

## **Secretases in Alzheimer's disease: Novel insights into proteolysis of APP and TREM2**

Stefan F. Lichtenthaler<sup>1,2,3</sup>, Sarah K. Tschirner<sup>1,2</sup> and Harald Steiner<sup>1,4</sup>

<sup>1</sup>German Center for Neurodegenerative Diseases (DZNE), Munich, Germany;

<sup>2</sup>Neuroproteomics, School of Medicine, Klinikum rechts der Isar, Technical University of Munich, 81675 Munich, Germany

<sup>3</sup>Munich Cluster for Systems Neurology (SyNergy), Munich, Germany

<sup>4</sup>Biomedical Center (BMC), Division of Metabolic Biochemistry, Faculty of Medicine, LMU Munich

To whom correspondence should be addressed:

[stefan.lichtenthaler@dzne.de](mailto:stefan.lichtenthaler@dzne.de) or [harald.steiner@med.uni-muenchen.de](mailto:harald.steiner@med.uni-muenchen.de)

Postal address: Feodor-Lynen-Street 17, 81377 Munich, Germany, +49.89.4400.46426 (SFL) or +49.89.4400.46535 (HS), Fax: +49.89.4400.46408

### **Highlights**

- Sfrp1, RECK, Atxn1 and Nrf2 control ADAM10 and BACE1 activity and are linked to AD
- New familial Alzheimer Uppsala mutation shifts APP cleavage from  $\alpha$ - to  $\beta$ -secretase
- Structure and pharmacology studies support drug development of  $\gamma$ -secretase modulators
- TREM2-agonistic antibodies show potential for reducing brain amyloidosis in mice

**Short Title: Secretases in Alzheimer's disease**

## **Abstract**

Secretases are a group of proteases that are major drug targets considered for prevention and treatment of Alzheimer's disease (AD). Secretases do not only process the AD-linked neuronal amyloid precursor protein (APP), but also the triggering receptor expressed on myeloid cells 2 (TREM2), thereby controlling microglial functions. This review highlights selected, recent discoveries for the  $\alpha$ -secretases ADAM10 and ADAM17, the  $\beta$ -secretase BACE1 and  $\gamma$ -secretase and their link to AD. New genetic evidence strengthens the role for  $\alpha$ -secretases in AD through cleavage of APP and TREM2. Novel proteins were linked to AD which control  $\alpha$ - and  $\beta$ -secretase activity through transcriptional and post-translational mechanisms. Finally, new opportunities, but also challenges are discussed for pharmacologically targeting  $\beta$ - and  $\gamma$ -secretase cleavage of APP and  $\alpha$ -secretase cleavage of TREM2, with the aim to prevent or treat AD.

**Keywords: Alzheimer's disease, APP, RECK, secretases, Sfrp1, TREM2**

## Introduction

Alzheimer's disease (AD) is the most common neurodegenerative disease and affects more than 50 million patients worldwide. The onset of clinical AD symptoms is preceded by an approximately 20 year-long asymptomatic disease pathogenesis that comprises a complex series of molecular and cellular events, referred to as the amyloid cascade hypothesis [1] (Fig. 1) which is based on genetic and epidemiological evidence [2]. The cascade starts with the formation of small, oligomeric aggregates of the hydrophobic amyloid  $\beta$  ( $A\beta$ ) peptide, which proteolytically derives from the amyloid precursor protein (APP). Because  $A\beta$  is mostly produced and released from neurons, AD pathogenesis was long seen from a neuron-centric perspective. However, this view changed in the recent past with the discovery that numerous genetic AD risk factors for late onset AD (LOAD), such as apolipoprotein E (ApoE) and triggering receptor expressed on myeloid cells 2 (TREM2), are strongly or even exclusively expressed in microglia [3]. This observation led to the discovery of beneficial, but also detrimental functions for microglia in AD pathogenesis. As immune cells of the brain, microglia can adapt to changing environments by modulating gene expression from homeostatic to protective disease-associated states [4,5]. Microglia can phagocytose and, thus, reduce abundance of  $A\beta$ , but also contribute to  $A\beta$  aggregation and excessive neuroinflammation (Fig. 1). TREM2 has emerged as a major player governing beneficial microglial defense responses to pathological  $A\beta$  deposition including microglial proliferation, chemotaxis, motility and survival in addition to  $A\beta$  phagocytosis [3]. As a defense mechanism to AD pathology, TREM2 appears to sense  $A\beta$  aggregates early on and, together with ApoE, with which TREM2 physically interacts, triggers a probably beneficial compaction of amyloid plaques to reduce neuritic damage as well as to prevent the release of toxic oligomeric  $A\beta$  forms from the outer plaque halos by providing a microglial barrier function [3].

Proteolysis controls abundance or activity of most proteins. In AD, proteolysis has been studied most intensively for APP as it regulates generation and abundance of  $A\beta$  [1]. Similar proteolytic processing pathways have been identified more recently for TREM2 and control the phagocytic activity of microglia, which is one mechanism to degrade  $A\beta$  [3]. Pharmacological modulation of APP and TREM2 proteolysis has been and is tested in clinical trials and may lead to a causative treatment or prevention of AD [3,6]. Notably, although received with controversy in the field, immunotherapy-mediated removal of  $A\beta$  using the antibody aducanumab has recently been conditionally approved as the first causal therapy for AD in the U.S.

This review article focuses on the proteolytic processing of APP and TREM2. We will first introduce the proteases – referred to as secretases – that cleave APP and TREM2 and then

highlight recent discoveries on their genetic linkage to AD and their cellular regulation and provide an update on their suitability and use as drug targets for AD.

### **Secretases mediate APP processing and control A $\beta$ generation**

APP is a type I transmembrane protein and the precursor of the A $\beta$  peptide (Fig. 2). Cleavage of APP by  $\alpha$ -,  $\beta$ -,  $\delta$ -,  $\eta$ -secretase results in secretion of the large extracellular APP domain (Fig. 2) [7]. This process is referred to as ectodomain shedding and occurs for hundreds of membrane proteins besides APP, including TREM2 [8]. In contrast,  $\gamma$ -secretase gradually cleaves APP within its transmembrane domain (TMD) thereby releasing 37-43 residue-long secreted A $\beta$  peptides [9]. Depending on which proteases cleave APP, distinct proteolytic products are generated, which have different physiological functions or may contribute to AD [7] (Fig. 2).

In the following we focus on  $\alpha$ -,  $\beta$ - and  $\gamma$ -secretase, which have been drug targets in clinical trials for AD and for which major new developments occurred during the past few years. The  $\alpha$ -secretase activity cleaves APP in the middle of the A $\beta$  domain, which precludes A $\beta$  generation, and is predominantly mediated by 'a disintegrin and metalloprotease 10' (ADAM10) in the nervous system, but additional proteases such as the ADAM10-homolog ADAM17 may also contribute [10]. A pharmacological activation of APP cleavage by the  $\alpha$ -secretase ADAM10 is feasible in vitro and in AD patients, as tested in a small phase 2a study [11,12], but safety and clinical benefit of this approach need to be tested in larger patient cohorts.

Processing of APP first by  $\beta$ - and then  $\gamma$ -secretase leads to A $\beta$  generation (Fig. 2A).  $\beta$ -site APP cleaving enzyme 1 (BACE1) is the major  $\beta$ -secretase [13]. BACE2 is a homolog of BACE1, but cleaves within the A $\beta$  domain of APP and can prevent A $\beta$  generation [13,14]. Additional proteases, including meprin  $\beta$  and ADAMTS4, can cleave close to the BACE1 cleavage site and generate N-terminally truncated A $\beta$  peptides (Fig. 2B), some of which such as A $\beta$ 4-x are abundant in amyloid plaques [15,16]. Interestingly, BACE1 can also cleave within the A $\beta$  sequence after A $\beta$  has been generated, which may constitute an A $\beta$  clearance function that may be used to monitor AD progression [17].  $\gamma$ -Secretase is a protein complex in the membrane consisting of four subunits, the proteolytically active subunit presenilin (PS) and the three non-proteolytic subunits nicastrin (NCT), anterior pharynx defective-1 (APH-1) and presenilin enhancer 2 (PEN-2) required for complex assembly, stabilization and maturation [9] (Fig. 2C). Numerous PS mutations were identified in dominantly inherited forms of AD,

which shift the major  $\gamma$ -secretase cleavage site from position 40 to 42 or in some cases also to 43 (Fig. 2D), thus resulting in longer A $\beta$  peptides which aggregate more rapidly and form the major constituent of amyloid plaques [9].

### **Secretases cleave TREM2 and control its function in microglial phagocytosis**

TREM2 undergoes cleavages by several secretases in microglia (Fig. 3). In a manner similar to APP, TREM2 can be cleaved in its ectodomain by ADAM10 and/or ADAM17 whereby the major extracellular part of TREM2 is removed [18-21]. The TREM2 ectodomain comprises the functionally important ligand-binding domain, which senses anionic lipids exposed by damaged or dying cells, myelin or A $\beta$  [3]. An alternative ectodomain shedding cleavage by meprin  $\beta$  was reported to occur in macrophages [22]. Ectodomain shedding of TREM2 most likely is a major factor to cease cell-autonomous signaling, which is mediated by its membrane-bound binding partner DNAX-activation protein 12 (DAP12). Phosphorylation of DAP12 and subsequent binding of the kinase Syk induces downstream signaling pathways that control crucial microglial functions including microglial phagocytosis of loosely aggregated A $\beta$  of amyloid plaque halos [3]. Removal of A $\beta$  in the brain by this mechanism is stimulated by full-length (FL) TREM2 [3]. Certain mutations of TREM2 located in the ectodomain such as R47H, R62H or H157Y are associated with an increased risk of LOAD [3]. They functionally impact the ligand binding domain, interfere with maturation, ectodomain shedding or cell surface transport of TREM2 [18,20,21,23]. Similar to a knockout of TREM2, they have been shown to cause a loss of function (LOF) phenotype with impaired phagocytic activity [3].

The resulting TREM2 C-terminal fragments left in the membrane after ectodomain shedding can be further processed by  $\gamma$ -secretase thereby eventually terminating TREM2/DAP12-mediated signaling [18,24]. Cleavage occurs in a flexible region of its TMD after alanine A192 [25]. Interestingly, a mutation directly at the cleavage site (H157Y) has been identified in two Chinese AD patients (Fig. 3A) [26]. Its biochemical consequences have not yet been investigated but may yield interesting insights into the role of  $\gamma$ -secretase in TREM2 biology.

### **Genetics continues to provide novel insights into AD pathogenesis**

Identification of mutated genes or genetic associations has been instrumental in unraveling the molecular and cellular causes of AD. For example, as for PS, numerous amino acid mutations were found in APP that cause dominantly inherited AD [27]. Most of these mutations occur outside of the A $\beta$  sequence but in the vicinity of the  $\beta$ - and  $\gamma$ -secretase cleavage sites and generally enhance production of either the total amount of A $\beta$  or specifically the long, more rapidly aggregating A $\beta$ <sub>42</sub> peptide, thus accelerating AD pathogenesis. Very recently, a new

mutation was identified next to the  $\alpha$ -secretase cleavage site, which reduced cleavage of APP by  $\alpha$ -secretase and increased its processing by  $\beta$ -secretase, resulting in enhanced A $\beta$  generation. This Uppsala mutation results in deletion of amino acids 19-24 of the A $\beta$  sequence of APP (Fig. 2E) [28]. Interestingly, the same region within A $\beta$  was previously identified as a substrate inhibitory domain that suppresses A $\beta$  generation [29]. In addition to its A $\beta$ -increasing effect it also enhances A $\beta$  aggregation, similar to single amino acid mutations identified previously within the A $\beta$  sequence [27] (Fig. 2E) and provides further evidence for the central role of A $\beta$  oligomers early in AD pathogenesis.

New genetic associations to AD were also reported for ADAM10 and ADAM17. One family with a nonsense mutation (Y167\*) within the ADAM10 prodomain was identified. This predicted haploinsufficiency segregated with the disease and resulted in less  $\alpha$ -secretase cleavage of APP, as determined by CSF analysis of APP cleavage products [30]. ADAM10 was also linked to AD through genome-wide association studies [31,32], but it remains unclear how the ADAM10 locus alters ADAM10 expression and whether it affects A $\beta$  generation in neurons – similar to the nonsense mutation described above – or TREM2 shedding and phagocytosis in microglia, where ADAM10 is also expressed.

For ADAM17 a loss-of-function mutation (R215I) within the prodomain was associated with increased AD risk [33]. Mechanistically, this mutation was shown to enhance A $\beta$  levels in vitro, but additional mechanisms cannot yet be ruled out and need to be explored. This is particularly relevant, because ADAM17 acts a) as a TREM2 sheddase and, thus, may alter microglial phagocytosis of A $\beta$ , and b) cleaves TNF, thereby converting it to a pro-inflammatory, soluble cytokine, which is implicated in AD pathogenesis [34].

The role of TREM2 shedding in AD was further strengthened with the identification of the membrane-spanning 4-domains subfamily A (MS4A) gene cluster as a modulator of both soluble TREM2 and AD risk [35]. While that study revealed a colocalization of the MS4A protein MS4A4A with TREM2, further studies are needed to understand how this protein mechanistically controls TREM2 secretase cleavage.

### **Novel players regulating secretase activities**

The past two years also provided important new insights into cellular mechanisms controlling secretase activity, in particular of the  $\alpha$ -secretase ADAM10 and the  $\beta$ -secretase BACE1. ADAM10 activity is controlled at multiple levels [10,36]. Now, two additional ADAM10 inhibitory proteins - secreted frizzled-related protein 1' (Sfrp1) and the glycosylphosphatidylinositol

(GPI)-anchored membrane protein 'reversion-inducing cysteine-rich protein with Kazal motifs (RECK) - were directly linked to AD [37,38]. Both proteins were discovered to have enhanced abundance in AD brains, whereas genetic ablation of either protein enhanced ADAM10-cleavage of APP and reduced amyloid pathology in AD mice. Whether both proteins may be targeted pharmacologically in a safe manner remains to be studied.

For BACE1 exciting progress has been made in understanding its transcriptional regulation and the link thereof to AD. The gene expression regulatory protein ataxin-1 (Atxn1), which is linked to the neurodegenerative disorder spinocerebellar ataxia type 1, but also to AD, was found to repress BACE1 expression and control hippocampal neurogenesis and amyloid pathology in mouse brain [39]. Another transcription factor, 'nuclear factor erythroid-derived 2-related factor 2' (Nrf2), was also identified as a suppressor of BACE1 transcription [40]. Pathophysiological relevance of this Nrf2-dependent mechanism was established by demonstrating that NRF2 expression is reduced in AD brain and correlated with increased BACE1 protein levels.

BACE1 function may also be controlled by post-translational mechanisms, including by the intracellular trafficking protein  $\gamma$ -ear-containing ADP-ribosylation factor-binding protein 3 (GGA3). Last year, a loss-of-function mutation in GGA3 (Ins545T) was discovered that cosegregates with LOAD and mechanistically raises BACE1 abundance, leading to axonal trafficking disruption and axonal swellings in AD mice [41].

### **Therapeutic targeting of APP and TREM2 secretases**

Proteases are classical targets for therapeutic interventions and this includes the Alzheimer secretases. Over the past few years, important new developments occurred for the BACE1 and  $\gamma$ -secretase-cleavage of APP and for the ADAM10/ADAM17-mediated shedding of TREM2.

Regarding BACE1 inhibition, six different BACE1-targeted small molecule inhibitors were tested until recently in phase 2 and 3 clinical trials. Although the drugs showed excellent target engagement and lowered A $\beta$  in human brain and CSF, the trials were prematurely terminated because of futility, an unfavorable risk/benefit ratio or because of the occurrence of mild cognitive, but reversible worsening that is seen as an unacceptable side effect [42,43]. Analysis of the trial results has led to suggestions for future clinical trials, including the use of BACE inhibitors rather for prevention than for treatment of AD and a reduction of the inhibitor dose to avoid the side effects [6,13]. Another option to prevent the cognitive worsening may

be a cotreatment of BACE1 inhibitors together with activators of metabotropic glutamate receptor 1, which mitigated synaptic deficits upon BACE1 inhibition in mice [44]. An important task for the near future is to identify the molecular cause of the cognitive worsening, which is presumably caused by a too strong inhibition of cleavage of APP or another neuronal BACE1 substrate. Based on studies in adult mice, seizure protein 6, close homolog of L1, and an A $\beta$ -independent proteolytic fragment of APP, referred to as A $\eta\alpha$ , are likely candidates [45-47], but new BACE1 substrates continue to be identified [48-50], although their physiological BACE1-dependent functions have not yet been established. New BACE1 inhibitors are being developed and this does not only include small molecule drugs, but also BACE1-targeted antibodies [51].

$\gamma$ -Secretase mediates the final cleavages in the generation of A $\beta$  from APP and has thus been a major drug target since the discovery of PS as its catalytic component just over 20 years ago.  $\gamma$ -Secretase inhibitors (GSIs) have been abandoned due to severe side effects in clinical trials, which can at least in part be attributed to the inhibition of the generation of a key signaling molecule from Notch1, one of the most critical  $\gamma$ -secretase substrates. Instead of  $\gamma$ -secretase inhibition, modulation of the enzyme's processivity activity in the generation of A $\beta$ 42/43 into presumably harmless shorter A $\beta$  peptides such as A $\beta$ 38 by so-called  $\gamma$ -secretase modulators (GSMs) is considered a safe approach. Unlike GSIs, GSMs do not target the signaling cleavage functions of  $\gamma$ -secretase mediated by the intracellular domains of a number of other substrates besides Notch1. While the generation of these compounds has lacked behind the clinical development of BACE inhibitors, powerful GSMs with great promise for clinical trials are now emerging. One of these compounds combines high efficacy with an advanced safety profile in preclinical models [52]. Moreover, a novel GSM has been identified, which can also lower the pathogenic A $\beta$ 43 variant [53]. This and other GSMs are capable of breaking the resistance of a subset of PS FAD mutations in lowering A $\beta$ 42/43 informing the design of potential future GSM trials within FAD cohorts [54].

Another important step forward that may aid rational drug development are the recently obtained cryo-electron microscopy (cryo-EM) structures of  $\gamma$ -secretase in complex with APP or Notch1 substrates, different GSIs or a GSM at atomic resolution [55-57]. The GSM binding site was identified to locate on the extracellular side of PS in vicinity to its hydrophilic loop 1 and a nearby loop of the NCT ectodomain. This region had been implicated earlier as a site of GSM action [58,59]. While these structures might be a starting point for the development of improved APP-selective GSIs and could possibly lead to a revival of  $\gamma$ -secretase inhibition as a strategy for AD therapy, GSIs such as MRK-560 have been successfully repurposed in



preclinical models to interfere with enhanced Notch signaling in certain cancers such as T-ALL offering potential for further clinical development [60].

An additional drug target to modulate brain amyloidogenesis that has emerged in the past years is TREM2. Since its phagocytic activity towards A $\beta$  is mediated by the non-shed FL protein at the cell surface, inhibition of TREM2 ectodomain shedding by ADAMs might be beneficial in AD. Because ADAM10 has numerous neuronal ADAM10 substrates [10,61,62], the possibility of specifically inhibiting TREM2 shedding by targeting its ADAM10/17 cleavage site was tested with an antibody binding near to the cleavage site at histidine 157 (Fig. 3) [19-21]. As a proof of concept, one antibody, 4D9, reduces TREM2 shedding, increases the levels of FL TREM2 and thereby increases intracellular signaling leading to microglial activation including enhanced removal of A $\beta$  plaque halos in the brain of an AD mouse model [63]. Other antibodies raised against the TREM2 ectodomain that like 4D9 may all bind in the stalk region [3] showed similar microglial responses to AD pathology as 4D9 [64-66]. One of these antibodies, AL002c, that presumably like 4D9 blocks TREM2 shedding has already advanced into early phase clinical trials [65]. The relevance of TREM2 shedding for AD pathogenesis is further supported with the discovery of a LOF TREM2 mutation directly at the cleavage site (H157Y), which enhances shedding and reduces FL TREM2, causing reduced phagocytic activity and a genetic association to increased AD risk [20,21].

In addition to FL TREM2 signaling, which initiates a defense response to A $\beta$ , the interaction of the shed soluble TREM2 ectodomain (sTREM2) with A $\beta$  oligomers [67,68] may be protective against AD as well. As recently shown, sTREM2 cannot only bind A $\beta$  oligomers but also disaggregate them and block A $\beta$ -mediated neurotoxicity. The R47H AD risk variant is deficient in these activities and rather promotes A $\beta$  aggregation into larger aggregates [69]. Notably, antibodies against the TREM2 ectodomain may also bind to sTREM2 [63,66]. Moreover, sTREM2 may have also signaling functions [70]. Thus, the therapeutic implications of reducing sTREM2 by antibody-mediated inhibition of FL TREM2 shedding require more research.

## **Conclusion**

For many years, AD secretase research focused on the secretase substrate APP and its proteolytic conversion to A $\beta$ . Although clinical trials with BACE1- and  $\gamma$ -secretase-targeted drugs did not (yet) lead to approval of AD drugs, interest in the AD secretases has strongly increased over the past few years, in particular with the discovery that the secretase-mediated shedding of TREM2 in microglia can be pharmacologically inhibited to enhance TREM2 function, for example in microglial phagocytosis of A $\beta$ . First clinical trials targeting this

mechanism are ongoing or being prepared. Additionally, new, but safer clinical trials are now being considered with BACE1 inhibitors [6]. The detailed structural understanding of the mechanism of action of GSIs and GSMs may foster the development of new improved clinical candidates for AD trials. Another important area of research is the regulation of secretase activity, which may allow to identify new ways to therapeutically modulate secretase activity. Additionally, the detailed study of secretase substrates and functions beyond APP cleavage is intensively investigated, partly due to the desire to predict and understand mechanism-based side effects resulting from targeting secretase activity and partly driven by the basic research interest to understand the fundamental role of secretases in development, maintenance and function of the nervous system and other organs. Because secretases shed hundreds of single-span membrane proteins [8], it is likely that additional membrane proteins linked to AD will move to the center of secretase research in AD. This includes the LDL receptor-related protein 1 (LRP1), which is shed in the brain [48,71] and controls tau endocytosis and spreading between neurons [72], but potentially also the microglia-expressed AD risk genes CD33 and complement receptor 1 and the microglia-activation protein CLEC7a [2]. Taken together, we are in an exciting time for secretase research in AD which no longer focuses just on APP but also TREM2, and likely more AD-linked secretase substrates in the future.

### **Acknowledgement**

We are grateful for funding by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) through research consortium 263531414/FOR2290 and under Germany's Excellence Strategy within the framework of the Munich Cluster for Systems Neurology (EXC 2145 SyNergy – ID 390857198). We are also grateful to the Federal Ministry of Education and Research for grant JPND PMG-AD (JPco-fuND2 2019 Personalised Medicine for Neurodegenerative Diseases 01ED2002B) and CLINSPECT-M. We thank Kai Schlepckow and Christian Haass for helpful discussions and comments on the manuscript.

### **Declaration of interest**

H.S. and S.K.T. declare no conflict of interest. S.F.L. has research collaborations on BACE inhibitors with Novartis and Shionogi.

## References

1. Selkoe DJ, Hardy J: **The amyloid hypothesis of Alzheimer's disease at 25 years.** *EMBO molecular medicine* (2016) **8**(6):595-608.
2. Scheltens P, De Strooper B, Kivipelto M, Holstege H, Chetelat G, Teunissen CE, Cummings J, van der Flier WM: **Alzheimer's disease.** *Lancet* (2021) **397**(10284):1577-1590.
3. Lewcock JW, Schlepckow K, Di Paolo G, Tahirovic S, Monroe KM, Haass C: **Emerging Microglia Biology Defines Novel Therapeutic Approaches for Alzheimer's Disease.** *Neuron* (2020) **108**(5):801-821.
4. Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, David E, Baruch K, Lara-Astaiso D, Toth B, Itzkovitz S *et al*: **A Unique Microglia Type Associated with Restricting Development of Alzheimer's Disease.** *Cell* (2017) **169**(7):1276-1290 e1217.
5. Mazaheri F, Snaidero N, Kleinberger G, Madore C, Daria A, Werner G, Krasemann S, Capell A, Trumbach D, Wurst W, Brunner B *et al*: **TREM2 deficiency impairs chemotaxis and microglial responses to neuronal injury.** *EMBO reports* (2017) **18**(7):1186-1198.
6. McDade E, Voytyuk I, Aisen P, Bateman RJ, Carrillo MC, De Strooper B, Haass C, Reiman EM, Sperling R, Tariot PN, Yan R *et al*: **The case for low-level BACE1 inhibition for the prevention of Alzheimer disease.** *Nat Rev Neurol* (2021 in press).
7. Muller UC, Deller T, Korte M: **Not just amyloid: physiological functions of the amyloid precursor protein family.** *Nat Rev Neurosci* (2017) **18**(5):281-298.
8. Lichtenthaler SF, Lemberg MK, Fluhner R: **Proteolytic ectodomain shedding of membrane proteins in mammals-hardware, concepts, and recent developments.** *EMBO J* (2018) **37**(15).
9. Steiner H, Fukumori A, Tagami S, Okochi M: **Making the final cut: pathogenic amyloid-beta peptide generation by gamma-secretase.** *Cell Stress* (2018) **2**(11):292-310.
10. Saftig P, Lichtenthaler SF: **The alpha secretase ADAM10: A metalloprotease with multiple functions in the brain.** *Progress in neurobiology* (2015) **135**(1-20).
11. Endres K, Fahrenholz F, Lotz J, Hiemke C, Teipel S, Lieb K, Tuscher O, Fellgiebel A: **Increased CSF APPs-alpha levels in patients with Alzheimer disease treated with acitretin.** *Neurology* (2014) **83**(21):1930-1935.
12. Brummer T, Muller SA, Pan-Montojo F, Yoshida F, Fellgiebel A, Tomita T, Endres K, Lichtenthaler SF: **NrCAM is a marker for substrate-selective activation of ADAM10 in Alzheimer's disease.** *EMBO molecular medicine* (2019) **11**(4).
13. Hampel H, Vassar R, De Strooper B, Hardy J, Willem M, Singh N, Zhou J, Yan R, Vanmechelen E, De Vos A, Nistico R *et al*: **The beta-Secretase BACE1 in Alzheimer's Disease.** *Biol Psychiatry* (2021) **89**(8):745-756.

14. Alic I, Goh PA, Murray A, Portelius E, Gkanatsiou E, Gough G, Mok KY, Koschut D, Brunmeir R, Yeap YJ, O'Brien NL *et al*: **Patient-specific Alzheimer-like pathology in trisomy 21 cerebral organoids reveals BACE2 as a gene dose-sensitive AD suppressor in human brain.** *Mol Psychiatry* (2020).
15. Walter S, Jumpertz T, Huttenrauch M, Ogorek I, Gerber H, Storck SE, Zampar S, Dimitrov M, Lehmann S, Lepka K, Berndt C *et al*: **The metalloprotease ADAMTS4 generates N-truncated Abeta4-x species and marks oligodendrocytes as a source of amyloidogenic peptides in Alzheimer's disease.** *Acta Neuropathol* (2019) **137**(2):239-257.
16. Bien J, Jefferson T, Causevic M, Jumpertz T, Munter L, Multhaup G, Weggen S, Becker-Pauly C, Pietrzik CU: **The metalloprotease meprin beta generates amino terminal-truncated amyloid beta peptide species.** *J Biol Chem* (2012) **287**(40):33304-33313.
17. Liebsch F, Kulic L, Teunissen C, Shobo A, Ulku I, Engelschalt V, Hancock MA, van der Flier WM, Kunach P, Rosa-Neto P, Scheltens P *et al*: **Abeta34 is a BACE1-derived degradation intermediate associated with amyloid clearance and Alzheimer's disease progression.** *Nature communications* (2019) **10**(1):2240.
18. Kleinberger G, Yamanishi Y, Suarez-Calvet M, Czirr E, Lohmann E, Cuyvers E, Struyfs H, Pettkus N, Wenninger-Weinzierl A, Mazaheri F, Tahirovic S *et al*: **TREM2 mutations implicated in neurodegeneration impair cell surface transport and phagocytosis.** *Sci Transl Med* (2014) **6**(243):243ra286.
19. Feuerbach D, Schindler P, Barske C, Joller S, Beng-Louka E, Worringer KA, Kommineni S, Kaykas A, Ho DJ, Ye C, Welzenbach K *et al*: **ADAM17 is the main sheddase for the generation of human triggering receptor expressed in myeloid cells (hTREM2) ectodomain and cleaves TREM2 after Histidine 157.** *Neuroscience letters* (2017) **660**(109-114).
20. Schlepckow K, Kleinberger G, Fukumori A, Feederle R, Lichtenthaler SF, Steiner H, Haass C: **An Alzheimer-associated TREM2 variant occurs at the ADAM cleavage site and affects shedding and phagocytic function.** *EMBO molecular medicine* (2017) **9**(10):1356-1365.
21. Thornton P, Sevalle J, Deery MJ, Fraser G, Zhou Y, Stahl S, Franssen EH, Dodd RB, Qamar S, Gomez Perez-Nievas B, Nicol LS *et al*: **TREM2 shedding by cleavage at the H157-S158 bond is accelerated for the Alzheimer's disease-associated H157Y variant.** *EMBO molecular medicine* (2017) **9**(10):1366-1378.
22. Berner DK, Wessolowski L, Armbrust F, Schneppenheim J, Schlepckow K, Koudelka T, Scharfenberg F, Lucius R, Tholey A, Kleinberger G, Haass C *et al*: **Meprin beta cleaves TREM2 and controls its phagocytic activity on macrophages.** *FASEB J* (2020) **34**(5):6675-6687.
23. Song W, Hooli B, Mullin K, Jin SC, Cella M, Ulland TK, Wang Y, Tanzi RE, Colonna M: **Alzheimer's disease-associated TREM2 variants exhibit either decreased or increased ligand-dependent activation.** *Alzheimers Dement* (2017) **13**(4):381-387.

24. Wunderlich P, Glebov K, Kemmerling N, Tien NT, Neumann H, Walter J: **Sequential proteolytic processing of the triggering receptor expressed on myeloid cells-2 (TREM2) protein by ectodomain shedding and gamma-secretase-dependent intramembranous cleavage.** *J Biol Chem* (2013) **288**(46):33027-33036.
25. Steiner A, Schlepckow K, Brunner B, Steiner H, Haass C, Hagn F: **gamma-Secretase cleavage of the Alzheimer risk factor TREM2 is determined by its intrinsic structural dynamics.** *EMBO J* (2020) **39**(20):e104247.
26. Jiang T, Tan L, Chen Q, Tan MS, Zhou JS, Zhu XC, Lu H, Wang HF, Zhang YD, Yu JT: **A rare coding variant in TREM2 increases risk for Alzheimer's disease in Han Chinese.** *Neurobiology of aging* (2016) **42**(217 e211-213).
27. Chavez-Gutierrez L, Szaruga M: **Mechanisms of neurodegeneration - Insights from familial Alzheimer's disease.** *Semin Cell Dev Biol* (2020) **105**(75-85).
28. Pagnon de la Vega M, Giedraitis V, Michno W, Kilander L, Güner G, Zielinski M, Löwenmark M, Brundin R, Danfors T, Söderberg L, Alafuzoff I *et al*: **The Uppsala APP deletion causes early onset autosomal dominant Alzheimer's disease by altering APP processing and increasing amyloid- $\beta$  fibril formation.** *Science translational medicine* (2021 in press).
29. Tian Y, Bassit B, Chau D, Li YM: **An APP inhibitory domain containing the Flemish mutation residue modulates gamma-secretase activity for Abeta production.** *Nat Struct Mol Biol* (2010) **17**(2):151-158.
30. Aguero P, Sainz MJ, Garcia-Ayllon MS, Saez-Valero J, Tellez R, Guerrero-Lopez R, Perez-Perez J, Jimenez-Escrig A, Gomez-Tortosa E: **alpha-Secretase nonsense mutation (ADAM10 Tyr167\*) in familial Alzheimer's disease.** *Alzheimers Res Ther* (2020) **12**(1):139.
31. Kunkle BW, Grenier-Boley B, Sims R, Bis JC, Damotte V, Naj AC, Boland A, Vronskaya M, van der Lee SJ, Amlie-Wolf A, Bellenguez C *et al*: **Genetic meta-analysis of diagnosed Alzheimer's disease identifies new risk loci and implicates Abeta, tau, immunity and lipid processing.** *Nat Genet* (2019) **51**(3):414-430.
32. Jansen IE, Savage JE, Watanabe K, Bryois J, Williams DM, Steinberg S, Sealock J, Karlsson IK, Hagg S, Athanasiu L, Voyle N *et al*: **Genome-wide meta-analysis identifies new loci and functional pathways influencing Alzheimer's disease risk.** *Nat Genet* (2019) **51**(3):404-413.
33. Hartl D, May P, Gu W, Mayhaus M, Pichler S, Spaniol C, Glaab E, Bobbili DR, Antony P, Koegelsberger S, Kurz A *et al*: **A rare loss-of-function variant of ADAM17 is associated with late-onset familial Alzheimer disease.** *Mol Psychiatry* (2020) **25**(3):629-639.
34. Torres-Acosta N, O'Keefe JH, O'Keefe EL, Isaacson R, Small G: **Therapeutic Potential of TNF-alpha Inhibition for Alzheimer's Disease Prevention.** *J Alzheimers Dis* (2020) **78**(2):619-626.

35. Deming Y, Filipello F, Cignarella F, Cantoni C, Hsu S, Mikesell R, Li Z, Del-Aguila JL, Dube U, Farias FG, Bradley J *et al*: **The MS4A gene cluster is a key modulator of soluble TREM2 and Alzheimer's disease risk.** *Sci Transl Med* (2019) **11**(505).
36. Marcello E, Borroni B, Pelucchi S, Gardoni F, Di Luca M: **ADAM10 as a therapeutic target for brain diseases: from developmental disorders to Alzheimer's disease.** *Expert Opin Ther Targets* (2017) **21**(11):1017-1026.
37. Esteve P, Rueda-Carrasco J, Ines Mateo M, Martin-Bermejo MJ, Draffin J, Pereyra G, Sandonis A, Crespo I, Moreno I, Aso E, Garcia-Esparcia P *et al*: **Elevated levels of Secreted-Frizzled-Related-Protein 1 contribute to Alzheimer's disease pathogenesis.** *Nat Neurosci* (2019) **22**(8):1258-1268.
38. Nakamura M, Li Y, Choi BR, Matas-Rico E, Troncoso J, Takahashi C, Sockanathan S: **GDE2-RECK controls ADAM10 alpha-secretase-mediated cleavage of amyloid precursor protein.** *Sci Transl Med* (2021) **13**(585).
39. Suh J, Romano DM, Nitschke L, Herrick SP, DiMarzio BA, Dzhala V, Bae JS, Oram MK, Zheng Y, Hooli B, Mullin K *et al*: **Loss of Ataxin-1 Potentiates Alzheimer's Pathogenesis by Elevating Cerebral BACE1 Transcription.** *Cell* (2019) **178**(5):1159-1175 e1117.
40. Bahn G, Park JS, Yun UJ, Lee YJ, Choi Y, Park JS, Baek SH, Choi BY, Cho YS, Kim HK, Han J *et al*: **NRF2/ARE pathway negatively regulates BACE1 expression and ameliorates cognitive deficits in mouse Alzheimer's models.** *Proc Natl Acad Sci U S A* (2019) **116**(25):12516-12523.
41. Lomoio S, Willen R, Kim W, Ho KZ, Robinson EK, Prokopenko D, Kennedy ME, Tanzi RE, Tesco G: **Gga3 deletion and a GGA3 rare variant associated with late onset Alzheimer's disease trigger BACE1 accumulation in axonal swellings.** *Sci Transl Med* (2020) **12**(570).
42. Egan MF, Kost J, Voss T, Mukai Y, Aisen PS, Cummings JL, Tariot PN, Vellas B, van Dyck CH, Boada M, Zhang Y *et al*: **Randomized Trial of Verubecestat for Prodromal Alzheimer's Disease.** *N Engl J Med* (2019) **380**(15):1408-1420.
43. Henley D, Raghavan N, Sperling R, Aisen P, Raman R, Romano G: **Preliminary Results of a Trial of Atabecestat in Preclinical Alzheimer's Disease.** *N Engl J Med* (2019) **380**(15):1483-1485.
44. Das B, Singh N, Yao AY, Zhou J, He W, Hu X, Yan R: **BACE1 controls synaptic function through modulating release of synaptic vesicles.** *Mol Psychiatry* (2021).
45. Ou-Yang MH, Kurz JE, Nomura T, Popovic J, Rajapaksha TW, Dong H, Contractor A, Chetkovich DM, Tourtellotte WG, Vassar R: **Axonal organization defects in the hippocampus of adult conditional BACE1 knockout mice.** *Sci Transl Med* (2018) **10**(459).
46. Zhu K, Xiang X, Filser S, Marinkovic P, Dorostkar MM, Crux S, Neumann U, Shimshek DR, Rammes G, Haass C, Lichtenthaler SF *et al*: **Beta-Site Amyloid Precursor Protein**

- Cleaving Enzyme 1 Inhibition Impairs Synaptic Plasticity via Seizure Protein 6.** *Biol Psychiatry* (2018) **83**(5):428-437.
47. Willem M, Tahirovic S, Busche MA, Ovsepian SV, Chafai M, Kootar S, Hornburg D, Evans LD, Moore S, Daria A, Hampel H *et al*: **eta-Secretase processing of APP inhibits neuronal activity in the hippocampus.** *Nature* (2015) **526**(7573):443-447.
  48. Tushaus J, Muller SA, Kataka ES, Zauha J, Sebastian Monasor L, Su M, Guner G, Jocher G, Tahirovic S, Frishman D, Simons M *et al*: **An optimized quantitative proteomics method establishes the cell type-resolved mouse brain secretome.** *EMBO J* (2020) **39**(20):e105693.
  49. Rudan Njavro J, Klotz J, Dislich B, Wanngren J, Shmueli MD, Herber J, Kuhn PH, Kumar R, Koeglsperger T, Conrad M, Wurst W *et al*: **Mouse brain proteomics establishes MDGA1 and CACHD1 as in vivo substrates of the Alzheimer protease BACE1.** *FASEB J* (2020) **34**(2):2465-2482.
  50. Kim W, Watanabe H, Lomoio S, Tesco G: **Spatiotemporal processing of neural cell adhesion molecules 1 and 2 by BACE1 in vivo.** *J Biol Chem* (2021) **296**(100372).
  51. Kariolis MS, Wells RC, Getz JA, Kwan W, Mahon CS, Tong R, Kim DJ, Srivastava A, Bedard C, Henne KR, Giese T *et al*: **Brain delivery of therapeutic proteins using an Fc fragment blood-brain barrier transport vehicle in mice and monkeys.** *Sci Transl Med* (2020) **12**(545).
  52. Rynearson KD, Ponnusamy M, Prikhodko O, Xie Y, Zhang C, Nguyen P, Hug B, Sawa M, Becker A, Spencer B, Florio J *et al*: **Preclinical validation of a potent gamma-secretase modulator for Alzheimer's disease prevention.** *J Exp Med* (2021) **218**(4).
  53. Trambauer J, Rodriguez Sarmiento RM, Fukumori A, Feederle R, Baumann K, Steiner H: **Abeta43-producing PS1 FAD mutants cause altered substrate interactions and respond to gamma-secretase modulation.** *EMBO reports* (2020) **21**(1):e47996.
  54. Trambauer J, Fukumori A, Steiner H: **Pathogenic Abeta generation in familial Alzheimer's disease: novel mechanistic insights and therapeutic implications.** *Curr Opin Neurobiol* (2020) **61**(73-81).
  55. Yang G, Zhou R, Guo X, Yan C, Lei J, Shi Y: **Structural basis of gamma-secretase inhibition and modulation by small molecule drugs.** *Cell* (2021) **184**(2):521-533 e514.
  56. Yang G, Zhou R, Zhou Q, Guo X, Yan C, Ke M, Lei J, Shi Y: **Structural basis of Notch recognition by human gamma-secretase.** *Nature* (2019) **565**(7738):192-197.
  57. Zhou R, Yang G, Guo X, Zhou Q, Lei J, Shi Y: **Recognition of the amyloid precursor protein by human gamma-secretase.** *Science* (2019) **363**(6428).
  58. Takeo K, Tanimura S, Shinoda T, Osawa S, Zahariev IK, Takegami N, Ishizuka-Katsura Y, Shinya N, Takagi-Niidome S, Tominaga A, Ohsawa N *et al*: **Allosteric regulation of gamma-secretase activity by a phenylimidazole-type gamma-secretase modulator.** *Proc Natl Acad Sci U S A* (2014) **111**(29):10544-10549.

59. Petit D, Hitzemberger M, Lismont S, Zoltowska KM, Ryan NS, Mercken M, Bischoff F, Zacharias M, Chavez-Gutierrez L: **Extracellular interface between APP and Nicastrin regulates Abeta length and response to gamma-secretase modulators.** *EMBO J* (2019) **38**(12).
60. Habets RA, de Bock CE, Serneels L, Lodewijckx I, Verbeke D, Nittner D, Narlawar R, Demeyer S, Dooley J, Liston A, Taghon T *et al*: **Safe targeting of T cell acute lymphoblastic leukemia by pathology-specific NOTCH inhibition.** *Sci Transl Med* (2019) **11**(494).
61. Kuhn PH, Colombo AV, Schusser B, Dreymueller D, Wetzel S, Schepers U, Herber J, Ludwig A, Kremmer E, Montag D, Muller U *et al*: **Systematic substrate identification indicates a central role for the metalloprotease ADAM10 in axon targeting and synapse function.** *eLife* (2016) **5**(
62. Gunner G, Cheadle L, Johnson KM, Ayata P, Badimon A, Mondo E, Nagy MA, Liu L, Bemiller SM, Kim KW, Lira SA *et al*: **Sensory lesioning induces microglial synapse elimination via ADAM10 and fractalkine signaling.** *Nat Neurosci* (2019) **22**(7):1075-1088.
63. Schlepckow K, Monroe KM, Kleinberger G, Cantuti-Castelvetri L, Parhizkar S, Xia D, Willem M, Werner G, Pettkus N, Brunner B, Sulzen A *et al*: **Enhancing protective microglial activities with a dual function TREM2 antibody to the stalk region.** *EMBO molecular medicine* (2020) **12**(4):e11227.
64. Price BR, Sudduth TL, Weekman EM, Johnson S, Hawthorne D, Woolums A, Wilcock DM: **Therapeutic Trem2 activation ameliorates amyloid-beta deposition and improves cognition in the 5XFAD model of amyloid deposition.** *J Neuroinflammation* (2020) **17**(1):238.
65. Wang S, Mustafa M, Yuede CM, Salazar SV, Kong P, Long H, Ward M, Siddiqui O, Paul R, Gilfillan S, Ibrahim A *et al*: **Anti-human TREM2 induces microglia proliferation and reduces pathology in an Alzheimer's disease model.** *J Exp Med* (2020) **217**(9).
66. Fassler M, Rappaport MS, Cuno CB, George J: **Engagement of TREM2 by a novel monoclonal antibody induces activation of microglia and improves cognitive function in Alzheimer's disease models.** *J Neuroinflammation* (2021) **18**(1):19.
67. Lessard CB, Malnik SL, Zhou Y, Ladd TB, Cruz PE, Ran Y, Mahan TE, Chakrabaty P, Holtzman DM, Ulrich JD, Colonna M *et al*: **High-affinity interactions and signal transduction between Abeta oligomers and TREM2.** *EMBO molecular medicine* (2018) **10**(11).
68. Zhong L, Wang Z, Wang D, Wang Z, Martens YA, Wu L, Xu Y, Wang K, Li J, Huang R, Can D *et al*: **Amyloid-beta modulates microglial responses by binding to the triggering receptor expressed on myeloid cells 2 (TREM2).** *Molecular neurodegeneration* (2018) **13**(1):15.



69. Vilalta A, Zhou Y, Sevalle J, Griffin JK, Satoh K, Allendorf DH, De S, Puigdellivol M, Bruzas A, Burguillos MA, Dodd RB *et al*: **Wild-type sTREM2 blocks Abeta aggregation and neurotoxicity, but the Alzheimer's R47H mutant increases Abeta aggregation.** *J Biol Chem* (2021) **296**(100631).
70. Zhong L, Chen XF, Wang T, Wang Z, Liao C, Wang Z, Huang R, Wang D, Li X, Wu L, Jia L *et al*: **Soluble TREM2 induces inflammatory responses and enhances microglial survival.** *J Exp Med* (2017) **214**(3):597-607.
71. Liu Q, Zhang J, Tran H, Verbeek MM, Reiss K, Estus S, Bu G: **LRP1 shedding in human brain: roles of ADAM10 and ADAM17.** *Molecular neurodegeneration* (2009) **4**(17).
72. Rauch JN, Luna G, Guzman E, Audouard M, Challis C, Sibih YE, Leshuk C, Hernandez I, Wegmann S, Hyman BT, Gradinaru V *et al*: **LRP1 is a master regulator of tau uptake and spread.** *Nature* (2020) **580**(7803):381-385.

### **Selection of candidate highlight papers 2019, 2020, 2021**

#### **Papers of special interest (\*):**

- Bahn PNAS 2019: A signaling pathway involving the transcription factor Nrf2 is shown to suppress BACE1 transcription and ameliorate cognitive deficits in an Alzheimer's disease mouse model.
- Brummer EMBO Mol Med 2019: Using proteomics and CSF samples from a phase 2a clinical trial, this study reports that activation of the  $\alpha$ -secretase ADAM10 may be feasible in a substrate-specific manner.
- Das Mol Psychiatry 2021: This study reports that positive allosteric modulators of metabotropic glutamate receptor 1 may alleviate cognitive worsening induced by clinically tested BACE1 inhibitors.
- Deming Sci Transl Med 2019: Using genetics and functional analysis, this study identified the MS4A gene cluster as a modifier of TREM2 shedding in Alzheimer's disease.
- Esteve Nat Neurosci 2019: This study demonstrates that the protein Sfrp1 increases A $\beta$  generation in mice through inhibition of ADAM10 and that Sfrp1 protein abundance is enhanced in brains from Alzheimer patients.
- Nakamura Sci Transl Med 2019: Using mice, this study unravels a neuronal signaling pathway that controls ADAM10 activity and A $\beta$  generation in mice.
- Pagnon de la Vega, Sci Transl Med 2021 in press: This study identifies the first deletion within the A $\beta$  sequence of APP that causes dominantly inherited Alzheimer's disease.

Mechanistically, the mutation is shown to shift APP processing from  $\alpha$ - to  $\beta$ -secretase and to modify A $\beta$  aggregation.

- Petit EMBO J 2020: This paper shows that a NCT loop region located in the vicinity of the hydrophilic loop 1 of PS contributes to the modulatory activity of GSMs on APP processing.
- Ryneerson J Exp Med 2021: The authors report a novel GSM which shows promising safety and efficacy properties in preclinical models including non-human primates and thus has potential to advance to a safe clinical candidate for the treatment of AD.
- Suh Cell 2019: In mice loss of ataxin-1 enhances Alzheimer's disease pathogenesis through increasing transcription of the  $\beta$ -secretase BACE1.
- Trambauer EMBO Rep. 2020: This paper reports on the identification of a novel GSM which can also lower the pathogenic A $\beta$ 43 species that is particularly produced in higher amounts by a subset of PS FAD mutants.
- Vialta J Biol Chem 2021: This study describes an anti-aggregatory function of the shed sTREM2 for A $\beta$  oligomers, which is compromised in TREM2 coding variants associated with increased AD risk.

#### **Papers of outstanding interest (\*\*):**

- Price et al. J Neuroinflamm 2020: This paper shows that an agonistic antibody against the TREM2 ectodomain elicits microglial activation resulting in reduced amyloidosis and improved cognition in an AD mouse model.
- Schlepckow EMBO Mol Med 2020: This work shows that an agonistic antibody 4D9 against the TREM2 extracellular stalk region blocks ectodomain shedding and triggers microglial defense responses to amyloid pathology in an in vivo model of AD.
- Wang J Exp Med 2020: This study shows that the agonistic antibody AL002c against the ectodomain of human TREM2 induces microglial defense responses against amyloidosis that reduces the formation of neuritic plaque pathology in an AD mouse model. The authors further show that AL002c is safe and well-tolerated in a first phase I clinical trial.
- Fassler J Neuroinflamm 2021 The authors report microglial defense responses triggered by an antibody against the TREM2 ectodomain including improved cognition in an AD mouse model.
- Yang Cell 2021: By identifying the  $\gamma$ -secretase binding sites of GSIs of different functional types as well as that of a GSM at atomic resolution, the authors provide molecular bases for the mode of action of these compounds. The structures reveal that GSIs occupy the region of the APP and Notch1 substrate  $\beta$ -strands proving an

explanation of their inhibition mechanism, while the GSM binds at an extracellular PS/NCT interface.

- Zhou Science 2019: Groundbreaking structural analysis of a  $\gamma$ -secretase enzyme-substrate complex showing that the APP substrate forms a hybrid  $\beta$ -sheet with the enzyme whereby the initial substrate cleavage sites are unfolded. Basically identical substrate-binding features were published in parallel by the same group for Notch1 [54].

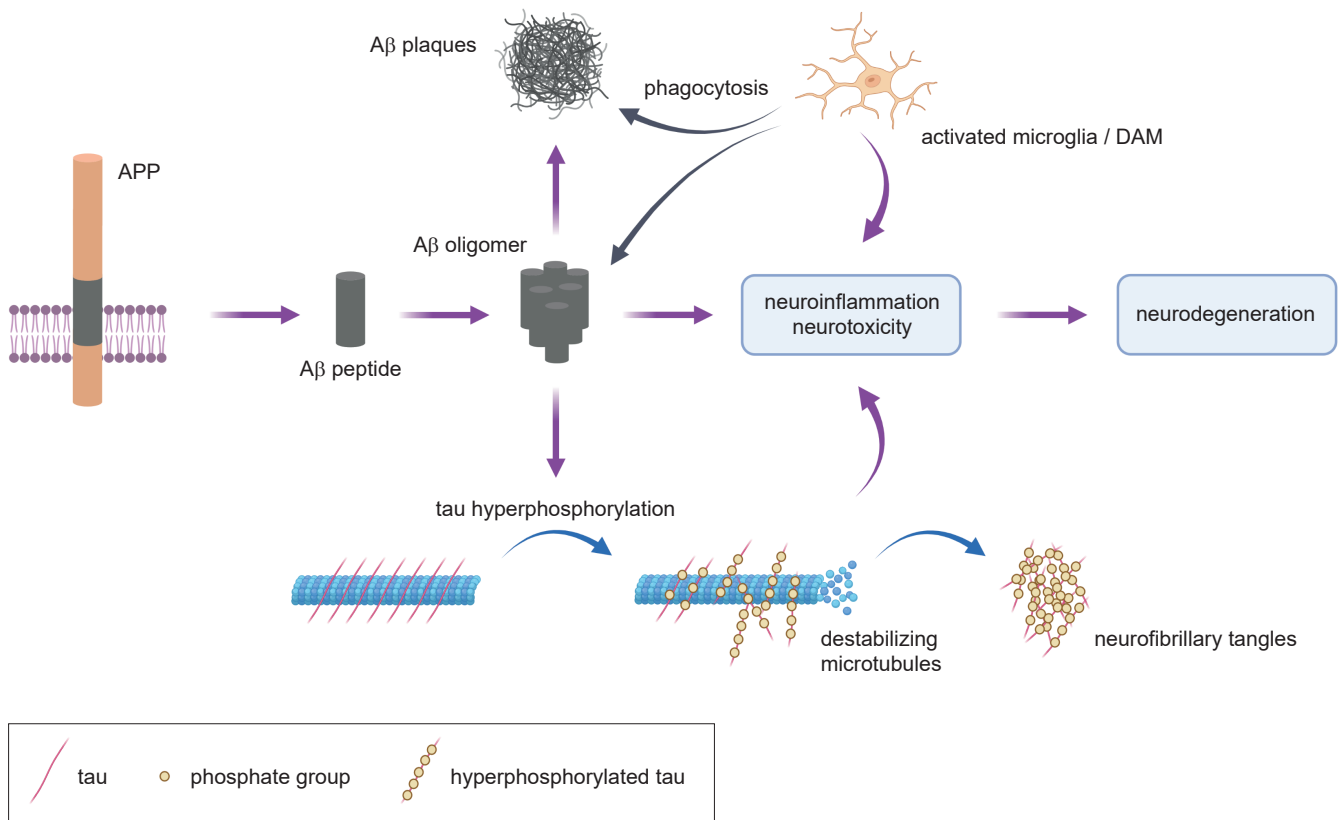
## Figure Legends

**Fig. 1: Amyloid cascade hypothesis of Alzheimer's disease.** The amyloid cascade is a simplified model to illustrate key pathological changes during AD pathogenesis.  $A\beta$  is a proteolytic cleavage product of APP. Oligomeric  $A\beta$  aggregates induce neurotoxicity and chronic neuroinflammation and lead to aggregation of hyperphosphorylated forms of the microtubule-associated protein tau and finally to neurodegeneration and the onset of clinical AD symptoms.  $A\beta$  and tau aggregates grow over time, are used for the clinical diagnosis of AD upon autopsy and are then referred to as amyloid plaques and neurofibrillary tangles.  $A\beta$  has a short half-life and can be removed by microglia, which, in the course of AD pathology, switch from a homeostatic to an activated state. While  $A\beta$  phagocytosis by microglia reduces amyloid load, chronically activated microglia can also enhance the neuroinflammatory process.

**Fig. 2: Proteolytic processing of APP and its role in AD. A-D.** In the amyloidogenic pathway, APP processing by  $\beta$ - and  $\gamma$ -secretase generates  $A\beta$  (A) (marked in grey).  $\beta$ -Secretase activity is mediated by BACE1 and to a lower extent by meprin  $\beta$ , ADAMTS4 and potentially other proteases (B).  $\gamma$ -Secretase is a hetero-tetrameric protease complex consisting of the subunits shown in (C).  $\gamma$ -Secretase has different cleavage sites – predominantly after amino acid 40, but also 42 of  $A\beta$  – so that  $A\beta$  presents as a heterogeneous peptide mixture including  $A\beta_{40}$  as most abundant peptide.  $A\beta_{42}$  is more hydrophobic, is the major constituent of amyloid plaques and is generated to a larger extent when  $\gamma$ -secretase is mutated in familial forms of AD (D). In the anti-amyloidogenic pathway,  $\alpha$ -secretase cleavage – mediated by ADAM10 and related proteases – prevents  $A\beta$  generation. **E.** Familial AD mutations around the cleavage site of  $\alpha$ -secretase may lower the anti-amyloidogenic APP processing and/or increase the aggregation propensity of  $A\beta$ . In contrast, the  $A\beta$ -lowering Icelandic mutation (blue) protects against AD. **F.** AEP/legumain acts as  $\delta$ -secretase and may enhance  $\beta$ -secretase cleavage and  $A\beta$  generation. **G.** Processing of APP by  $\eta$ -secretase, followed by  $\alpha$ - or  $\beta$ -secretase, generates the  $A\eta$  peptide ( $A\eta\alpha$ ,  $A\eta\beta$ ), which is distinct from  $A\beta$ , but may exert neurotoxic functions in its  $A\eta\alpha$  variant form.  $\eta$ -Secretase activity may be mediated by MT5-MMP, but also by other, as yet unidentified proteases. For more details on APP processing, see text.

**Fig. 3: Proteolytic processing of TREM2 and its role in  $A\beta$  phagocytosis. A.** TREM2 processing pathways. TREM2 is cleaved in the stalk region of its ectodomain by ADAM10 and/or ADAM17. Alternatively, meprin  $\beta$  cleaves within the ectodomain more N-terminally. Both cleavages leave C-terminal fragments in the membrane which are subsequently cleaved by  $\gamma$ -secretase. The cleavages terminate TREM2 signaling, which is mediated by FL TREM2 in complex with DAP12 and leads to activation of homeostatic microglia thereby triggering protective activities including  $A\beta$  phagocytosis. The asterisk in the membrane-proximal stalk

region denotes the ADAM protease cleavage site at amino acid 157. A point mutation at this site, H157Y, which causes AD, enhances ADAM protease cleavage of TREM2, presumably reducing the protective TREM2 functions. **B.** Therapeutic targeting of TREM2 shedding. Agonistic antibodies such as 4D9 that prevent ectodomain shedding likely cause receptor cross-linking and trigger defense mechanisms to AD pathology from FL TREM2-mediated signaling.



**Fig. 1**

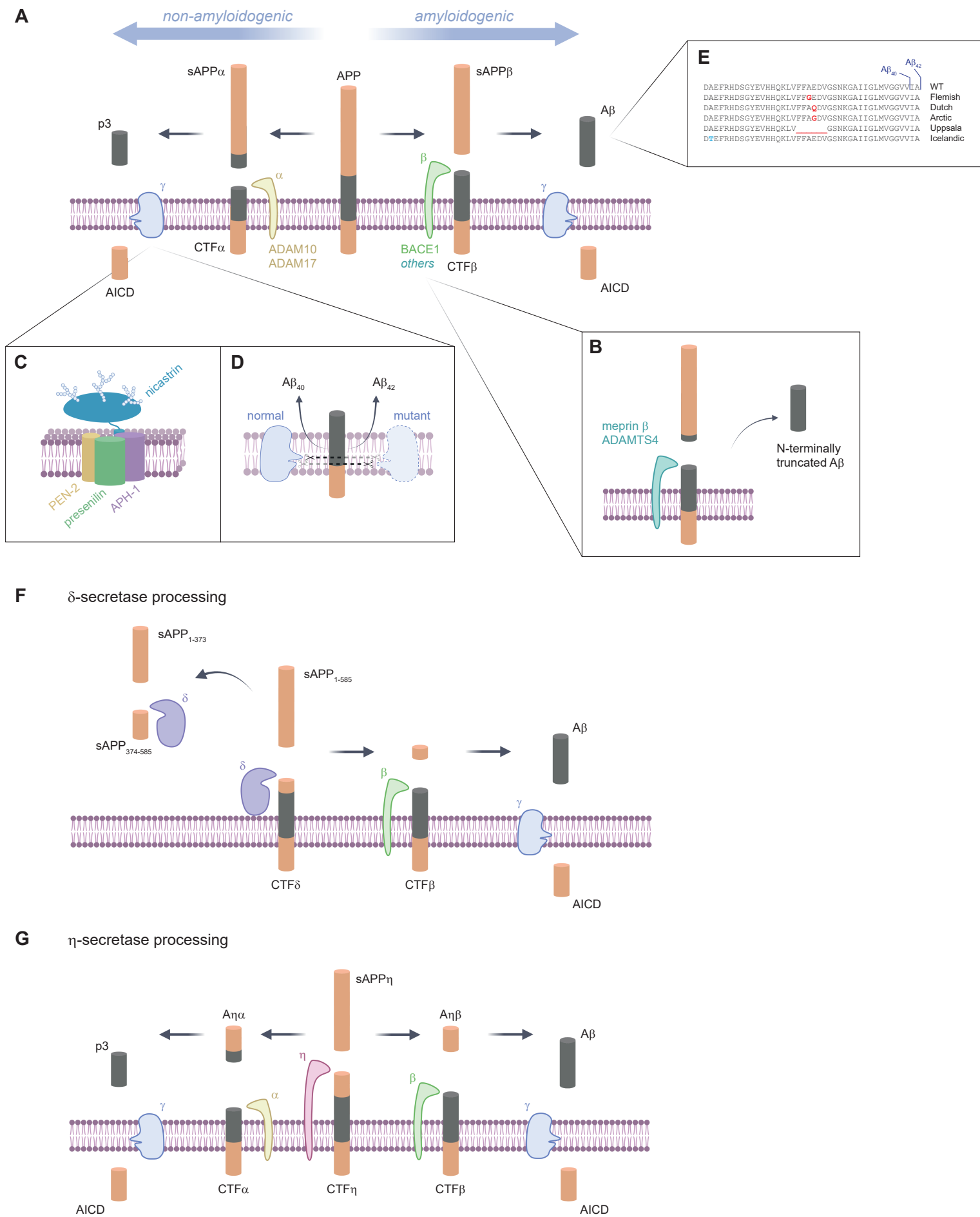
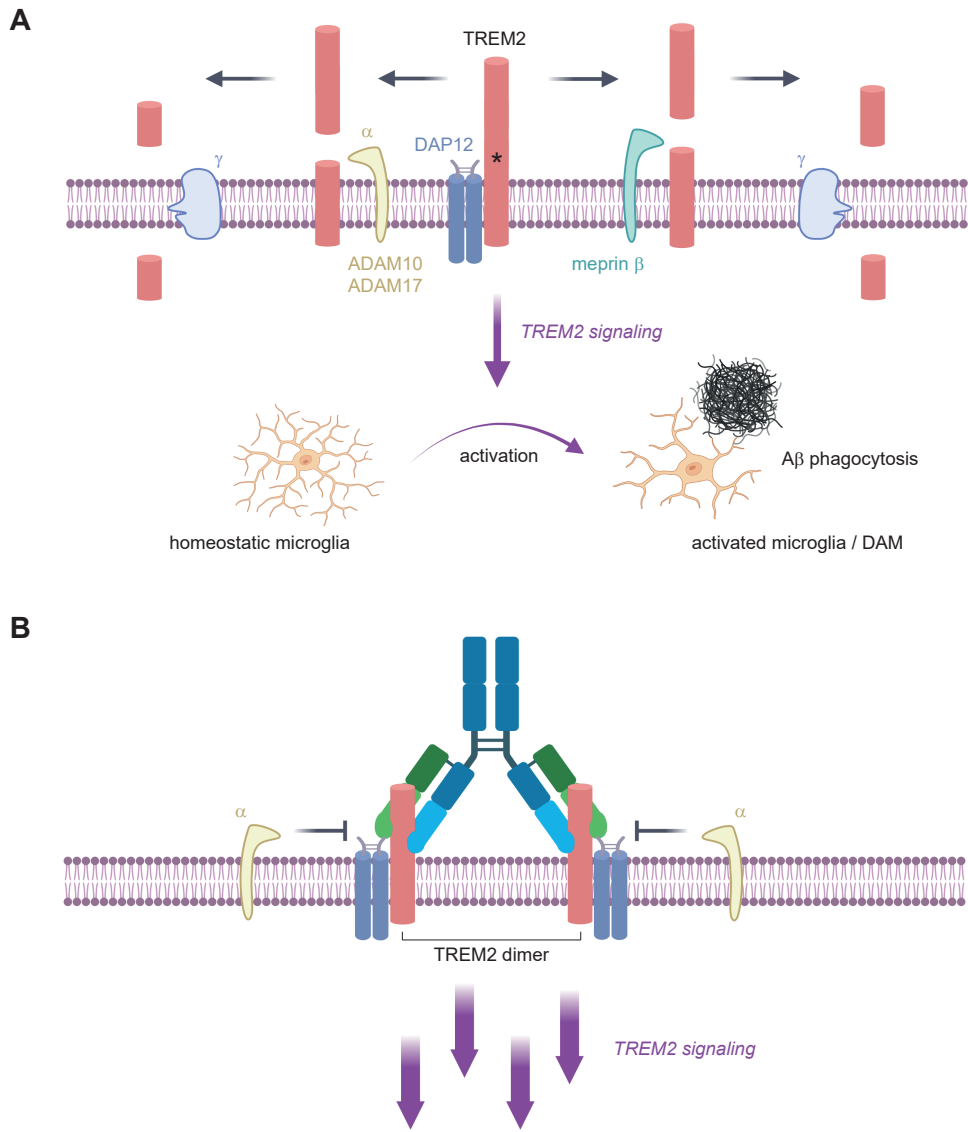


Fig. 2



**Fig. 3**