS et al. IL-36 promotes myeloid cell infiltration, activation, and inflammatory activity in skin. J Immunol 2014; 192:6053–61.
- 73 Buhl AL, Wenzel J. Interleukin-36 in infectious and inflammatory skin diseases. Front Immunol 2019; 10:1162.
- 74 Marrakchi S, Guigue P, Renshaw BR et al. Interleukin-36-receptor antagonist deficiency and generalized pustular psoriasis. N Engl J Med 2011; 365:620–8.
- 75 Körber A, Mössner R, Renner R et al. Mutations in IL36RN in patients with generalized pustular psoriasis. J Invest Dermatol 2013; 133:2634–7.
- 76 Onoufriadis A, Simpson MA, Pink AE et al. Mutations in IL36RN/ IL1F5 are associated with the severe episodic inflammatory skin disease known as generalized pustular psoriasis. Am J Hum Genet 2011; 89:432–7.
- 77 Sugiura K, Takemoto A, Yamaguchi M et al. The majority of generalized pustular psoriasis without psoriasis vulgaris is caused by deficiency of interleukin-36 receptor antagonist. J Invest Dermatol 2013; 133:2514–21.
- 78 Takahashi T, Fujimoto N, Kabuto M et al. Mutation analysis of IL36RN gene in Japanese patients with palmoplantar pustulosis. J Dermatol 2017; 44:80–3.

- 79 Bissonnette R, Suárez-Fariñas M, Li X et al. Based on molecular profiling of gene expression, palmoplantar pustulosis and palmoplantar pustular psoriasis are highly related diseases that appear to be distinct from psoriasis vulgaris. PLOS ONE 2016; 11:e0155215.
- 80 Jordan CT, Cao L, Roberson ED et al. PSORS2 is due to mutations in CARD14. Am J Hum Genet 2012; 90:784–95.
- 81 Bertin J, Wang L, Guo Y et al. CARD11 and CARD14 are novel caspase recruitment domain (CARD)/membrane-associated guanylate kinase (MAGUK) family members that interact with BCL10 and activate NF-κB. J Biol Chem 2001; 276:11877–82.
- 82 Madonna S, Girolomoni G, Dinarello CA et al. The significance of IL-36 hyperactivation and IL-36R targeting in psoriasis. Int J Mol Sci 2019; 20:3318.
- 83 Albanesi C, Madonna S, Gisondi P et al. The interplay between keratinocytes and immune cells in the pathogenesis of psoriasis. Front Immunol 2018; 9:1549.
- 84 Boutet MA, Nerviani A, Pitzalis C. IL-36, IL-37, and IL-38 cytokines in skin and joint inflammation: a comprehensive review of their therapeutic potential. Int J Mol Sci 2019; 20:1257.
- 85 Mercurio L, Morelli M, Scarponi C et al. IL-38 has an antiinflammatory action in psoriasis and its expression correlates with disease severity and therapeutic response to anti-IL-17A treatment. Cell Death Dis 2018; **9**:1104.
- 86 D'Erme AM, Wilsmann-Theis D, Wagenpfeil J et al. IL-36γ (IL-1F9) is a biomarker for psoriasis skin lesions. J Invest Dermatol 2015; 135:1025–32.
- 87 Giannoudaki E, Stefanska AM, Lawler H et al. SIGIRR negatively regulates IL-36-driven psoriasiform inflammation and neutrophil infiltration in the skin. J Immunol 2021; 207:651–60.
- 88 Henry CM, Sullivan GP, Clancy DM et al. Neutrophil-derived proteases escalate inflammation through activation of IL-36 family cytokines. Cell Rep 2016; 14:708–22.
- 89 Sidoroff A, Halevy S, Bavinck JN et al. Acute generalized exanthematous pustulosis (AGEP) – a clinical reaction pattern. J Cutan Pathol 2001; 28:113–19.
- 90 Feldmeyer L, Heidemeyer K, Yawalkar N. Acute generalized exanthematous pustulosis: pathogenesis, genetic background, clinical variants and therapy. Int J Mol Sci 2016; 17:1214.
- 91 Meier-Schiesser B, Feldmeyer L, Jankovic D et al. Culprit drugs induce specific IL-36 overexpression in acute generalized exanthematous pustulosis. J Invest Dermatol 2019; 139:848–58.
- 92 Satoh TK, Mellett M, Meier-Schiesser B et al. IL-36 γ drives skin toxicity induced by EGFR/MEK inhibition and commensal Cutibacterium acnes. J Clin Invest 2020; **130**:1417–30.
- 93 Hessam S, Sand M, Gambichler T et al. Interleukin-36 in hidradenitis suppurativa: evidence for a distinctive proinflammatory role and a key factor in the development of an inflammatory loop. Br J Dermatol 2018; 178:761–7.
- 94 Thomi R, Kakeda M, Yawalkar N et al. Increased expression of the interleukin-36 cytokines in lesions of hidradenitis suppurativa. J Eur Acad Dermatol Venereol 2017; 31:2091–6.
- 95 Di Caprio R, Balato A, Caiazzo G et al. IL-36 cytokines are increased in acne and hidradenitis suppurativa. Arch Dermatol Res 2017; **309**:673-8.
- 96 Wolk K, Brembach TC, Šimaitė D et al. Activity and components of the granulocyte colony-stimulating factor pathway in hidradenitis suppurativa. Br J Dermatol 2021; 185:164–76.
- 97 Mühl H, Pfeilschifter J. Interleukin-18 bioactivity: a novel target for immunopharmacological anti-inflammatory intervention. Eur J Pharmacol 2004; 500:63–71.
- 98 Roth W, Kumar V, Beer HD et al. Keratin 1 maintains skin integrity and participates in an inflammatory network in skin through interleukin-18. J Cell Sci 2012; 125:5269–79.

- 99 Seco T, Cerqueira A, Costa A et al. Adult-onset Still's disease: typical presentation, delayed diagnosis. Cureus 2020; **12**:e8510.
- 100 Feist E, Mitrovic S, Fautrel B. Mechanisms, biomarkers and targets for adult-onset Still's disease. Nat Rev Rheumatol 2018; 14:603–18.
- 101 Jamilloux Y, Gerfaud-Valentin M, Martinon F et al. Pathogenesis of adult-onset Still's disease: new insights from the juvenile counterpart. Immunol Res 2015; 61:53–62.
- 102 Kawashima M, Yamamura M, Taniai M et al. Levels of interleukin-18 and its binding inhibitors in the blood circulation of patients with adult-onset Still's disease. Arthritis Rheum 2001; 44:550–60.
- 103 Chen DY, Lan JL, Lin FJ et al. Proinflammatory cytokine profiles in sera and pathological tissues of patients with active untreated adult onset Still's disease. J Rheumatol 2004; 31:2189–98.
- 104 Girard C, Rech J, Brown M et al. Elevated serum levels of free interleukin-18 in adult-onset Still's disease. Rheumatology (Oxford) 2016; 55:2237–47.
- 105 Nam SW, Kang S, Lee JH et al. Different features of interleukin-37 and interleukin-18 as disase activity markers of adult-onset Still's disease. J Clin Med 2021; 10:910.
- 106 Chen S, Jiang F, Ren J et al. Association of IL-18 polymorphisms with rheumatoid arthritis and systemic lupus erythematosus in Asian populations: a meta-analysis. BMC Med Genet 2012; 13:107.
- 107 Wang D, Drenker M, Eiz-Vesper B et al. Evidence for a pathogenetic role of interleukin-18 in cutaneous lupus erythematosus. Arthritis Rheum 2008; 58:3205–15.
- 108 Xiang M, Feng Y, Wang Y et al. Correlation between circulating interleukin-18 level and systemic lupus erythematosus: a metaanalysis. Sci Rep 2021; 11:4707.
- 109 Aghdashi M, Aribi S, Salami S. Serum levels of IL-18 in Iranian females with systemic lupus erythematosus. *Med Arch* 2013; 67:237–40.
- 110 Mikita N, Ikeda T, Ishiguro M et al. Recent advances in cytokines in cutaneous and systemic lupus erythematosus. J Dermatol 2011; 38:839–49.
- 111 Ciążyńska M, Olejniczak-Staruch I, Sobolewska-Sztychny D et al. The role of NLRP1, NLRP3, and AIM2 inflammasomes in psoriasis: review. Int J Mol Sci 2021; 22:5898.
- 112 Companjen A, van der Wel L, van der Fits L et al. Elevated interleukin-18 protein expression in early active and progressive plaque-type psoriatic lesions. Eur Cytokine Netw 2004; 15:210–16.
- 113 Forouzandeh M, Besen J, Keane RW et al. The inflammasome signaling proteins ASC and IL-18 as biomarkers of psoriasis. Front Pharmacol 2020; 11:1238.
- 114 US Food and Drug Administration Kineret [prescribing information]. Available at: https://www.accessdata.fda.gov/drugsatfda_ docs/label/2012/103950s5136lbl.pdf (last accessed 31 January 2022).
- 115 European Medicines Agency. Kineret [prescribing information]. Available at: https://www.ema.europa.eu/en/documents/productinformation/kineret-epar-product-information_en.pdf (last accessed 31 January 2022).
- 116 US Food and Drug Administration. Arcalyst [prescribing information]. Available at: https://www.accessdata.fda.gov/drugsatfda_ docs/label/2008/125249lbl.pdf (last accessed 31 January 2022).
- 117 US Food and Drug Administration Ilaris [prescribing information]. Available at: https://www.accessdata.fda.gov/drugsatfda_ docs/label/2016/BLA125319_858687lbl.pdf (last accessed 31 January 2022).
- 118 European Medicines Agency. Ilaris [prescribing information]. Available at: https://www.ema.europa.eu/en/documents/product-information/ilaris-epar-product-information_en.pdf (last accessed 31 January 2022).

- 119 de Koning HD, Bodar EJ, Simon A et al. Beneficial response to anakinra and thalidomide in Schnitzler's syndrome. Ann Rheum Dis 2006; 65:542–4.
- 120 Martinez-Taboada VM, Fontalba A, Blanco R et al. Successful treatment of refractory Schnitzler syndrome with anakinra: comment on the article by Hawkins et al. Arthritis Rheum 2005; 52:2226–7.
- 121 Krause K, Weller K, Stefaniak R et al. Efficacy and safety of the interleukin-1 antagonist rilonacept in Schnitzler syndrome: an open-label study. Allergy 2012; 67:943–50.
- 122 Fabiani C, Vitale A, Emmi G et al. Interleukin (IL)-1 inhibition with anakinra and canakinumab in Behçet's disease-related uveitis: a multicenter retrospective observational study. Clin Rheumatol 2017; 36:191–7.
- 123 Feist E, Quartier P, Fautrel B et al. Efficacy and safety of canakinumab in patients with Still's disease: exposure–response analysis of pooled systemic juvenile idiopathic arthritis data by age groups. Clin Exp Rheumatol 2018; 36:668–75.
- 124 Naumann L, Feist E, Natusch A et al. IL1-receptor antagonist anakinra provides long-lasting efficacy in the treatment of refractory adult-onset Still's disease. Ann Rheum Dis 2010; **69**:466–7.
- 125 Lequerré T, Quartier P, Rosellini D et al. Interleukin-1 receptor antagonist (anakinra) treatment in patients with systemic-onset juvenile idiopathic arthritis or adult onset Still disease: preliminary experience in France. Ann Rheum Dis 2008; 67:302–8.
- 126 Delluc A, Limal N, Puéchal X et al. Efficacy of anakinra, an IL1 receptor antagonist, in refractory Sweet syndrome. Ann Rheum Dis 2008; 67:278–9.
- 127 Quist SR, Kraas L. Treatment options for pyoderma gangrenosum. J Dtsch Dermatol Ges 2017; 15:34–40.
- 128 Behrens EM, Kreiger PA, Cherian S et al. Interleukin 1 receptor antagonist to treat cytophagic histiocytic panniculitis with secondary hemophagocytic lymphohistiocytosis. J Rheumatol 2006; 33:2081–4.
- 129 Cugno M, Borghi A, Marzano AV. PAPA, PASH and PAPASH syndromes: pathophysiology, presentation and treatment. Am J Clin Dermatol 2017; 18:555–62.
- 130 Wendling D, Prati C, Aubin F. Anakinra treatment of SAPHO syndrome: short-term results of an open study. Ann Rheum Dis 2012; 71:1098–100.
- 131 Stojanov S, Lapidus S, Chitkara P et al. Periodic fever, aphthous stomatitis, pharyngitis, and adenitis (PFAPA) is a disorder of innate immunity and Th1 activation responsive to IL-1 blockade. Proc Natl Acad Sci U S A 2011; 108:7148–53.
- 132 Rossi-Semerano L, Piram M, Chiaverini C et al. First clinical description of an infant with interleukin-36-receptor antagonist deficiency successfully treated with anakinra. Pediatrics 2013; 132: e1043–7.
- 133 Viguier M, Guigue P, Pagès C et al. Successful treatment of generalized pustular psoriasis with the interleukin-1-receptor antagonist Anakinra: lack of correlation with IL1RN mutations. Ann Intern Med 2010; 153:66–7.
- 134 Skendros P, Papagoras C, Lefaki I et al. Successful response in a case of severe pustular psoriasis after interleukin-1 β inhibition. Br J Dermatol 2017; **176**:212–15.
- 135 Tauber M, Viguier M, Alimova E et al. Partial clinical response to anakinra in severe palmoplantar pustular psoriasis. Br J Dermatol 2014; 171:646–9.
- 136 Leslie KS, Tripathi SV, Nguyen TV et al. An open-label study of anakinra for the treatment of moderate to severe hidradenitis suppurativa. J Am Acad Dermatol 2014; 70:243–51.
- 137 Tzanetakou V, Kanni T, Giatrakou S et al. Safety and efficacy of anakinra in severe hidradenitis suppurativa: a randomized clinical trial. JAMA Dermatol 2016; **152**:52–9.

- 138 Sun NZ, Ro T, Jolly P et al. Non-response to interleukin-1 antagonist canakinumab in two patients with refractory pyoderma gangrenosum and hidradenitis suppurativa. J Clin Aesthet Dermatol 2017; **10**:36–8.
- 139 Houriet C, Seyed Jafari SM, Thomi R et al. Canakinumab for severe hidradenitis suppurativa: preliminary experience in 2 cases. JAMA Dermatol 2017; 153:1195–7.
- 140 Kanni T, Argyropoulou M, Spyridopoulos T et al. MABp1 targeting IL-1 α for moderate to severe hidradenitis suppurativa not eligible for adalimumab: a randomized study. J Invest Dermatol 2018; 138:795–801.
- 141 Gottlieb A, Natsis NE, Kerdel F et al. A phase II open-label study of bermekimab in patients with hidradenitis suppurativa shows resolution of inflammatory lesions and pain. J Invest Dermatol 2020; **140**:1538–45.
- 142 Carrasco D, Stecher M, Lefebvre GC et al. An open label, phase 2 study of MABp1 monotherapy for the treatment of acne vulgaris and psychiatric comorbidity. J Drugs Dermatol 2015; 14:560–4.
- 143 Coleman KM, Gudjonsson JE, Stecher M. Open-label trial of MABp1, a true human monoclonal antibody targeting interleukin 1α , for the treatment of psoriasis. JAMA Dermatol 2015; **151**:555–6.
- 144 Ganesan R, Raymond EL, Mennerich D et al. Generation and functional characterization of anti-human and anti-mouse IL-36R antagonist monoclonal antibodies. MAbs 2017; **9**:1143–54.
- 145 Bachelez H, Choon SE, Marrakchi S et al. Inhibition of the interleukin-36 pathway for the treatment of generalized pustular psoriasis. N Engl J Med 2019; 380:981–3.
- 146 Misiak-Galazka M, Zozula J, Rudnicka L. Palmoplantar pustulosis: recent advances in etiopathogenesis and emerging treatments. Am J Clin Dermatol 2020; 21:355–70.
- 147 Mrowietz U, Burden AD, Pinter A et al. Spesolimab, an antiinterleukin-36 receptor antibody, in patients with palmoplantar pustulosis: results of a phase IIa, multicentre, double-blind, randomized, placebo-controlled pilot study. Dermatol Ther (Heidelb) 2021; 11:571–85.
- 148 Khanskaya I, Pinkstaff J, Marino MH et al. A phase 1 study of ANB019, an anti-IL-36 receptor monoclonal antibody, in healthy volunteers. Available at: https://www2.anaptysbio.com/wpcontent/uploads/ANB019-Phase-1-Study-Poster-EAACI-2018.pdf (last accessed 31 January 2022).
- 149 Chen Y-L, Gutowska-Owsiak D, Hardman CS et al. Proof-ofconcept clinical trial of etokimab shows a key role for IL-33 in atopic dermatitis pathogenesis. Sci Transl Med 2019; 11:eaax2945.
- 150 Nnane I, Frederick B, Yao Z et al. The first-in-human study of CNTO 7160, an anti-interleukin-33 receptor monoclonal antibody, in healthy subjects and patients with asthma or atopic dermatitis. Br J Clin Pharmacol 2020; 86:2507–18.
- 151 Gabay C, Fautrel B, Rech J et al. Open-label, multicentre, doseescalating phase II clinical trial on the safety and efficacy of tadekinig alfa (IL-18BP) in adult-onset Still's disease. Ann Rheum Dis 2018; 77:840–7.
- 152 Højen JF, Kristensen MLV, McKee AS et al. IL-1R3 blockade broadly attenuates the functions of six members of the IL-1 family, revealing their contribution to models of disease. Nat Immunol 2019; 20:1138–49.
- 153 Nordström D, Knight A, Luukkainen R et al. Beneficial effect of interleukin 1 inhibition with anakinra in adult-onset Still's disease. An open, randomized, multicenter study. J Rheumatol 2012; 39:2008–11.
- 154 Kedor C, Listing J, Zernicke J et al. Canakinumab for treatment of adult-onset Still's disease to achieve reduction of arthritic manifestation (CONSIDER): phase II, randomised, double-blind,

placebo-controlled, multicentre, investigator-initiated trial. Ann Rheum Dis 2020; **79**:1090-7.

- 155 Shahid Z, Kalayanamitra R, Patel R et al. Refractory Sweet syndrome treated with anakinra. Cureus 2019; 11:e4536.
- 156 Kluger N, Gil-Bistes D, Guillot B et al. Efficacy of anti-interleukin-1 receptor antagonist anakinra (Kineret®) in a case of refractory Sweet's syndrome. Dermatology 2011; 222:123–7.
- 157 Grayson PC, Yazici Y, Merideth M et al. Treatment of mucocutaneous manifestations in Behçet's disease with anakinra: a pilot open-label study. Arthritis Res Ther 2017; 19:69.
- 158 Tugal-Tutkun I, Pavesio C, De Cordoue A et al. Use of gevokizumab in patients with Behçet's disease uveitis: an international, randomized, double-masked, placebo-controlled study and openlabel extension study. Ocul Immunol Inflamm 2018; 26:1023–33.
- 159 Tugal-Tutkun IM, Kadayifcilar SM, Khairallah MM et al. Safety and efficacy of gevokizumab in patients with Behçet's disease uveitis: results of an exploratory phase 2 study. Ocul Immunol Inflamm 2017; 25:62–70.
- 160 Gül A, Tugal-Tutkun I, Dinarello CA et al. Interleukin-1 β -regulating antibody XOMA 052 (gevokizumab) in the treatment of acute exacerbations of resistant uveitis of Behcet's disease: an open-label pilot study. Ann Rheum Dis 2012; **71**:563–6.
- 161 Jennings L, Molloy O, Quinlan C et al. Treatment of pyoderma gangrenosum, acne, suppurative hidradenitis (PASH) with weight-based anakinra dosing in a hepatitis B carrier. Int J Dermatol 2017; 56:e128–9.
- 162 Casas C, Paul C, Lahfa M et al. Quantification of Demodex folliculorum by PCR in rosacea and its relationship to skin innate immune activation. Exp Dermatol 2012; 21:906–10.
- 163 Soylu A, Yıldız G, Torun Bayram M et al. IL-1β blockade in periodic fever, aphthous stomatitis, pharyngitis, and cervical adenitis (PFAPA) syndrome: case-based review. Rheumatol Int 2021; 41:183–8.
- 164 Abramovits W, Rivas Bejarano JJ, Valdecantos WC. Role of interleukin 1 in atopic dermatitis. Dermatol Clin 2013; 31:437–44.
- 165 Yamanaka K, Mizutani H. 'Inflammatory skin march': IL-1mediated skin inflammation, atopic dermatitis, and psoriasis to cardiovascular events. J Allergy Clin Immunol 2015; 136:823–4.
- 166 Mattii M, Ayala F, Balato N et al. The balance between pro- and anti-inflammatory cytokines is crucial in human allergic contact dermatitis pathogenesis: the role of IL-1 family members. Exp Dermatol 2013; 22:813–19.
- 167 Watanabe H, Gaide O, Pétrilli V et al. Activation of the IL-1 β -processing inflammasome is involved in contact hypersensitivity. J Invest Dermatol 2007; **127**:1956–63.
- 168 Groh M, Rogowska K, Monsarrat O et al. Interleukin-1 receptor antagonist for refractory anti-MDA5 clinically amyopathic dermatomyopathy. Clin Exp Rheumatol 2015; 33:904–5.
- 169 Landeck L, Visser M, Kezic S et al. ILIA-889 C/T gene polymorphism in irritant contact dermatitis. J Eur Acad Dermatol Venereol 2013; 27:1040–3.
- 170 Spiekstra SW, Toebak MJ, Sampat-Sardjoepersad S et al. Induction of cytokine (interleukin-1α and tumor necrosis factor-α) and chemokine (CCL20, CCL27, and CXCL8) alarm signals after allergen and irritant exposure. Exp Dermatol 2005; 14:109–16.
- 171 Conti P, Lauritano D, Caraffa A et al. New insight into systemic mastocytosis mediated by cytokines IL-1 β and IL-33: potential inhibitory effect of IL-37. Eur J Pharmacol 2019; **858**:172473.
- 172 Rowczenio DM, Pathak S, Arostegui JI et al. Molecular genetic investigation, clinical features, and response to treatment in 21 patients with Schnitzler syndrome. Blood 2018; 131:974–81.
- 173 Tran T-A, Pariente D, Guitton C et al. Treatment of Erdheim–Chester disease with canakinumab. Rheumatology 2014; **53**:2312–14.

- 174 Darstein F, Kirschey S, Heckl S et al. Successful treatment of Erdheim–Chester disease with combination of interleukin-1targeting drugs and high-dose glucocorticoids. Intern Med J 2014; 44:90–2.
- 175 Courcoul A, Vignot E, Chapurlat R. Successful treatment of Erdheim–Chester disease by interleukin-1 receptor antagonist protein. Joint Bone Spine 2014; 81:175–7.
- 176 Hussein MR, Hassan HI, Hofny ER et al. Alterations of mononuclear inflammatory cells, CD4/CD8⁺ T cells, interleukin 1 β , and tumour necrosis factor α in the bronchoalveolar lavage fluid, peripheral blood, and skin of patients with systemic sclerosis. J Clin Pathol 2005; **58**:178–84.
- 177 Martínez-Godínez MA, Cruz-Domínguez MP, Jara LJ et al. Expression of NLRP3 inflammasome, cytokines and vascular mediators in the skin of systemic sclerosis patients. Isr Med Assoc J 2015; 17:5–10.
- 178 Krause K, Mahamed A, Weller K et al. Efficacy and safety of canakinumab in urticarial vasculitis: an open-label study. J Allergy Clin Immunol 2013; 132:751–4.
- 179 Kisla Ekinci RM, Balci S, Bisgin A et al. Renal amyloidosis in deficiency of adenosine deaminase 2: successful experience with canakinumab. *Pediatrics* 2018; **142**:e20180948.
- 180 Brzoza Z, Rymarczyk B, Grzeszczak W et al. Interleukin 1 gene polymorphisms presumably participate in the pathogenesis of chronic spontaneous autoreactive urticaria. J Interferon Cytokine Res 2020; 40:497–500.
- 181 Ben-Zvi I, Kukuy O, Giat E et al. Anakinra for colchicine-resistant familial Mediterranean fever: a randomized, double-blind, placebo-controlled trial. Arthritis Rheumatol 2017; 69:854–62.
- 182 Herlin T, Fiirgaard B, Bjerre M et al. Efficacy of anti-IL-1 treatment in Majeed syndrome. Ann Rheum Dis 2013; 72:410–13.
- 183 Schmidt E, Mittnacht A, Schömig H et al. Detection of IL-1α, IL-1β and IL-1 receptor antagonist in blister fluid of bullous pemphigoid. J Dermatol Sci 1996; 11:142–7.
- 184 Brenner M, Ruzicka T, Plewig G et al. Targeted treatment of pyoderma gangrenosum in PAPA (pyogenic arthritis, pyoderma gangrenosum and acne) syndrome with the recombinant human interleukin-1 receptor antagonist anakinra. Br J Dermatol 2009; 161:1199–201.
- 185 Bhardwaj S, Rani S, Srivastava N et al. Increased systemic and epidermal levels of IL-17A and IL-1 β promotes progression of nonsegmental vitiligo. Cytokine 2017; **91**:153–61.
- 186 Lachmann HJ, Kone-Paut I, Kuemmerle-Deschner JB et al. Use of canakinumab in the cryopyrin-associated periodic syndrome. N Engl J Med 2009; 360:2416–25.
- 187 Hautala T, Vähäsalo P, Kuismin O et al. A family with A20 haploinsufficiency presenting with novel clinical manifestations and challenges for treatment. J Clin Rheumatol 2021; 27:e583–7.
- 188 De Benedetti F, Gattorno M, Anton J et al. Canakinumab for the treatment of autoinflammatory recurrent fever syndromes. N Engl J Med 2018; 378:1908–19.
- 189 Moghaddas F, Llamas R, De Nardo D et al. A novel pyrinassociated autoinflammation with neutrophilic dermatosis mutation further defines 14-3-3 binding of pyrin and distinction to familial Mediterranean fever. Ann Rheum Dis 2017; 76:2085–94.
- 190 Greally P, Hussain MJ, Price JF et al. Interleukin-1α and soluble interleukin-2 receptor in atopic dermatitis. Arch Dis Child 1992; 67:1413.
- 191 Suhng E, Kim BH, Choi YW et al. Increased expression of IL-33 in rosacea skin and UVB-irradiated and LL-37-treated HaCaT cells. Exp Dermatol 2018; 27:1023–9.
- 192 Meephansan J, Komine M, Tsuda H et al. Expression of IL-33 in the epidermis: the mechanism of induction by IL-17. J Dermatol Sci 2013; 71:107–14.

- 193 Conti P, Pregliasco FE, Bellomo RG et al. Mast cell cytokines IL-1, IL-33, and IL-36 mediate skin inflammation in psoriasis: a novel therapeutic approach with the anti-inflammatory cytokines IL-37, IL-38, and IL-1Ra. Int J Mol Sci 2021; 22:8076.
- 194 Mok MY, Huang FP, Ip WK et al. Serum levels of IL-33 and soluble ST2 and their association with disease activity in systemic lupus erythematosus. Rheumatology 2009; 49:520–7.
- 195 Yanaba K, Yoshizaki A, Asano Y et al. Serum IL-33 levels are raised in patients with systemic sclerosis: association with extent of skin sclerosis and severity of pulmonary fibrosis. Clin Rheumatol 2011; 30:825–30.
- 196 Manetti M, Ibba-Manneschi L, Liakouli V et al. The IL1-like cytokine IL33 and its receptor ST2 are abnormally expressed in the affected skin and visceral organs of patients with systemic sclerosis. Ann Rheum Dis 2010; 69:598–605.
- 197 Kay AB, Clark P, Maurer M et al. Elevations in T-helper-2initiating cytokines (interleukin-33, interleukin-25 and thymic stromal lymphopoietin) in lesional skin from chronic spontaneous ('idiopathic') urticaria. Br J Dermatol 2015; **172**:1294–302.
- 198 Wakatabi K, Komine M, Meephansan J et al. The levels of soluble ST2 in sera and bullous fluid from patients with bullous pemphigoid. Eur J Dermatol 2012; 22:333–6.
- 199 Kim DJ, Baek SY, Park MK et al. Serum level of interleukin-33 and soluble ST2 and their association with disease activity in patients with Behcet's disease. J Korean Med Sci 2013; 28:1145–53.
- 200 Li P, Ma H, Han D et al. Interleukin-33 affects cytokine production by keratinocytes in vitiligo. Clin Exp Dermatol 2015; 40:163– 70.
- 201 Chu M, Wong CK, Cai Z et al. Elevated expression and proinflammatory activity of IL-36 in patients with systemic lupus erythematosus. Molecules 2015; 20:19588–604.
- 202 Xu D, Mu R, Wei X. The roles of IL-1 family cytokines in the pathogenesis of systemic sclerosis. Front Immunol 2019; **10**:2025.
- 203 Żebrowska A, Woźniacka A, Juczyńska K et al. Correlation between IL36α and IL17 and activity of the disease in selected autoimmune blistering diseases. Mediators Inflamm 2017; 2017:8980534.
- 204 Suárez-Fariñas M, Ungar B, Correa da Rosa J et al. RNA sequencing atopic dermatitis transcriptome profiling provides insights into novel disease mechanisms with potential therapeutic implications. J Allergy Clin Immunol 2015; 135:1218–27.
- 205 Sato S, Chiba T, Nakahara T et al. Upregulation of IL-36 cytokines in folliculitis and eosinophilic pustular folliculitis. Australas J Dermatol 2020; 61:e39–45.
- 206 Maczynska I, Millo B, Ratajczak-Stefańska V et al. Proinflammatory cytokine (IL-1 β , IL-6, IL-12, IL-18 and TNF- α) levels in sera of patients with subacute cutaneous lupus erythematosus (SCLE). Immunol Lett 2006; **102**:79–82.
- 207 Gangemi S, Merendino RA, Guarneri F et al. Serum levels of interleukin-18 and s-ICAM-1 in patients affected by psoriasis: preliminary considerations. J Eur Acad Dermatol Venereol 2003; 17:42–6.
- 208 Johansen C, Moeller K, Kragballe K et al. The activity of caspase-1 is increased in lesional psoriatic epidermis. J Invest Dermatol 2007; 127:2857–64.
- 209 Konishi H, Tsutsui H, Murakami T et al. IL-18 contributes to the spontaneous development of atopic dermatitis-like inflammatory skin lesion independently of IgE/stat6 under specific pathogenfree conditions. Proc Natl Acad Sci U S A 2002; 99:11340–5.
- 210 Zedan K, Rasheed Z, Farouk Y et al. Immunoglobulin e, interleukin-18 and interleukin-12 in patients with atopic dermatitis: correlation with disease activity. J Clin Diagn Res 2015; 9: WC01-5.

- 211 Puxeddu I, Italiani P, Giungato P et al. Free IL-18 and IL-33 cytokines in chronic spontaneous urticaria. Cytokine 2013; **61**:741–3.
- 212 Galbiati V, Papale A, Galli CL et al. Role of ROS and HMGB1 in contact allergen-induced IL-18 production in human keratinocytes. J Invest Dermatol 2014; 134:2719–27.
- 213 Lee D, Hong SK, Park SW et al. Serum levels of IL-18 and sIL-2R in patients with alopecia areata receiving combined therapy with oral cyclosporine and steroids. Exp Dermatol 2010; 19:145–7.
- 214 Park HJ, Kim HS, Kim HJ et al. Immunohistochemical characterization of cutaneous drug eruptions by STI571. J Dermatol Sci 2005; **38**:9–15.
- 215 Fujimori Y, Takatsuka H, Takemoto Y et al. Elevated interleukin (IL)-18 levels during acute graft-versus-host disease after allogeneic bone marrow transplantation. Br J Haematol 2000; 109:652–7.
- 216 Park HJ, Kim JE, Lee JY et al. Increased expression of IL-18 in cutaneous graft-versus-host disease. Immunol Lett 2004; 95:57–61.
- 217 Brydges SD, Mueller JL, McGeough MD et al. Inflammasomemediated disease animal models reveal roles for innate but not adaptive immunity. Immunity 2009; 30:875–87.
- 218 Novick D, Elbirt D, Dinarello CA et al. Interleukin-18 binding protein in the sera of patients with Wegener's granulomatosis. J Clin Immunol 2009; 29:38–45.
- 219 Scala E, Pallotta S, Frezzolini A et al. Cytokine and chemokine levels in systemic sclerosis: relationship with cutaneous and internal organ involvement. Clin Exp Immunol 2004; 138:540–6.
- 220 Kanameishi S, Nakamizo S, Endo Y et al. High level of serum human interleukin-18 in a patient with pyogenic arthritis, pyoderma gangrenosum and acne syndrome. J Eur Acad Dermatol Venereol 2017; 31:e115–16.
- 221 Wada T, Toma T, Miyazawa H et al. Longitudinal analysis of serum interleukin-18 in patients with familial Mediterranean

fever carrying MEFV mutations in exon 10. Cytokine 2018; **104**:143–6.

- 222 Haznedaroglu S, Oztürk MA, Sancak B et al. Serum interleukin 17 and interleukin 18 levels in familial Mediterranean fever. Clin Exp Rheumatol 2005; 23 (4 Suppl. 38):S77–80.
- 223 Kim M, Kim KE, Jung HY et al. Recombinant erythroid differentiation regulator 1 inhibits both inflammation and angiogenesis in a mouse model of rosacea. Exp Dermatol 2015; 24:680–5.
- 224 Przepiera-Będzak H, Fischer K, Brzosko M. Serum interleukin-18, fetuin-a, soluble intercellular adhesion molecule-1, and endothelin-1 in ankylosing spondylitis, psoriatic arthritis, and SAPHO syndrome. Int J Mol Sci 2016; **17**:1255.
- 225 Fang H, Shao S, Cao T et al. Increased expression of NLRP3 inflammasome components and interleukin-18 in patients with bullous pemphigoid. J Dermatol Sci 2016; 83:116–23.
- 226 Tirado SA, Ponce ORM, Montes de Oca SG. [Interleukin 18 as a marker of immunological activity in pemphigus vulgaris]. Dermatol Rev Mex 2009; 53:215–18 (in Spanish).
- 227 Prasinou M, Smith R, Vrettos A et al. The role of IL-18 in Behcet's disease; a potential therapeutic target. Autoimmun Rev 2020; 19:102613.
- 228 Migliorini P, Italiani P, Pratesi F et al. Cytokines and soluble receptors of the interleukin-1 family in Schnitzler syndrome. Scand J Rheumatol 2019; 48:235–8.
- 229 Hashkes PJ, Spalding SJ, Giannini EH et al. Rilonacept for colchicine-resistant or -intolerant familial Mediterranean fever: a randomized trial. Ann Intern Med 2012; 157:533–41.
- 230 Mantero JC, Kishore N, Ziemek J et al. Randomised, doubleblind, placebo-controlled trial of IL1-trap, rilonacept, in systemic sclerosis. A phase I/II biomarker trial. Clin Exp Rheumatol 2018; 36 (Suppl. 113):146–9.