



## HIGHLIGHTS

Claudia C. Beerweiler et al.

### **REVIEW**

# Asthma and allergic diseases: Cross talk of immune system and environmental factors

Claudia C. Beerweiler<sup>1,3</sup> , Rebecca K. Masanetz<sup>2</sup> and Bianca Schaub<sup>1,3</sup>

- <sup>1</sup> Pediatric Allergology, Department of Pediatrics, Dr. von Hauner Children's Hospital, University Hospital, LMU Munich, Munich, Germany
- <sup>2</sup> Department of Molecular Neurology, Friedrich-Alexander-Universität Erlangen-Nürnberg and Universitätsklinikum Erlangen, Erlangen, Germany
- <sup>3</sup> Member of German Center for Lung Research DZL, LMU Munich, Munich, Germany

Air pollution and immune-related diseases including allergy and asthma are constantly on the rise worldwide. Thus, a comprehensive investigation of environmentally induced immune regulation is required for a deeper understanding of disease pathogenesis, progression as well as prevention. Here, we summarize the current knowledge on environmental factors such as microbiome or geographical locations with harmful or protective effects for human health and their different routes of exposure. This review comprises a brief outline regarding the latest findings on the interaction of environmental factors with innate and adaptive regulation of the immune system, exemplarily for one protective and one harmful environmental factor, respectively.

**Keywords:** Air pollution ⋅ Allergy ⋅ Asthma ⋅ Environment ⋅ Immune regulation

#### Introduction

Over the last decades, the prevalence of immune-related diseases including allergy and asthma has increased progressively within the last decades [1–3]. There is growing evidence suggesting a decisive role of environmental influences in shaping early immune maturation including the host's responses to allergens [4–6]. Thus, an increase in immune-related diseases is suggested to be influenced by (disadvantageous) environmental changes including the increment of harmful and the loss of protective exposures associated with progressive western/industrialized/urbanassociated lifestyle. The rapidly advancing change in lifestyle as well as environments have in a way challenged the regulatory capacity of the healthy human immune system prompting an increasing interest to identify and evaluate harmful and protective factors.

Correspondence: Prof. Bianca Schaub e-mail: Bianca.Schaub@med.uni-muenchen.de This review summarizes the current knowledge on environmental factors with harmful or protective effects and their different routes of exposure. In addition, a brief outline about the latest findings on the interaction of environmental factors with innate and adaptive components of the immune system from *in vivo* studies is given exemplary for one protective and one harmful environmental factor, respectively.

## Routes of exposure

The human organism is constantly exposed to diverse biological, chemical, and physical influences from the external environment. Conceptually, cumulative lifetime environmental exposure and related physiological responses are termed *human exposome* [7] that affects health, behavior, and physiology. In humans, several efficient barrier organs regulate the responses to environmental exposures that enter primarily via the skin, mucosal tissue, and inhalation. Importantly, the microbiota is involved

in early-life imprinting of local immune compartments, thereby promoting immune tolerance to antigens and wound healing [8, 9].

#### Skin exposure

The multiple layers and mechanical barriers of the skin control penetration of environmental substances, thereby protecting the organism from exposure-related effects. Physical defense mechanisms including functional integrity of barriers formed by hair follicles, skin glands, and blood vessels prevent systemic entry and effects of external factors. The outermost barrier bordering the external environment, the stratum corneum, consists of tightly connected differentiated keratinocytes that perform first defense innate immune functions [10]. Additionally, innate and adaptive immune cells including dermal and epidermal dendritic cells, macrophages, lymphocytes, mast cells, as well as complement factors are involved in immune responses in the skin. However, absorption of skin-penetrating substances through an intact and functional dermal epithelial barrier is possible [10].

#### Mucosal tissue exposure

Mucosal surface integrity and mucus secretion in the eye and nose, the upper respiratory tract, the gastrointestinal tract (GIT), and the urogenital tract are critical to impede diffusion and mediate elimination of invaded pollutants. Dysfunctional mucosal epithelial barriers have been associated with antigen invasion, inflammatory activation of epithelial cells, and allergic reactions leading to the development of allergic diseases [11, 12]. The GIT that is exposed to the external environment through ingested particles taken up by diet, inhalation, or other interventions exhibits a barrier formed by tightly connected epithelial cells lined with mucus and microbiota. However, exposure to noxious factors can mediate intestinal barrier defects resulting in epithelial translocation of microbes and initiate inflammatory immune reactions [13]. Mucosal tissue is further exposed by inhaled particles and substances circulating in the air are trapped by nasal hairs and cilia in the nasal cavity, avoiding deeper penetration into the trachea. Mucus secreted by goblet cells and hair-like cilia lining the trachea and bronchi trap foreign particles and push them back outwards through constant cilial movement. Despite these highly efficient mechanisms, particulate matter (PM) with a diameter of  $\leq$ 10  $\mu$ m (PM<sub>10</sub>) reach the upper respiratory tract and ultrafine particles (PM<sub>0.1</sub>) can even get deep inside the lung affecting lung alveoli and local microbiota, penetrating the lung epithelial barrier and entering the systemic blood circulation [14]. The extent of inhalation exposure depends on the size and penetration capacity of inhaled matter, on indoor and outdoor air quality, and preliminary organ dam-

# Environmental factors with immunoreactive potential

Environmental exposures influencing the human organism comprise small particles and chemical residuals, physical influences, microbes, geographical location, as well as lifestyle choices. Depending on the exposure time, extent, and dosage, environmental factors can either elicit immune-potentiating or immunosuppressive responses. Importantly, environmental exposures add up during an individual's lifespan and can mediate delayed reactions. Importantly, evidence suggests that combinations of environmental influences such as combined wood smoke and diesel particles will act in an additive or synergistic manner [15]. For the sake of brevity, we here review the immunoreactive effect of individual environmental factors and will not discuss the impact of the combination of environmental influences.

Some environmental factors have beneficial effects on immune priming and function as well as disease prevention such as plant-derived bioaerosols and dust. Specifically, farm-related exposures induce anti-inflammatory immune reactions, alleviate allergic and autoimmune responses, and if occurring in the prenatal or early-life period, reduce the risk for autoimmune diseases [16–21]. Other naturally occurring bioactive compounds with beneficial immunomodulating properties comprise polyphenols that are subclassified into flavonoids and non-flavonoids and display an anti-neuroinflammatory potential and alleviate inflammation-associated depressive symptoms [22].

In contrast, air pollutants such as PM, a complex air suspension of solid and liquid, organic and inorganic particles, as well as nanoparticles, environmental contaminants of small dimensions (1-100 nm), can negatively alter immune activity and function [23, 18]. In mice, chronic ultrafine PM (PM<sub>2.5</sub>), diesel exhaust particles, and engineered nanoparticles induced systemic and neuroinflammatory immune responses [4, 23]. In humans, maternal exposure to traffic-related air pollutants, such as nitrogen oxide, PM, and ozone (O<sub>3</sub>), has been associated with increased inflammatory cytokine levels in newborns, impairing immune system development and postnatal immune functions [24, 23]. Further airborne factors with immunoreactive and inflammatory potential comprise cigarette smoke [25], organic agricultural dust [26], and microplastics [27, 28]. Chemicals, that is, poly- and perfluorinated alkyl substances (PFAS), were detected in human blood sera and have been associated with impaired vaccine-induced antibody responses [29-31]. However, the authors from a recent review article on the immunomodulation and exposure of PFAS conclude that more evidence is needed due to inconsistent results on the immunomodulatory effect of PFAS exposure [29]. In addition, prenatal exposure to phthalates that act as endocrine disruptors has been associated with male reproductive disorders in mice. Few clinical studies were performed which revealed that phthalate metabolites can cross the placental barrier subsequently impairing genital development in human male fetuses [32, 33]. Moreover, environmental phthalate exposure in children was associated with allergic symptoms such as atopic dermatitis, asthma, and allergic rhinitis [34]. Chemical

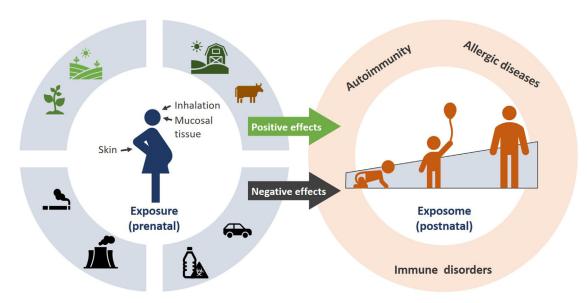


Figure 1. Prenatal and postnatal exposures: routes, effects, and outcomes. Starting in the prenatal period through maternal exposure, individuals are exposed to a variety of environmental influences by different routes (left circle). These can either elicit positive and beneficial effects on immune priming or predispose for the development of immune-related diseases later in life. Accumulation of environmental influences over the lifetime (exposome) results in the development of immune disorders at different ages.

exposure also affects the microbiota that is essential for intact host immune functioning. In children, the chemical exposome was related to altered early-life microbiota development, potentially favoring altered immune functions and associated diseases later in life such as asthma [35, 13].

Overall, environmental exposure is especially critical during vulnerable developmental periods and *in utero* or neonatal exposure alter immune maturation, function, and development (Fig. 1).

# Influence of environmental factors on innate and adaptive immune responses

As a result of the variability of environmental factors, the following paragraph aims to set the spotlight exemplarily on one of each protective and harmful factor for asthma and allergic diseases (Fig. 2). Concerning protective properties, several epidemiological studies worldwide have shown significantly lower prevalence in asthma and allergic diseases for children growing up on farms, a phenomenon known as "protective farm effect". Herein, contact to livestock and fodder, consumption of raw milk and microbial diversity are crucial [20, 36-40, 21, 41]. Environmentally mediated allergy and asthma-protective effects were replicated robustly in Amish children compared to children from Hutterite people (5.2% vs. 21.3% asthma prevalence). Despite their similar geographic and genetic background, both groups differ in their farming lifestyle with more traditional farming in the Amish and highly industrialized practices in the Hutterites [42]. The protective capacity of these rural agricultural environments has already been linked to a variety of immunological components. Regarding the innate part of the immune system, farm exposure was associated with altered functional properties of dendritic cells (DC), cells that are in first contact with environmental factors and thus influence T-cell responses. Immunomodulatory effects by decreasing myeloid dendritic cell (mDC) proportions related to farming lifestyle were shown [43, 44] coinciding with significantly elevated expression of the pathogen sensing innate immune receptors, such as Toll-like receptor (TLR)2, TLR7, TLR8, and cluster of differentiation (CD) 14 [6, 45]. In addition, gene-by-environment interaction for the single nucleotide polymorphism rs2230626 in the tumor necrosis factor, alpha-induced protein 3 (TNFAIP3) gene could have been connected with asthma and allergy protective effects in children growing up in farm-related environments [46]. Regarding the adaptive branch of the immune system, reduced lymphocyte proliferation in farm children and a positive association between early (in utero) exposure to farming lifestyle and regulatory T lymphocyte (Treg) numbers as well as T<sub>reg</sub> function was shown. However, an immunological switch results in lower T<sub>reg</sub> levels in farm children at school age suggesting inter alia the existence of a critical window during early healthy immune maturation both in terms of flexibility and variability but also in timing of immunity [5, 47-49]. Moreover, farm exposure was connected to a T helper lymphocyte (TH)1 dominating cytokine profile with elevated interleukin (IL)-12, tumor necrosis factor alpha (TNF-α), and interferon gamma (IFN-γ) levels [50, 51] accompanied by reduced levels of the T<sub>H</sub>2 cytokine IL-5 [5].

The rapid increase in asthma prevalence in recent decades may therefore be partly mediated by the absence of a protective farm-related environment due to progressively dominating urbanized lifestyles accompanied by a growing trend for exposure to harmful environmental factors potentially playing a causal role in disease development. Epidemiological data, published by the

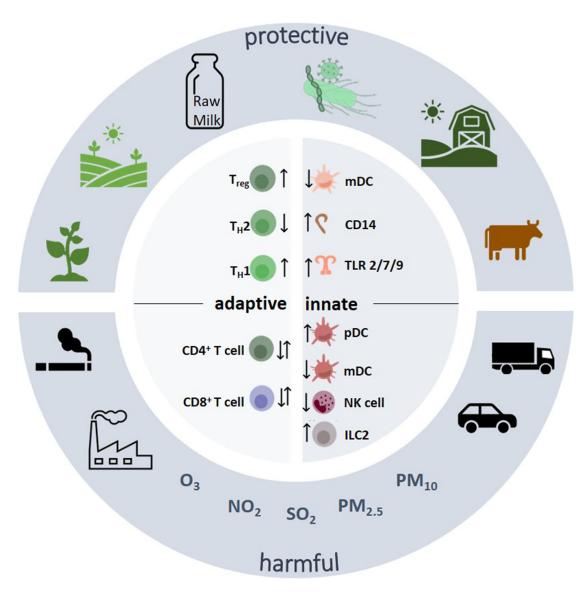


Figure 2. Influence of asthma and allergy protective as well as harmful environmental factors on innate and adaptive immune cells as described in invivo studies. Arrows indicate changes in cellular composition ( $\uparrow$ : upregulation,  $\downarrow$ : downregulation,  $\uparrow\downarrow$ : different findings in different studies,  $T_{reg}$ : regulatory T Lymphocyte,  $T_H1$ : T helper lymphocyte 1,  $T_H2$ : T helper lymphocyte 2, CD: cluster of differentiation, mDC: myeloid dendritic cell, pDC: plasmacytoid dendritic cell, TLR: Toll-like receptor, NK: natural killer, ILC2: innate lymphoid cell 2, NO<sub>2</sub>: nitrogen dioxide, O<sub>3</sub>: ozone, PM<sub>10</sub>: particulate matter with a diameter of  $\leq$ 10  $\mu$ m, PM<sub>2.5</sub>: particulate matter with a diameter of  $\leq$ 2.5  $\mu$ m, SO<sub>2</sub>: sulfur dioxide).

World Health Organization (WHO) as part of the 2018 update, indicate that 90% of people are exposed to air containing high levels of indoor and outdoor pollutants. Moreover, more than 80% of urban living people breathe air containing pollutant levels that exceed the WHO guidelines [52]. Thus, elevated asthma risk in children could have already been linked to prenatal harmful environmental exposure, namely cigarette smoke, accompanied by impaired  $T_{\rm H}1$  responses shifted toward a  $T_{\rm H}2$  dominating phenotype and increased pro-inflammatory conditions (secretion of IL-17A, IL-8, IL-6, and TNF- $\alpha$ ) [53]. However, a causal link between asthma and air pollution in general, not only cigarette smoke, has been suggested. Regarding the innate branch of the immune system, higher numbers in plasmacytoid dendritic cells (pDCs) as well as reduced numbers ofmDCs and natural killer (NK)

cells were described after *in utero* exposure to air pollution [54]. Moreover, in a human study including 118 healthy and asthmatic patients, exposure to  $PM_{10}$  and  $PM_{2.5}$  was shown to positively correlate with innate lymphoid cell type 2 (ILC2) frequencies, innate cells functioning in a similar way to asthma and allergy-mediating  $T_{\rm H}2$  lymphoid cells [55, 56]. Furthermore,  $PM_{10}$  exposure was related with increasing ILC2 frequencies in severe asthmatics compared to mild asthmatics with  $PM_{10}$  exposure shown to act in a rather chronic way since ILC2 levels were shown to correlate with the altitude of exposure level. No such correlation for  $PM_{2.5}$  in severe asthmatics could be described. ILC1 cells were shown to correlate with  $O_3$ ,  $NO_2$ , and carbon monoxide (CO) exposure, but not with PM [56]. Concerning the adaptive branch of the immune system, latest publications described, although contradictory,

disturbed lymphocyte distributions after in utero exposure to airpolluting substances including PM<sub>2.5</sub>, PM<sub>10</sub>, NO<sub>2</sub>, or SO<sub>2</sub>. While Martins Costa Gomes and co-workers described reduced numbers of CD4<sup>+</sup> and elevated numbers of CD8<sup>+</sup> cells after SO<sub>2</sub> exposure, Garcia-Serna and colleagues reported reduced numbers of CD8+ T cells, decreased numbers of  $T_{\text{reg}}$  cells, and elevated numbers of T<sub>H</sub>1 cells connected with PM exposure during pregnancy [57, 54]. Moreover, gestational exposure to traffic-related air pollution (TRAP; mix of NO<sub>2</sub>, PM<sub>2.5</sub>, PM<sub>10</sub>, O<sub>3</sub>) was also shown to increase pro-inflammatory (IL-1β and IL-6), T<sub>H</sub>2 related (IL-13), and immunomodulatory (IL-10) cytokines in newborn offspring [24]. Contrarily, Hahn and colleagues showed a reduced production of IL-6, TNF- $\alpha$ , and IL-10 in cord blood mononuclear cells (CBMCs) of children from mothers exposed to higher levels of PM2.5 during pregnancy [58]. Beyond the in vivo data, the focus of this review, Deckers et al. as well as Krusche et al. described recent findings regarding the mechanisms that confer protection against asthma and allergies including mouse and in vitro studies [59, 60].

#### Conclusion

Multiple environmental factors, classified either as protective or harmful, interact with and thus modulate the innate as well as the adaptive part of the human immune system and its functionality via different routes of exposure. Here, early life represents a critical window of opportunity but also vulnerability influencing the subsequent development of a child's immune system. Pre- as well as postnatal environmental exposures shape the still developing immune system by regulation of innate and adaptive cellular differentiation, (un)balancing T cell responses, as well as modulation of cytokine secretion. However, the exact cellular mechanisms underlying these harmful or protective environmental factors on disease prevention and/or disease progression including asthma and allergic diseases still require further research. This is key for future prevention strategies as well as for efficient counselling of government and authorities.

Acknowledgements: This work was supported by the DFG grants SCHA 997/9-1, SCHA 997/10-1, and SCHA 997/11-1. Open access funding enabled and organized by Projekt DEAL.

Conflict of interest: The authors declare no commercial or financial conflict of interest.

Author contributions: CB and BS conceived the study. CB and RM drafted the manuscript; all authors have revised and approved the final manuscript.

Data availability statement: Data sharing is not applicable to this article as no new data were created or analyzed in this study.

#### References

- 1 Poethko-Müller, C., Thamm, M. and Thamm, R., Allergic rhinitis and asthma among children and adolescents in Germany. Results of the cross-sectional KiGGS Wave 2 study and trends. *J. Health Monitor.* 2018. 3: 52–56.
- 2 Eder, W., Ege, M. J. and von Mutius, E., The asthma epidemic. N Engl J Med. 2006. 355: 2226–2235.
- 3 Global Asthma Network. The global asthma report 2018. Global Asthma Network, Auckland, New Zealand, 2018.
- 4 Morris-Schaffer, K., Merrill, A. K., Wong, C., Jew, K., Sobolewski, M. and Cory-Slechta, D. A., Limited developmental neurotoxicity from neonatal inhalation exposure to diesel exhaust particles in C57BL/6 mice. *Part. Fibre Toxicol.* 2019. 16: 1.
- 5 Schaub, B., Liu, J., Höppler, S., Schleich, I., Huehn, J., Olek, S., Wieczorek, G. et al., Maternal farm exposure modulates neonatal immune mechanisms through regulatory T cells. *J Allergy Clin Immunol*. 2009. 123: 774.e5–782.e5.
- 6 Loss, G., Bitter, S., Wohlgensinger, J., Frei, R., Roduit, C., Genuneit, J., Pekkanen, J. et al., Prenatal and early-life exposures alter expression of innate immunity genes: the PASTURE cohort study. *J Allergy Clin Immunol*. 2012. 130: 523.e9–530.e9.
- 7 Miller, G. W. and Jones, D. P., The nature of nurture: refining the definition of the exposome. *Toxicological Sciences*. 2014. 137: 1–2.
- 8 Constantinides, M. G., Link, V. M., Tamoutounour, S., Wong, A. C., Perez-Chaparro, P. J., Han, S. J., Chen, Y. E. et al., MAIT cells are imprinted by the microbiota in early life and promote tissue repair. *Science*. 2019. 366: eaax6624.
- 9 Gollwitzer, E. S., Saglani, S., Trompette, A., Yadava, K., Sherburn, R., McCoy, K. D., Nicod, L. P. et al., Lung microbiota promotes tolerance to allergens in neonates via PD-L1. Nat Med. 2014. 20: 642–647.
- 10 Passeron, T., Zouboulis, C. C., Tan, J., Andersen, M. L., Katta, R., Lyu, X., Aguilar, L. et al., Adult skin acute stress responses to short-term environmental and internal aggression from exposome factors. *J Eur Acad Dermatol Venereol.* 2021. 35: 1963–1975.
- 11 Noureddine, N., Chalubinski, M. and Wawrzyniak, P., The role of defective epithelial barriers in allergic lung disease and asthma development. J. Asthma Allergy. 2022. 15: 487–504.
- 12 Singh, N., Diebold, Y., Sahu, S. K. and Leonardi, A., Epithelial barrier dysfunction in ocular allergy. *Allergy*. 2022. 77: 1360–1372.
- 13 Ayeni, K. I., Berry, D., Wisgrill, L., Warth, B. and Ezekiel, C. N., Early-life chemical exposome and gut microbiome development: African research perspectives within a global environmental health context. *Trends Micro-biol.* 2022. 30: 1084–1100.
- 14 El Morabet, R., Effects of outdoor air pollution on human health. Reference module in earth systems and environmental sciences. Elsevier, 2018.
- 15 Adetona, A. M., Adetona, O., Gogal, R. M., Diaz-Sanchez, D., Rathbun, S. L. and Naeher, L. P., Impact of work task-related acute occupational smoke exposures on select proinflammatory immune parameters in wildland firefighters. *J Occup Environ Med.* 2017. 59: 679–690.
- 16 Frei, R., Heye, K. and Roduit, C., Environmental influences on childhood allergies and asthma—the Farm effect. *Pediatric Allergy Immunol.* 2022. 33: e13807.
- 17 Steiman, C. A., Evans, M. D., Lee, K. E., Lasarev, M. R., Gangnon, R. E., Olson, B. F., Barnes, K. L. et al., Patterns of farm exposure are associ-

- ated with reduced incidence of atopic dermatitis in early life. J Allergy Clin Immunol. 2020. 146: 1379.e6-1386.e6.
- 18 Martikainen, M. V., Rönkkö, T. J., Schaub, B., Täubel, M., Gu, C., Wong, G. W., Li, J. et al., Integrating farm and air pollution studies in search for immunoregulatory mechanisms operating in protective and high-risk environments. Pediatric Allergy Immunol. 2018. 29: 815–822.
- 19 Stiehm, M., Bufe, A. and Peters, M., Proteolytic activity in cowshed dust extracts induces C5a release in murine bronchoalveolar lavage fluids which may account for its protective properties in allergic airway inflammation. Thorax. 2013. 68: 31-38.
- 20 Illi, S., Depner, M., Genuneit, J., Horak, E., Loss, G., Strunz-Lehner, C., Büchele, G. et al., Protection from childhood asthma and allergy in Alpine farm environments—the GABRIEL advanced studies. J Allergy Clin Immunol. 2012. 129: 1470.e6-1477.e6.
- 21 Douwes, J., Cheng, S., Travier, N., Cohet, C., Niesink, A., McKenzie, J., Cunningham, C. et al., Farm exposure in utero may protect against asthma, hay fever and eczema. Eur Respir J. 2008. 32: 603-611.
- 22 Tayab, M. A., Islam, M. N., Chowdhury, K. A. A. and Tasnim, F. M., Targeting neuroinflammation by polyphenols: a promising therapeutic approach against inflammation-associated depression. Biomed Pharmacother. 2022. 147: 112668.
- 23 Gong, J.-Y., Holt, M. G., Hoet, P. H. M. and Ghosh, M., Neurotoxicity of four frequently used nanoparticles: a systematic review to reveal the missing data. Arch Toxicol. 2022. 96: 1141-1212.
- 24 García-Serna, A. M., Martín-Orozco, E., Jiménez-Guerrero, P., Hernández-Caselles, T., Pérez-Fernández, V., Cantero-Cano, E., Muñoz-García, M. et al., Cytokine profiles in cord blood in relation to prenatal trafficrelated air pollution: the NELA cohort. Pediatric Allergy Immunol. 2022. 33: e13732.
- 25 Ou, G., Zhu, M., Huang, Y., Luo, W., Zhao, J., Zhang, W., Xia, H. et al., HSP60 regulates the cigarette smoke-induced activation of TLR4-NF-kB-MyD88 signalling pathway and NLRP3 inflammasome. Int Immunopharmacol. 2022. 103: 108445.
- 26 Ulu, A., Sveiven, S., Bilg, A., Velazquez, J. V., Diaz, M., Mukherjee, M., Yuil-Valdes, A. G. et al., IL-22 regulates inflammatory responses to agricultural dust-induced airway inflammation. Toxicol. Appl. Pharmacol. 2022. 446: 116044.
- 27 Jenner, L. C., Rotchell, J. M., Bennett, R. T., Cowen, M., Tentzeris, V. and Sadofsky, L. R., Detection of microplastics in human lung tissue using μFTIR spectroscopy. Sci. Total Environ. 2022. 831: 154907.
- 28 Li, Y., Shi, T., Li, X., Sun, H., Xia, X., Ji, X., Zhang, J. et al., Inhaled tire-wear microplastic particles induced pulmonary fibrotic injury via epithelial cytoskeleton rearrangement. Environ. Int. 2022. 164:
- 29 Antoniou, E., Colnot, T., Zeegers, M. and Dekant, W., Immunomodulation and exposure to per- and polyfluoroalkyl substances: an overview of the current evidence from animal and human studies. Arch Toxicol. 2022. 96: 2261-2285
- 30 Shih, Y.-H., Blomberg, A. J., Bind, M.-A., Holm, D., Nielsen, F., Heilmann, C., Weihe, P. et al., Serum vaccine antibody concentrations in adults exposed to per- and polyfluoroalkyl substances: a birth cohort in the Faroe Islands. J Immunotoxicol. 2021. 18: 85-92.
- 31 Kielsen, K., Shamim, Z., Ryder, L. P., Nielsen, F., Grandjean, P., Budtz-Jørgensen, E. and Heilmann, C., Antibody response to booster vaccination with tetanus and diphtheria in adults exposed to perfluorinated alkylates. J Immunotoxicol. 2016. 13: 270-273.
- 32 Yu, C., Lu, J., Zhao, J., Zhao, T., Long, C., Lin, T., Wu, S. et al., Maternal phthalate exposure during pregnancy and male reproductive disorders: a systematic review and metaanalysis. Turk J Pediatr. 2022. 64: 187.

- 33 Martino-Andrade, A. J., Liu, F., Sathyanarayana, S., Barrett, E. S., Redmon, J. B., Nguyen, R. H. N., Levine, H. et al., Timing of prenatal phthalate exposure in relation to genital endpoints in male newborns. Andrology. 2016. 4: 585-593
- 34 Lee, J. Y., Lee, J., Huh, D. A. and Moon, K. W., Association between environmental exposure to phthalates and allergic disorders in Korean children: Korean National Environmental Health Survey (KoNEHS) 2015-2017. Int. J. Hygiene Environ. Health. 2021. 238: 113857.
- 35 Arrieta, M. C., Stiemsma, L. T., Dimitriu, P. A., Thorson, L., Russell, S., Yurist-Doutsch, S., Kuzeljevic, B. et al., Early infancy microbial and metabolic alterations affect risk of childhood asthma. Sci Transl Med. 2015.
- 36 Timm, S., Frydenberg, M., Janson, C., Campbell, B., Forsberg, B., Gislason, T., Holm, M. et al., The urban-rural gradient in asthma: a populationbased study in northern Europe. Int J Environ Res Public Health. 2015. 13:
- 37 Ege, M. J., Frei, R., Bieli, C., Schram-Bijkerk, D., Waser, M., Benz, M. R., Nyberg, F. et al., Not all farming environments protect against the development of asthma and wheeze in children. J Allergy Clin Immunol. 2007. 119: 1140-1147.
- 38 Roduit, C., Wohlgensinger, J., Frei, R., Bitter, S., Bieli, C., Loeliger, S., Büchele, G. et al., Prenatal animal contact and gene expression of innate immunity receptors at birth are associated with atopic dermatitis. JAllergy Clin Immunol. 2011. 127: 179-185.
- 39 von Mutius, E. and Vercelli, D., Farm living: effects on childhood asthma and allergy. Nat Rev Immunol. 2010. 10: 861-868.
- 40 Birzele, L. T., Depner, M., Ege, M. J., Engel, M., Kublik, S., Bernau, C., Loss, G. J. et al., Environmental and mucosal microbiota and their role in childhood asthma. Allergy. 2017. 72: 109-119.
- 41 Loss, G., Depner, M., Ulfman, L. H., van Neerven, R. J. J., Hose, A. J., Genuneit, J., Karvonen, A. M. et al., Consumption of unprocessed cow's milk protects infants from common respiratory infections. J Allergy Clin Immunol, 2015, 135: 56-62.
- 42 Holbreich, M., Genuneit, J., Weber, J., Braun-Fahrländer, C., Waser, M. and M. E. von, Amish children living in northern Indiana have a very low prevalence of allergic sensitization. J Allergy Clin Immunol. 2012. 129: 1671-1673.
- 43 Kääriö, H., Nieminen, J. K., Karvonen, A. M., Huttunen, K., Schröder, P. C., Vaarala, O., von Mutius, E. et al., Circulating dendritic cells, farm exposure and asthma at early age. Scand J Immunol. 2016. 83: 18-25.
- 44 Martikainen, M. V., Kääriö, H., Karvonen, A., Schröder, P. C., Renz, H., Kaulek, V., Dalphin, J. C. et al., Farm exposures are associated with lower percentage of circulating myeloid dendritic cell subtype 2 at age 6. Allergy. 2015. 70: 1278-1287.
- 45 Lauener, R. P., Birchler, T., Adamski, J., Braun-Fahrländer, C., Bufe, A., Herz, U., von Mutius, E. et al., Expression of CD14 and Toll-like receptor 2 in farmers' and nonfarmers' children. Lancet. 2002. 360: 465-466.
- 46 Schuijs, M. J., Willart, M. A., Vergote, K., Gras, D., Deswarte, K., Ege, M. J., Madeira, F. B. et al., Farm dust and endotoxin protect against allergy through A20 induction in lung epithelial cells. Science. 2015. 349: 1106-
- 47 Lluis, A., Depner, M., Gaugler, B., Saas, P., Casaca, V. I., Raedler, D., Michel, S. et al., Increased regulatory T-cell numbers are associated with farm milk exposure and lower atopic sensitization and asthma in childhood. JAllergy Clin Immunol. 2014. 133: 551-559.
- 48 Ege, M. J., Herzum, I., Büchele, G., Krauss-Etschmann, S., Lauener, R. P., Roponen, M., Hyvärinen, A. et al., Prenatal exposure to a farm environment modifies atopic sensitization at birth. J Allergy Clin Immunol. 2008. 122: 407-412, 412.e1-4.

15214141, 2023, 6, Downloaded from https://onlinelibrary.wiley.com/doi/10.1002/eji.202249981, Wiley Online Library on [21/03/2024]. See the Terms and Conditions (https://onlinelibrary.wiley.com/terms-and-conditions) on Wiley Online Library for rules of use; OA articles are governed by the applicable Ceative Commons License

- 49 Schröder, P. C., Illi, S., Casaca, V. I., Lluis, A., Böck, A., Roduit, C., Depner, M. et al., A switch in regulatory T cells through farm exposure during immune maturation in childhood. *Allergy*. 2017. 72: 604–615.
- 50 Pfefferle, P. I., Büchele, G., Blümer, N., Roponen, M., Ege, M. J., Krauss-Etschmann, S., Genuneit, J. et al., Cord blood cytokines are modulated by maternal farming activities and consumption of farm dairy products during pregnancy: the PASTURE Study. J Allergy Clin Immunol. 2010. 125: 108.e1-3-115.e1-3.
- 51 Kääriö, H., Huttunen, K., Karvonen, A. M., Schaub, B., M. E. von, Pekkanen, J., Hirvonen, M. R. et al., Exposure to a farm environment is associated with T helper 1 and regulatory cytokines at age 4.5 years. Clin Exp Allergy. 2016, 46: 71–77.
- 52 World Health Organization. WHO global ambient air quality database (update 2018), 2018.
- 53 Strzelak, A., Ratajczak, A., Adamiec, A. and Feleszko, W., Tobacco smoke induces and alters immune responses in the lung triggering inflammation, allergy, asthma and other lung diseases: a mechanistic review. *Int J Environ Res Public Health.* 2018. 15: 1033.
- 54 García-Serna, A. M., Hernández-Caselles, T., Jiménez-Guerrero, P., Martín-Orozco, E., Pérez-Fernández, V., Cantero-Cano, E., Muñoz-García, M. et al., Air pollution from traffic during pregnancy impairs newborn's cord blood immune cells: the NELA cohort. *Environ Res.* 2021. 198: 110468.
- 55 Klein Wolterink, R. G. J., Kleinjan, A., van Nimwegen, M., Bergen, I., de, Bruijn, M., Levani, Y. and Hendriks, R. W. et al., Pulmonary innate lymphoid cells are major producers of IL-5 and IL-13 in murine models of allergic asthma. Eur J Immunol. 2012. 42: 1106–1116.
- 56 Kim, J., Kim, Y. C., Ham, J., Sohn, K. H., Lee, S. Y., Chung, D. H., Cho, S. H. et al., The effect of air pollutants on airway innate immune cells in patients with asthma. *Allergy*. 2020. 75: 2372–2376.

- 57 Martins Costa Gomes, G., Karmaus, W., Murphy, V. E., Gibson, P. G., Percival, E., Hansbro, P. M., Starkey, M. R. et al., Environmental air pollutants inhaled during pregnancy are associated with altered cord blood immune cell profiles. *Int J Environ Res Public Health*. 2021. 18: 7431.
- 58 Hahn, J., Gold, D. R., Coull, B. A., McCormick, M. C., Finn, P. W., Perkins, D. L., Shiman, S. L. R. et al., Air pollution, neonatal immune responses, and potential joint effects of maternal depression. *Int J Environ Res Public Health*. 2021. 18: 5062.
- 59 Deckers, J., Marsland, B. J. and von, Mutius, E., Protection against allergies: microbes, immunity, and the farming effect. Eur J Immunol. 2021. 51: 2387–2398.
- 60 Krusche, J., Basse, S. and Schaub, B., Role of early life immune regulation in asthma development. Semin Immunopathol. 2020. 42: 29–42.

Abbreviations: CD: cluster of differentiation  $\cdot$  ILC: innate lymphoid cell  $\cdot$  PM: particulate matter  $\cdot$  PM10: particulate matter with a diameter of  $\leq$  10  $\mu$ m

Full correspondence: Prof. Bianca Schaub, Dr. von Hauner Children's Hospital, University Hospital, Dep. of Allergology, LMU Munich, Lindwurmstr. 4, 80337 Munich, Germany e-mail: Bianca.Schaub@med.uni-muenchen.de

Received: 30/9/2022 Revised: 11/1/2023 Accepted: 13/1/2023

Accepted article online: 16/1/2023