

Perspective

The evolving landscape of Alzheimer's disease therapy: From A β to tauJean-Philippe Courade,^{1,13} Henrik Zetterberg,^{2,3,4,5,6,7,8} Günter U. Höglunger,^{9,10,11} and Ilse Dewachter^{12,13,*}¹Discoveric bio alpha, Bahnhofstrasse 1, 8808 Pfäffikon, Switzerland²Department of Psychiatry and Neurochemistry, Institute of Neuroscience and Physiology, the Sahlgrenska Academy at the University of Gothenburg, Mölndal, Sweden³Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden⁴Department of Neurodegenerative Disease, UCL Institute of Neurology, Queen Square, London, UK⁵UK Dementia Research Institute at UCL, London, UK⁶Hong Kong Center for Neurodegenerative Diseases, InnoHK, Hong Kong, China⁷Wisconsin Alzheimer's Disease Research Center, University of Wisconsin School of Medicine and Public Health, University of Wisconsin-Madison, Madison, WI, USA⁸Centre for Brain Research, Indian Institute of Science, Bangalore, India⁹Department of Neurology, LMU University Hospital, Ludwig-Maximilians-Universität (LMU) München, Munich, Germany¹⁰German Center for Neurodegenerative Diseases (DZNE), Munich, Germany¹¹Munich Cluster for Systems Neurology (SyNergy), Munich, Germany¹²Department of Neurosciences, Biomedical Research Institute BIOMED, Hasselt University, Hasselt, Belgium¹³These authors contributed equally*Correspondence: ilse.dewachter@uhasselt.be<https://doi.org/10.1016/j.cell.2025.11.033>

SUMMARY

A marked evolution in Alzheimer's disease (AD) therapy research is ongoing. In this perspective, we highlight emerging outcomes of tau-targeting approaches with disease-modifying potential evidenced by PET-based slowing of tau accumulation and early signs of cognitive benefit. We outline how decades of iterative amyloid β (A β)-trial refinement leading to the recent successes of approved anti-A β therapies have set the stage for accelerated optimization of next-generation trials. We summarize key learnings from first-generation tau immunotherapies and how these paved the way for early achievements in tau trials, while many challenges remain. Finally, we discuss the back-translation of clinical outcomes into fundamental insights on human tau pathobiology, and we outline challenges and future directions for AD therapy development including combination therapy and targets beyond A β /tau. Together, this provides a framework for next-generation AD and tau-therapy development toward increasingly efficient disease-halting interventions.

INTRODUCTION

In this perspective, we first outline recent breakthroughs that opened a new era in Alzheimer's disease (AD) research and treatment. These include the recent US Food and Drug Administration (FDA) approvals of anti-amyloid β (A β) therapies—lecanemab and donanemab—resulting from decades of setbacks yielding incremental success in A β trial optimization. Emerging outcomes of tau trials are fast following the A β successes, showing slower tau accumulation (measured by tau PET [positron emission tomography]) and early signs of cognitive benefit. While primary endpoints were not met, and corroboration in multiple, large future trials will be critical, these outcomes represent first proof of concept in humans of biological disease halting and present an important step forward for tau therapeutic agents. We here provide an integrated analysis of the tau trial outcomes alongside lessons learned from A β trials, which yield a roadmap for therapy optimization. We also discuss clinically validated insights in light of fundamental tau pathobiology and remaining challenges. Look-

ing forward, we discuss opportunities for A β -tau combination therapies, as well as therapeutic strategies targeting other factors. Together, this provides a roadmap to guide accelerated, next-generation AD and tau-therapy development.

EVOLUTIONS IN AD THERAPY: FROM A β TO TAU

AD and other tauopathies

AD is the most frequent tauopathy and prevalent dementia, with an unmet medical need for therapy. Globally, AD accounts for approximately 32 out of 57 million dementia cases,¹⁻³ which are expected to climb to 153 million by 2050, posing a major health and socioeconomic challenge.² The AD brain is characterized by the presence of A β pathology (A), tau pathology (T), and neurodegeneration (N), as pathological hallmarks⁴⁻⁹ that serve as a clinical-biological framework^{10,11} of AD (Figure 1).

Both A β and tau represent key therapeutic targets. Accumulating evidence supports an initiatory role for A β in AD, while tau is proposed to have an executive role in the



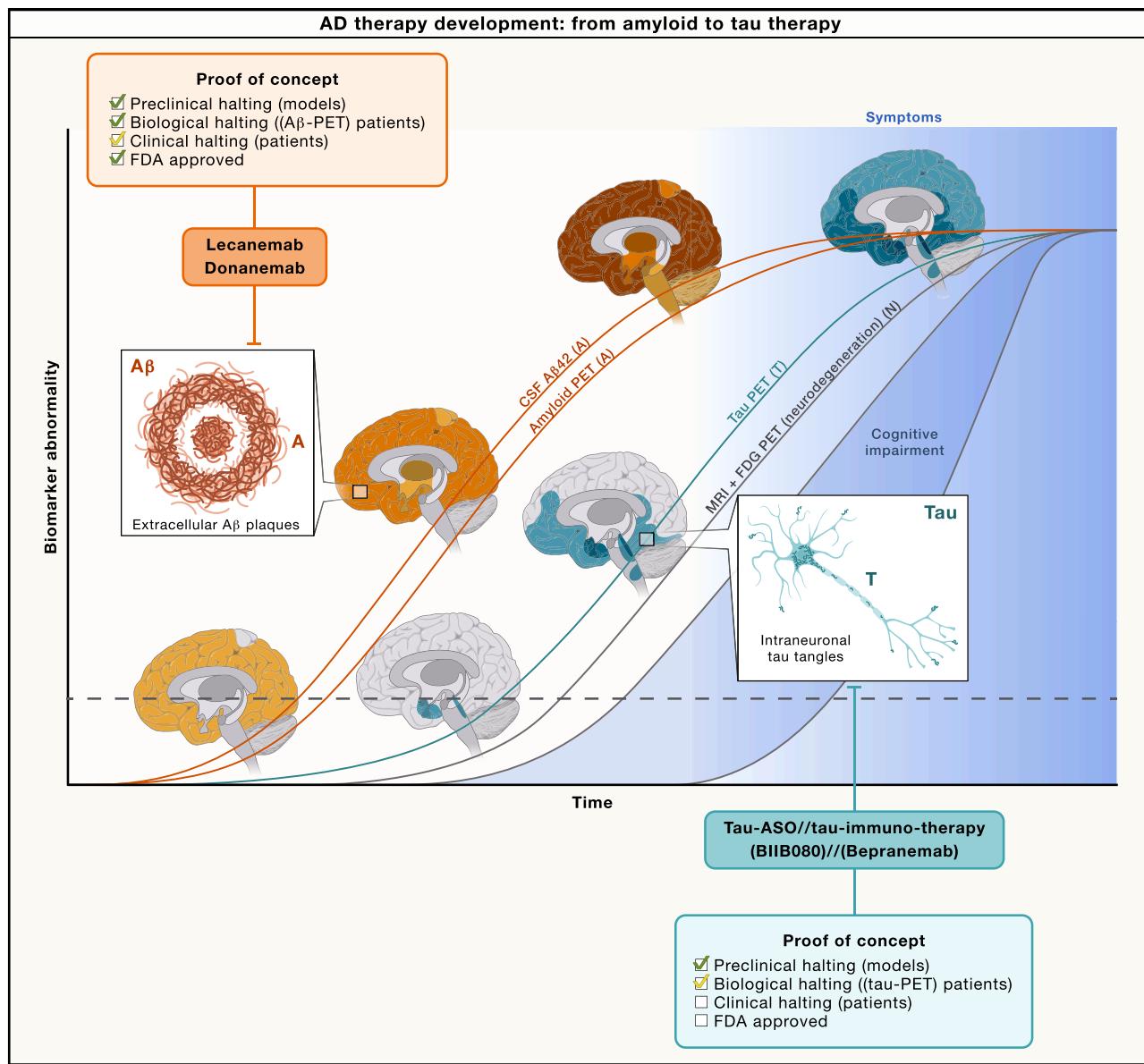


Figure 1. Evolutions in AD therapy—From A β to tau therapy

AD is characterized by the aggregation of A β (A) as extracellular A β plaques followed by aggregation of hyperphosphorylated tau (T) as intraneuronal neurofibrillary tangles. A β is considered the initiator of the disease process and tau the executor of the disease process, with tau aggregation strongly correlating with symptom progression. Recent developments led to FDA-approved anti-A β immunotherapy (lecanemab and donanemab), while emerging data indicate early signs of biological halting based on tau PET measured slowing of tau pathology for tau-immunotherapy (bepranemab, E2814) and antisense oligonucleotide (ASO)-based therapy (BIIIB080). The clinical-biological framework for AD, based on the presence of ATN-pathologies (A, A β pathology; T, tau pathology; N, neurodegeneration; and I, inflammation), provided a critical—biological—framework enabling accelerated trial optimization toward clinical efficacy and approved therapies (schematic presentation of ATN framework, based on and modified from Jack et al.⁹ and Jucker and Walker¹²).

neurodegenerative process downstream of A β , including A β -independent roles.^{5–8,13–19} Indeed, tau pathology closely correlates with symptom progression.^{18–24} Besides AD, progressive tau aggregation characterizes a family of various neurodegenerative disorders, including progressive subcortical gliosis (PSP) and corticobasal degeneration (CBD), which represent primary tauopathies and additional unmet medical needs.^{16,25,26} Genetic tauopathies with autosomal dominant

mutations in the *MAPT* gene unequivocally identify tau as a causal, driving factor in the neurodegenerative process in tauopathies.^{16,17,25–31}

Anti-A β immunotherapies: Recent breakthrough successes

Therapeutic strategies targeting A β , including anti-A β immunotherapy, have been intensively pursued to inhibit the initiation of the

AD pathogenic process.^{5,14,32–38} A β -targeting therapies underwent a long “unsuccessful” phase, often casting doubt on the underlying amyloid cascade hypothesis.^{35,36} This optimization track finally converged on the recent breakthrough in anti-AD therapies with FDA approvals of anti-A β immunotherapy therapies for lecanemab and donanemab.^{33,34,37,39–43} Both show marked, near complete clearance of A β plaques in a drug dose-dependent manner, accompanied by a moderate slowing of cognitive deterioration.^{33,34,40,41} In the phase 3 randomized, placebo-controlled CLARITY AD trial, anti-A β immunotherapy using lecanemab significantly reduced A β plaques, improved biomarker levels, and slowed cognitive decline by 27% in patients with early AD (Figures 1 and 2).⁴¹ Lecanemab received first accelerated and subsequently full FDA approval in June 2023. Similarly, promising results followed from the TRAILBLAZER-ALZ 2 phase 3 trial of donanemab, showing fast and efficient A β clearance and a 29% reduction in disease progression overall (CDR-SB [clinical dementia rating-sum of boxes]), which was up to 40% in patients with low to intermediate tau levels (Figures 1 and 2).⁴⁰ These results highlight the efficacy of A β -targeting immunotherapies, while not yet obtaining full clinical halting.^{33,34,40,41}

Lessons from anti-A β clinical trials

The long iterative optimization of A β -targeting therapies, not only converged on FDA-approved drugs but also provided a framework for accelerated AD therapy development. It took several decades on a path of setbacks and incremental partial successes to attain near complete clearance of A β pathology—reducing high A β PET to normal or low levels (from 70–120 centiloids to below 24 centiloids)—and concurrent moderate but significant clinical efficacy, now achieved with lecanemab and donanemab as FDA-approved therapies (Figures 1 and 2).^{33,40,41} Proof of anti-A β immunization to clear A β plaques was delivered for the first time in 1999 in preclinical models.⁴³ The availability of preclinical models robustly mimicking AD pathologies provided the crucial basis for this breakthrough. This preclinical proof, shown consistently and in different models, was followed by the first successful clearance of A β pathology in patients enrolled in anti-A β immunization trials, showing biological slowing of AD (first active and later passive immunization trials)^{37,39,44,45} (Figures 1 and 2). This landmark was however not immediately associated with effective halting of clinical symptoms^{37,39,44,45} and was also associated with serious adverse effects.^{37,39} Subacute meningoencephalitis in a subset of AD patients after active A β 42 immunization^{46,47} resulted in a more intensive pursuit of passive immunization approaches, which were considered safer.⁴⁸ However, the latter, including lecanemab and donanemab, are also not fully devoid of side effects, as A β -related imaging abnormalities (ARIA) are observed in subsets of patients and requires close mitigation, monitoring, and patient selection.^{33,34,40,41,48} Furthermore, the initial lack of clinical effect, despite indications of biological slowing, highlighted the need for further optimization of targeting strategies and trial design.

A variety of anti-A β passive immunization strategies have been tested in clinical trials with incremental success. This included the testing of multiple antibodies targeting different A β epitopes (various N-terminal or mid-region epitopes), and with different affinities for smaller to larger aggregates (including, non-exhaustively,

bapineuzumab, solanezumab, crenezumab, gantenerumab, aducanumab, lecanemab, and donanemab),^{33,34,44,45,48} on a path toward increasing insights into antibody mode of action (MoA) and more effective trials (Figure 2 and comprehensively reviewed in Boxer et al. and others^{33,34,40–42}). Failures in meeting primary endpoints not only resulted from non-optimal epitopes or targeting strategies, but also from a combination of different factors, including initiating treatment too late in the disease course, sub-optimal patient selection (clinical endpoint-based instead of biomarker-based), patient heterogeneity, and disregarding advanced tau status as an independent driver of disease progression. Furthermore, insufficient A β clearance—not reaching threshold for clinical benefit—and adverse effects halting trials might also have contributed to the lack of clinical benefit in A β trials.^{33,34}

However, these initial trial failures did pave the way for the definition of a biological framework of AD, based on ATN pathologies^{7–11} and groundbreaking biomarker development^{4,8,22–24,49–57} to monitor biology. In particular, the development of AD and tauopathy biomarkers, i.e., A β and tau PET, cerebrospinal fluid (CSF)- and blood-based biomarkers^{7,49–63} provided an unprecedented window into AD processes in the brain. This biological monitoring and biomarker advances have been crucial to transform AD trials across four domains: (1) accelerated optimization of targeting strategy (N-terminal epitope/A β aggregate conformation, immunoglobulin G [IgG] isotype, and MoA) vs. clinical outcomes and target engagement; (2) optimized patient stratification and defined an optimal window of intervention based on tau burden, co-pathologies, co-morbidities, genetics (e.g., apolipoprotein E [APOE] genotype), disease stage, and progression rate; (3) delivered rapid and sensitive measures of treatment efficacy, defining thresholds and dosing regimens to achieve the biological modification required for clinical benefit^{32–34}; (4) balanced safety and efficacy with biomarker-informed stratification of patients at risk of adverse effects (reviewed in Boxer et al.,³³ Jucker and Walker,³⁴ and Brody and Holtzman⁴⁸). The use of biological readouts and optimizing biological halting toward clinical efficacy was pivotal for developing approved therapies with clinical effect—lecanemab and donanemab—(Figure 2) resulting in significant but not yet complete disease halting.

From A β to tau therapy

Although anti-A β immunotherapies can almost completely clear A β pathology in most individuals over 12–18 months, they only partially normalize tau biomarkers (e.g., CSF and plasma tau, tau PET).^{40,41} One potential explanation is that tau pathophysiology may become or develop A β -independent, with tau pathology propagating in the brain independent of A β , as demonstrated in the primary tauopathies.^{30,31} Thus, the removal of the initial A β trigger may not sufficiently halt tau propagation in AD, highlighting the need to pursue anti-tau therapies, particularly in view of the unmet clinical need for effective, preferably complete halting of disease progression.

Whether tau therapies can be successful for AD remains to be seen, but there are several arguments supporting tau as a key driver and target in tauopathies. These include (1) tau genetic mutations in tauopathies indicating tau dysfunction as a process sufficient to drive neurodegenerative symptoms^{16,21,25,26}; (2) progressive spatio-temporal tau aggregation characterizing all

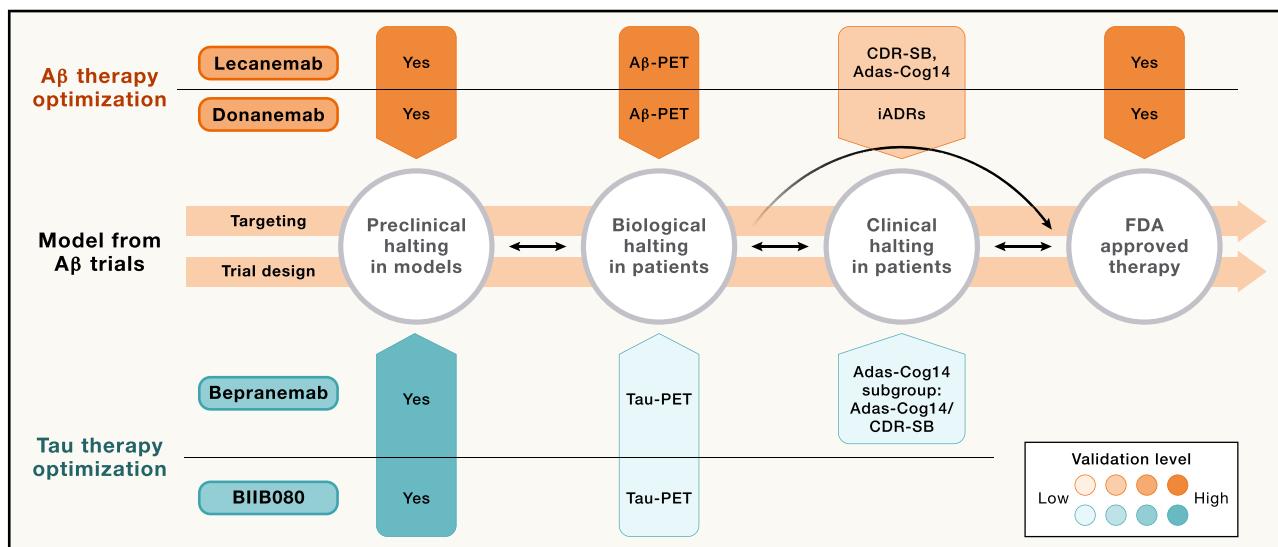


Figure 2. Lessons from anti-A_β clinical trials: Roadmap for accelerated trial optimization from preclinical to biological and clinical halting for regulatory agency therapy approval

Lessons from the long and incremental learning path from anti-A_β trial optimization toward approved anti-A_β therapies, provided a roadmap for accelerated trial optimization for anti-A_β therapies, tau therapies, and combined A_β-tau therapies. The roadmap encompasses toggling back and forth, to move from optimal preclinical validation to obtaining optimal biological validation toward improved clinical efficacy and regulatory agency-approved therapies. The stage for current therapies and trial outcomes for A_β (lecanemab and donanemab) and tau (bepranemab and BIIB080) along the roadmap are indicated. The A_β and tau trial and therapy optimization show different levels of validation and different levels of efficacy of achieved preclinical, biological, and clinical slowing of the disease process (as discussed). This roadmap enables accelerated development of increasingly effective therapies with controlled side effects, through continuous targeting optimization and trial design optimization.

tauopathies in association with symptoms and symptom progression^{3–11,13–19}, and (3) propagation of tau pathology between cells as a potential targetable process, contributing to pathogenetic processes.^{12,64–66} While some open questions and challenges remain, tau targeting is a straightforward approach with recent emerging data supporting its potential feasibility and safety.

The learnings from the anti-A_β trials generated a validated roadmap for accelerated trial optimization toward more clinically effective therapies with fewer side effects. The roadmap includes toggling back and forth from (1) preclinical validation to (2) proof of biological validation in patients, to increased (3) clinical efficacy, and (4) approved therapies (based on biological or clinical efficacy), by continuous optimization of targeting strategy and trial design (Figure 2), a roadmap now useful for tau-targeting therapies.

TAU THERAPIES

Tau pathobiology and tau therapies: Challenges and opportunities

Tau is a microtubule-binding protein mainly expressed in neurons and limited in glia in the adult human brain, which exists in 6 different isoforms that differ in the number of inserts (0, 1, or 2) in the N-terminal domain and number repeat domains (3 or 4) in the microtubule-binding region (MTBR)^{25–29,67,68} (Figure 3). As an intrinsically disordered protein, tau exerts its physiological functions, while in pathological conditions tau aggregates strongly correlate with progressive neurodegeneration and clinical symptoms in tauopathies.^{24–28}

In tauopathies, tau monomers aggregate into various oligomers and filaments, with different molecular and structural characteristics depending on the respective tauopathy^{69,71,72} but sharing a large MTBR region in their aggregate core. Tau aggregation is markedly accelerated by templated tau seeding, i.e., pre-aggregated tau inducing fast aggregation of physiological unfolded tau. Intercellular transmission of tau seeds then provides a compelling mechanism for the fast progressive development of tau pathology in tauopathies (Figure 3). Templated seeding and intercellular propagation of tau pathology has been shown *in vitro* and *in vivo* in preclinical models,^{64,65,73–75} and is supported by accumulating evidence in humans, while clinical validation is still missing.^{76–79} This process, discussed in detail below, encompasses an extracellular phase of pre-aggregated tau, which is potentially important in the context of tau immunotherapy.

Tau undergoes a variety of post-translational modifications (PTMs), including phosphorylation, glycosylation, acetylation, as well as truncation^{16,25,28,29} (Figure 3). Hyperphosphorylation is an important and intensively studied PTM in tauopathies, causing tau to detach from microtubules and facilitating tau aggregation. Tau can also be cleaved and secreted from neurons mostly as N-terminal fragments that can be measured in biofluids but are not prone to aggregate.⁸⁰ Conversely, tau fragments containing the MTBR region may misfold and seed tangle pathology and can also be released as MTBR tau fragments, detectable in biofluids albeit in lower quantities.^{49,58,63} MTBR-containing tau, full length or fragments, can serve as a substrate for tau aggregation, a process that also involves full-length tau containing both N- and C-terminal sequences, while full-length

tau seems more rarely released from neurons.⁸⁰ Besides fibrillar tau aggregates, oligomeric tau forms are particularly considered as toxic agents in the disease process.^{81,82} Furthermore, targeting tau based on pathogenic PTMs may leave physiological tau unaffected and functional but may also only target a subpopulation of pathological tau, leaving other toxic subpopulations untargeted. In the context of passive immunization, truncated tau species are of particular interest as epitope choice might define the targeting of total or a subset of tau pools (Figure 3).

Tau is hence a complex molecule forming a variety of tau species and polymorphs (3R/4R, PTMs, structure, and aggregation state), with physiological and pathological roles in tauopathies. While pathognomonic tau aggregates mainly accumulate intracellularly, in contrast to extracellular A β plaques, various tau forms are detected both intra- and extracellularly. These tau forms may contribute differently to physiological and pathogenic processes, including progressive tau pathology and/or tau-associated neurodegeneration, which are not necessarily identical processes and remain to be understood in detail.^{28,29} Based on this pathobiology, tau targeting presents with specific challenges and opportunities.

Tau therapies: Proof of concept in patients

Different strategies for halting pathological tau progression and its executive role in neurodegenerative tauopathies have been developed and are being pursued.^{15,17,27,70,83-86} Preclinically, small molecules as well as biological approaches are being explored at various stages of the development pipeline, including decreasing tau expression by antisense oligonucleotide (ASO) and small interfering RNA (siRNA) therapies, altering tau PTMs (e.g., phosphorylation, acetylation, glycosylation, and truncation), active tau vaccination, passive tau immunization, promoting tau degradation via proteolysis targeting chimera (PROTAC), tau aggregation inhibition, and O-GlcNAcase enzyme inhibitors.^{15,17,27,61,70,83-86} Emerging outcomes for tau-directed ASO and “second-generation” tau-immunotherapy therapies are showing the first signs of disease-modifying effects, based on slowing PET-assessed tau pathology progression and early signs of cognitive benefit.

Proof of concept for tau-directed ASO as a therapeutic strategy was first demonstrated in preclinical models, showing decreased tau pathology and tauopathy-induced neurodegenerative changes^{87,88} without major side effects. siRNA approaches for reducing tau expression have also been developed and tested preclinically in a murine tauopathy model.⁸⁹ Development of ASO tau therapy was built on and encouraged by early successes using ASO against mutant *SOD1* in genetic amyotrophic lateral sclerosis (ALS), which showed treatment-induced lowering of CSF and plasma concentrations of neurofilament light chain (NFL), an established biomarker for neurodegeneration intensity expected to translate into clinical benefit. On this basis, the FDA-approved tofersen for mutant *SOD1* ALS (FDA).^{90,91} ASO- or microRNA-based therapies also present promising clinical results in other neurodegenerative diseases (Huntington’s disease [adeno-associated virus (AAV)-based gene therapy]⁹² or spinal muscular atrophy^{93,94}).

Several ASO-based clinical trials aimed at reducing tau expression are ongoing (BIIB080 [in AD], NIO752 [in AD and

PSP], and LY3954068 [in AD]). Highly interesting and promising results emerged from a phase 1b trial of tau-directed ASO BIIB080 (NCT03186989), currently being evaluated in a phase 2 clinical study (CELIA study: NCT05399888). In the phase 1b study, a 36-week multiple-ascending-dose trial followed by a 64- or 71-week open-label long-term extension in 46 participants with mild AD, BIIB080 dose-dependently reduced soluble total-tau and p-tau in CSF and aggregated tau pathology measured with tau PET. Tau PET showed reduced tau accumulation in BIIB080 versus placebo arms at week 25 ($n = 13$), and at 100 weeks versus baseline across all regions assessed in the BIIB080 arm ($n = 12$).^{95,96} The result in the BIIB080 arm was associated with suggested favorable trends in cognitive outcomes^{95,96} (Alzforum ADPD 2023, CTAD 2023⁹⁷); BIIB080 was fast-tracked by the FDA in April 2025. The successful biological slowing of tau pathology with the tau-targeting ASO therapy, BIIB080, is an important breakthrough, reminiscent of the A β field (Figures 1 and 2), while follow-up studies are required.

Promising results are also emerging from second-generation passive tau immunotherapies⁹⁸ (Alzforum),⁹⁹⁻¹⁰¹ following first-generation tau immunotherapies that failed to show clinical efficacy. In these latest trials, passive tau immunotherapy with bepranemab now successfully slowed tau pathology progression (tau PET; slowing tau accumulation by 33%–58% versus placebo after 80 weeks of treatment) and cognitive decline (Alzheimer’s disease assessment scale-cognitive subscale 14-item version [ADAS-Cog14]; reduced by 21%–25% versus placebo) as secondary endpoints (TOGETHER-trial $n = 466$; Alzforum).⁹⁸⁻¹⁰¹ Although primary endpoints were not met in the whole study population, this trial now shows biological slowing of tau pathology (tau PET) associated with significant, moderate cognitive benefit (ADAS-Cog14) for tau-directed trials (Figures 1 and 2).

Along the same lines, the second-generation tau antibody E2814 slowed tau pathology progression, measured by PET imaging, albeit only in a few patients, and reduced CSF concentrations of MTBR-tau243, a tau fragment strongly correlated to tau PET, in a larger sample. Phase 2 data with additional second-generation tau antibodies are expected by the end of 2025 (E2814/etalanetug, posdinemab, BMS-986446, and MK-2214) and will be important to generate new insights for future trial development, including defining optimal targeting strategies, optimal biomarker-based patient selection, endpoints, and others further discussed below.

Together, these results provide now evidence of biological disease modification of tau pathology (bepranemab, E2814, and BIIB080) and emerging functional benefits (bepranemab) by tau-directed ASO therapy and passive tau-immunotherapy therapies. While corroboration of the outcomes in multiple and large future clinical trials will be critical, these pave the way for tau trial optimization and development of increasingly effective therapies.

Anti-tau immunotherapies: Optimization of tau targeting and clinical trial design

The recent achievements of tau-immunotherapy trials built on learnings in therapy development by optimizing targeting efficacy and trial design. Despite intracellular accumulation of tau aggregates

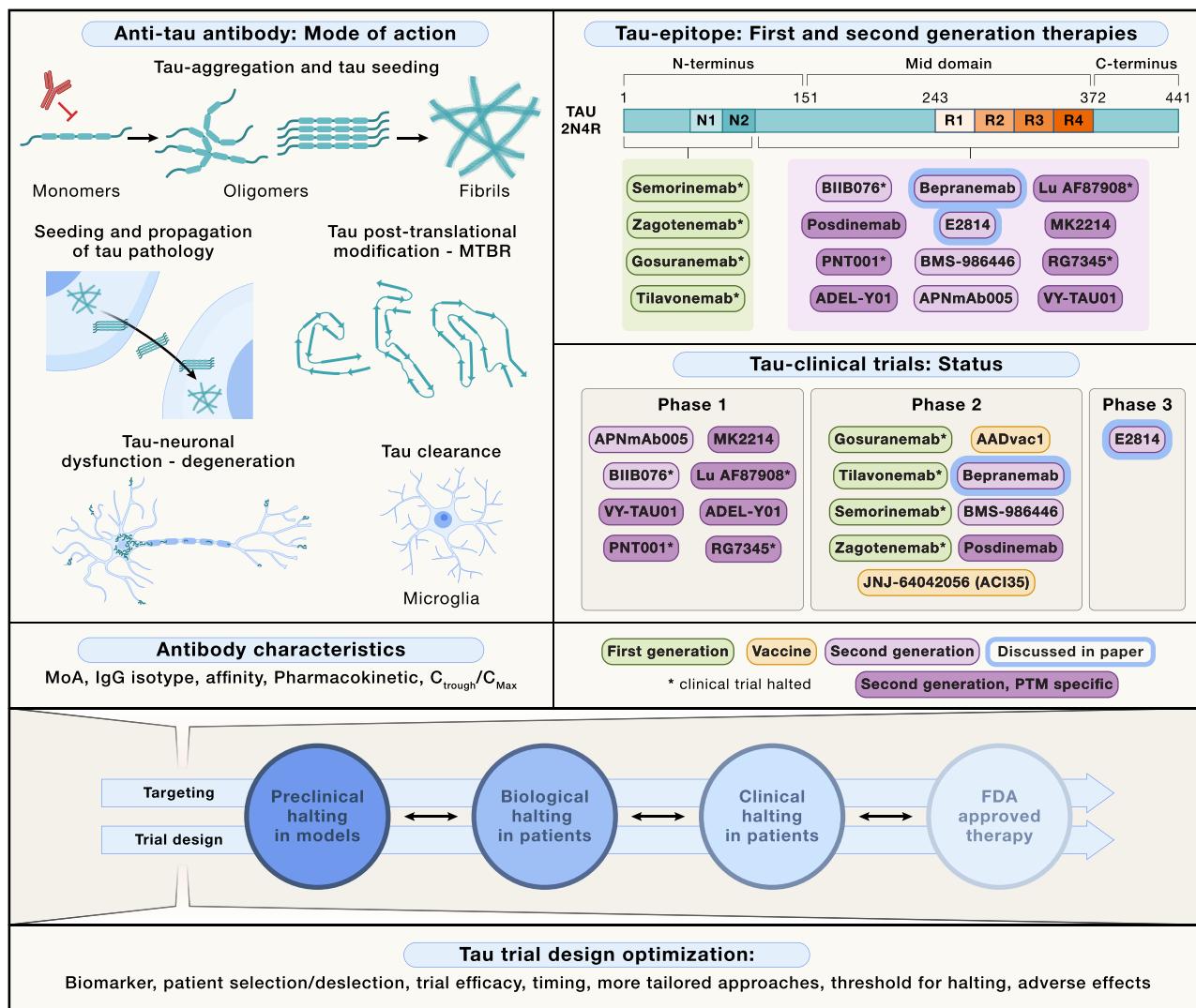


Figure 3. Lessons from anti-tau immunotherapies: Optimization of tau targeting and clinical trial design

Optimization of anti-tau immunotherapy is currently ongoing by continued targeting optimization and trial design optimization. A roadmap toggling between preclinical halting, to biological halting, to clinical efficiency is pursued. First- and second-generation therapies have been or are being developed, with first-generation therapies, targeting N-terminal tau epitopes. First- and second-generation antibodies display a variety of targeting epitopes, tau forms being targeted, antibody characteristics, as well as proven MoA of the antibody. Clinical trials are in different phases of development, and clinical trial design is being optimized based on biomarker use and specific parameters. (Schematic presentation of tau structures and tau propagation, based on and modified from Parra Bravo et al.²⁸ and Shi et al.,⁶⁹ tau clinical trials based on and modified from Cummings et al.⁷⁰).

presenting an additional challenge, tau immunization was successfully and reproducibly shown to halt tau pathology in preclinical models.^{27,102–107} This led to various trials pursuing tau immunotherapy pipeline including^{27,102–106,108,109} passive and active (e.g., AADvac-1 and JNJ-64042056 [ACI-35]) immunization therapies. Here, we focus on the progress of passive tau-immunization trials. It is important to note that anti-tau antibodies may strongly differ in their MoA and target different tau-related pathogenic processes and forms. The MoA may include blocking tau cleavage, PTMs, and misfolding (chaperone-like), blocking oligomeric tau, stimulating microglial phagocytosis (opsonization), interfering with seeding and transcellular tau propagation (extra-cellular release and uptake), and blocking intracellular effects of

tau, depending on the targeting strategy and the site of action (Figure 3).

First-generation anti-tau immunotherapies: N-terminal targeting of tau

First-generation passive tau immunotherapies targeted solely or partially the N-terminal domain of tau (Figure 3), including tilavonemab, gosuranemab, semorinemab, and zagotenemab.⁸⁵ However, they differ significantly in binding affinities (0.1 nM for gosuranemab; 3.8 nM for semorinemab; 20 nM for tilavonemab) and pharmacokinetic profiles (half-lives of > 550 h for gosuranemab and tilavonemab; 400 h for semorinemab). These very different properties led to varying target engagement in the brain for the same dose.¹¹⁰ Furthermore, most

first-generation tau antibodies (gosuranemab,¹¹¹ semorinemab,¹¹² and tilavonemab¹¹³) were of the IgG4 isotype, while zogotenemab (registered as NCT03518073) was an IgG1. IgG1 binds with higher affinity to Fc gamma receptors than IgG4, leading to better phagocytosis of the antibody-antigen complex by macrophages in the periphery or microglia in the CNS, accompanied by pro-inflammatory cytokine release. It had been shown that both tau antibodies with IgG1 or IgG1 with Fc mutation leading to complete lack of Fc gamma receptor binding could be effective, suggesting that effector function is not required for efficacy,¹¹⁴ while IgG1 might increase effector functions and intracellular uptake.^{105,115} However, IgG1 can lead to pro-inflammatory cytokine release, which would increase the risk of neuroinflammation.¹¹⁴ Therefore, it is important to consider the choice of IgG isotype, besides the tau epitope the antibody targets.²⁷

First-generation tau trials unfortunately failed endpoints. Tilavonemab (N-terminal AA 25–30) was not effective in PSP, with its relatively weak affinity potentially requiring higher doses to achieve high target engagement in the brain.¹¹³ Patient selection in this trial was based on clinical measures, without biologically confirmed tau pathology using biomarkers. Semorinemab (N-terminal AA 6–23) was not efficacious in AD, with a shorter half-life and medium affinity also potentially necessitating higher doses to attain high target engagement.¹¹² Patient selection included both clinical assessment and A β pathology positivity by PET or CSF biomarkers but not tau pathology biomarkers. Despite higher affinity, longer half-life, and target engagement in the CSF, gosuranemab (N-terminal AA 15–24) did not show positive effect on clinical outcomes or imaging biomarkers in PSP or AD,^{111,116} leading to the conclusion that targeting N-terminal epitope tau may not be efficacious in PSP. In the AD trial, patients were selected based on clinical deficits and positive A β PET scan but not based on tau pathology. Despite a sub-nanomolar affinity, zogotenemab (N-terminal AA 7–9) had no effect on tau PET in a phase 2 trial of early symptomatic AD.^{117,118} Interestingly, in this trial, patients were selected based on intermediate levels of brain tau on PET imaging.

Taken together, first-generation passive tau immunization did not achieve effects on tau pathology or efficacy with clinical endpoints. The most straightforward explanation could relate to the choice of N-terminal tau epitope, given that all first-generation anti-tau antibodies targeted this region of the protein. In view of the higher accessibility of N-terminal epitopes for antibodies compared with MTBR epitopes, their high immunogenicity, possibly less interference with the physiological role of tau, and the abundant presence of truncated tau encompassing N-terminal regions in biofluids, N-terminal tau targeting was a straightforward and valid strategy to initially pursue. Later on, however, a variety of truncations have been identified, including fragments with either N-terminal tau or MTBR-encompassing tau in human brain and in CSF.^{62,63,119–121} In addition, N-terminal fragments are not prone to aggregate, and N-terminal tau-targeting antibodies were shown to be less effective at inhibiting seeding and propagation of tau pathology *in vitro* and *in vivo* when using human postmortem extracts.^{103,107} Hence, the current data suggest that following tau truncation, it may not be efficacious to target non-MTBR-

containing fragments of tau to prevent seeding and propagation of tau pathology.^{62,63,74,75,103,107,122–125}

Second-generation anti-tau immunotherapies: Optimization of targeting and antibody design

In light of that, the second generation of passive tau immunization instead focused on epitopes closer to the aggregation- and seeding-prone MTBR region, which have the potential to block spread and seeding of tau pathology^{62,63,74,75,103,107,122–125} (Figure 3). This second generation includes BIIB076,¹²⁶ bepranemab,¹²⁷ E2814,¹²⁸ BMS986448 (PRX005), posdinemab (JNJ-63733657), Lu AF87908, MK-2214, APNmAb005, ADEL-Y01, VY-TAU01 (likely now called VY7523) (Figures 1 and 3). Like the first-generation therapies, these antibodies are quite different from each other and the data generated with one may not necessarily apply to others.

Some of these antibodies target specific tau PTMs, such as phosphorylation (p217 for posdinemab, p396 and 404 for Lu AF87908, p413 for MK-2214, and acetylated K280 for ADEL-Y01) that may increase engagement of specific pathological tau forms, leaving physiological tau unaffected. APNmAb005 binds a conformational epitope in tau oligomers, and hence binds tau in synaptosomes and insoluble fractions in human brain extracts better than monomeric, cytosolic tau.¹²⁷ BIIB076 targets the middle domain of tau.¹²⁸ Both antibodies halt tau seeding and propagation of tau pathology in preclinical models. Last, VY-TAU01 binds a C-terminal epitope.

In addition, most second-generation tau antibodies are now IgG1 isotype (BIIB076, E2814, BMS986448, Lu AF87908, APNmAb005, posdinemab, and ADEL-Y01) with stronger effector function than IgG4 (bepranemab, VY-TAU01), but which, conversely, may potentially increase neuroinflammation and ARIA risks, if combined with anti-A β antibodies. In this respect, it must be noted that the MoA of immunotherapy may be very different for A β and tau. For A β , strong effector function (IgG1) is important for efficacy, since microglia engagement via the Fc gamma receptor is key for phagocytosing extracellular A β aggregates. The effect is an active mechanism for clearing A β via microglia and is antibody maximum (peak) bloodstream concentration (C_{max})-driven, i.e., once a sufficient antibody concentration is reached in the target organ (brain), microglia actively phagocytose the aggregated A β . A sustained presence of high antibody concentration may not be necessary for removing plaques and a pharmacokinetic profile with rapid brain penetration but a shorter half-life could be appropriate.¹²⁹ For tau, however, it was shown that anti-tau antibodies with or without effector function can both be efficacious in preclinical research.¹¹⁴ The MoA may be related to blocking binding sites besides, or in addition to, opsonization and phagocytic uptake by microglia.

These trials are now at various stages, with some already ceasing clinical development. This is the case for BIIB076, which completed phase 1 trial (NCT03056729), and showed target engagement, based on reduced mid-region tau in CSF, 1 week after infusion, but whose side effects at the highest dose prompted trial termination. Lu AF87908 was in clinical development (NCT0414986) but is no longer listed in the Lundbeck portfolio; the reason for removing this antibody is unknown to us. Some antibodies are starting early clinical trials, such as VY-TAU01/VY7523 (NCT06874621), MK2214 (NCT05466422), APNmAb005

(NCT05344989), and ADEL-Y01 (NCT06247345), but no data are yet available regarding safety, pharmacokinetics, CSF target engagement, and efficacy. Some others have completed phase 1 trials, with safety and target engagement having been considered sufficient to move toward phase 2 proof-of-mechanism trials, E2814 (NCT04231513, NCT04971733, NCT05269394, NCT06602258, and NCT01760005), BMS986446 (NCT06084598 and NCT06268886), and posdinemab (NCT05407818, NCT03689153, NCT04619420, and NCT03375697).

First outcomes are emerging for two antibodies with epitopes in or close to MTBR. The second-generation antibody E2814 (Alzforum 1, Alzforum 2) was shown to reduce early and late tau pathology biomarkers in a subset of patients with dominantly inherited AD with mild-to-moderate cognitive impairment ($n = 7$) that received E2814 in escalating doses (750–4,500 mg) during 6 to 26 months. Two years of treatment reduced CSF p-tau217 and tau-MTBR levels by 50% and 75%, respectively, compared with DIAN observational study data (Wildsmith et al., CTAD24). Study limitations included a small sample size, unblinded design, and lack of a placebo arm. Interestingly, a few patients stabilized with a trend toward reduced tau by PET. Together, the data support target engagement and slowed progression of tau pathology (tau PET and tau MTBR) in a limited number of patients. Currently, E2814 is also being tested in combination with anti-A β therapies, with a readout expected in 2027–2028, further discussed below.

Bepranemab is another second-generation, MTBR-region-directed tau antibody with phase 2 data in AD (TOGETHER-trial $n = 466$; CTAD 2024, Alzforum).^{98,103,107} Bepranemab significantly slowed secondary cognitive (ADAS-Cog14) and tau pathology (measured by tau PET) endpoints in the whole trial population in a phase 2 double-blind, placebo-controlled trial with high (90 mg/kg) and low (45 mg/kg) doses (Alzforum).⁹⁸ This represents the first time that a tau antibody demonstrated an effect on cognitive and pathology endpoints. However, the effect was not statistically significant on the primary CDR-SB endpoint in the whole population, although it was statistically significant in a prespecified sub-analysis of subjects with low baseline tau or were APOE4-negative. Most interestingly, predefined sub-analyses in this clinical trial were formulated based on the hypothesis that (1) patients with low/medium- but proven-baseline tau burden would benefit more than patients with high tau burden, and (2) APOE4 carriers may react differently to non-carriers. These findings may help select patients more likely to respond to tau immunotherapies in future trials.

Bepranemab's MoA was selected on and shown to prevent intercellular propagation of tau seeding in preclinical models. As an antibody with low effector function (IgG4), bepranemab's effects on tau PET and ADAS-Cog14 suggest that strong effector function is not necessary for the MoA of tau antibodies in humans, aligned with preclinical data.¹¹⁴ Given bepranemab predominantly binds extracellularly released tau seeding species,^{103,107} the effect of bepranemab is expected to be C_{trough} -driven, i.e. a high and stable antibody concentration in the target organ (brain) is needed to capture seeding species as they are released from cells, favoring a pharmacokinetic profile with longer, stable half-life. Of note, several promising immunization approaches with a distinct goal of increasing intracellular target-

ing of tau and its pathogenetic effects to halt tauopathies are also at various stages of development (e.g., intrabodies, liposomal antibody administration, and others).^{27,86,104,115,130,131}

These emerging outcomes suggest that screening assays used to identify bepranemab and related tau-immunization strategies could translate to clinical effects, at least partially, and are providing insights into optimal tau-targeting epitopes (MTBR-encompassing domain and bepranemab/E2814) and antibody characteristics (IgG isotypes and pharmacokinetics) (Figure 3). In addition, emerging outcomes of various ongoing trials effectiveness in humans may further provide crucial insights about optimal tau-targeting strategies, as well as the exact pathogenetic forms of tau and their role in tau pathology progression and tau-induced neurodegenerative processes.

Second-generation anti-tau immunotherapies:

Optimization of clinical trial design

Clinical trial design is an important issue that, at least in part, may have contributed to the failure of first-generation anti-tau immunotherapies (Figure 3). However, adopting the framework provided by A β trials can help identifying critical parameters for optimal design of tau trials (Figures 1 and 2).

In terms of safety and adverse effects, tau immunotherapy seems to display a better safety profile compared to anti-A β immunotherapy. No ARIA has been described with any of the tau antibodies, and, more generally, no major safety concerns have been identified so far in non-human primate toxicology studies, nor in phase 1 or phase 2 studies of healthy volunteers, AD, or PSP patients with a variety of tau antibodies, apart from BIIB076 at high doses.

In terms of patient selection, using PET/CSF and/or blood-based biomarkers is pivotal to select patients more likely to respond to the treatment, define trial efficacy, set thresholds of biological effects that translate to functional outcomes, and shorten recruitment time. Successful outcomes in predefined bepranemab phase 2 sub-group analyses suggested that selecting patients positive for A β but lower, yet proven, baseline tau pathology or APOE4-negative (often with comparably lower tau pathology) is important for tau therapeutic antibodies. This is also being used now in other trials. In the TargetTau-1 trial for the tau antibody BMS986446, presence of tau pathology is part of the inclusion criteria. Moreover, for the AUTONOMY trial testing the p217-specific tau antibody posdinemab, plasma p-tau217 followed by intermediate (not high, not absent) levels of tau PET are inclusion criteria.

In AD and primary tauopathies, different biomarkers related to tau and A β are available. For tau, different tau forms can be measured in fluids yielding insight into different pathologies. For instance, p-tau may be more reflective of A β pathology while MTBR and “total” tau correlate with tau pathology, although the latter should be cautiously employed as not all tau species may be captured by antibodies or visible by mass spectroscopy. Blood biomarkers, such as plasma p-tau (e.g., p-tau-217) for A β -related tau phosphorylation⁵⁹ and eMTBR-tau243 for tau tangle pathology,⁵⁸ should facilitate easier and faster selection of patients with positive A β pathology and low/medium tau pathology using less invasive, cheaper, and readily available tests in larger number of centers. Additionally, glial fibrillary acidic protein (GFAP) and NFL, as well as other complementary markers

indicative of gliosis, neurodegeneration, and synaptic dysfunction (e.g., CSF neurogranin) can be useful in association with functional readouts.¹³² For tau PET, reduction in baseline values, progression rate of tau pathology, and/or progression to different brain regions can be used. Overall, the use and continuous optimization of biomarkers, both blood- and CSF-based and PET imaging,^{23,55,63} is of utmost importance in the context of patient selection.

Clearly, for all these biomarkers (imaging or biofluids), it will also be necessary to determine thresholds of biological effects that translate into symptom-slowng effects. Accordingly, among different anti-A β trials, increased biological effects on A β plaque clearance were associated with slowing of cognitive decline.^{32,33} Reducing A β pathology to a threshold below 24 centiloids (on PET, \pm 70% reduction) is considered necessary for clinical efficacy. Importantly, thresholds of biological effects for tau required for clinical efficacy may differ in AD compared with primary tauopathies.

Besides patient selection, another concern impacting trial outcomes is patient exclusion. Currently, patients are not excluded based on the presence of alpha-synuclein (aSyn) pathology, a co-morbid pathology in \sim 30% of patients clinically diagnosed with AD.¹³³ aSyn by itself can induce cognitive deficits, and Lewy body dementia can be misdiagnosed clinically as AD, as well as affect AD clinical trajectories.¹³⁴ Clinically diagnosed AD patients with aSyn may not respond to tau antibodies, enhancing clinical trial variability and impacting absence of efficacy outcomes. Optimized patient stratification based on excluding non-AD or AD patients with high co-morbidities and/or co-pathologies that drive cognitive deficits and symptom progression can lead to improved clinical trial design and outcomes. Other factors may include APOE genotype, stage of tau pathology, type of tauopathy, mono or combination therapy. This exclusion will be particularly important in initial tau-therapy validation, while future combined therapies can potentially enable treatment of patients with co-pathologies (aSyn and TDP43, among others) or different subgroups of patients.

The choice of endpoints is critical for demonstrating effective disease modification in clinical trials. The best-suited clinical endpoint depends strongly on the trial stage, target population, and regulatory strategy. The CDR-SB and iADRS (the integrated AD rating scale) are the current endpoints of choice following their use in the lecanemab and donanemab trials,^{40,41} respectively. The CDR-SB is widely accepted and recognized as a primary endpoint by regulatory agencies for its broad assessments of cognition and function. CDR-SB assesses six domains and is sensitive in early AD, showing dynamic range in early symptomatic stages (mild cognitive impairment [MCI] due to AD and mild AD dementia). However, it may be subject to inter-rater variability and has limited granularity. The iADRS is a composite measure of ADAS-Cog and Alzheimer's Disease Cooperative Study-Instrumental Activities of Daily Living (ADCS-iADL), which offers an integrated view of cognitive and functional decline and was used to validate donanemab and recent anti-A β trials. The iADRS has enhanced sensitivity to change, with a good signal-to-noise ratio in early AD and MCI but may still lack long-standing regulatory recognition despite the growing momentum. Both endpoints are suitable for disease-modifying therapy trials, but

neither are AD-specific nor directly confirm "disease modification," which requires additional biomarker evidence (e.g., A β or tau PET, CSF, or blood markers). Hence, current best practices for maximizing the success of AD trials may involve using CDR-SB or iADRS, supported with biomarker evidence to indicate disease modification. Preferentially, secondary measures such as CGIC/CaGIC (clinical/caregiver global impression of change), quality of life and caregiver burden should also be included.

Overall, targeted epitope, IgG isotype, PTM, and pharmacokinetic characteristics are important design features for tau antibodies. Additionally, key crucial trial design considerations include the use of biomarkers of target engagement and targeting efficacy, biomarker thresholds that translate to clinical efficacy, patient selection (who and when), as well as exclusion and a choice of endpoint critical for demonstrating effective disease modification.

Back-translating tau immunotherapy trials: Clinical validation tau pathology propagation and its mechanisms

Previous research demonstrated the concept of seeding and propagation of tau misfolding and pathology in preclinical tauopathy models.^{12,25,64–66,73–76,123,135,136} Extracellular release of misfolded tau and its subsequent uptake seeds aggregation of physiological intracellular tau in the recipient cell. This process bypasses the long lag phase of tau aggregation, accelerating and propagating tau pathology throughout the brain in functionally connected brain regions (Figures 3 and 4).^{12,25,64–66,73–76,123,135–139} Accumulating evidence showing the presence of seeding- and propagation-competent tau forms and tau seeds in human brains of patients with AD and primary tauopathies support these concepts.^{12,34,64–66,73–75,79,140} In human tauopathy patients, imaging data, functional connectivity, rate of disease progression, and tau seed analysis further support propagation of tau pathology and its contribution to progressive tau pathology and disease progression.^{30,31,51,78,79,141–143} However, clinical proof of propagation of tau pathology in human AD brains is now emerging from the effects of bepranemab on tau PET, and of E2814 on tau PET/MTBR-tau243, both designed to target seed-competent tau and its propagation in preclinical models. The current trial outcomes thereby support a role for tau-seed-based propagation of tau pathology as a clinically validated contributory mechanism to progressive AD in humans.

Mechanistically, several cellular and subcellular mechanisms can contribute to pathological tau propagation in the brain.^{28,144,145} Exosomes, tunneling nanotubes, unconventional secretion, extracellular vesicles, and multivesicular body release of tau seeds have been identified as mechanisms of tau release from cells.^{28,146} Conversely, different molecular mechanisms have been identified for cellular uptake of tau seeds, including by interaction with (low-density lipoprotein receptor-related protein 1) LRP1 and heparan sulfate proteoglycans (HSPGs),^{28,146–148} via MTBR region.^{28,131,147,148}

In the bepranemab and E2814 trials, most of the antibody is anticipated to be present in the extracellular compartment, where it targets extracellular, freely available seed-competent tau, without—or with limited—access, to tau in tunneling nanotubes or extracellular vesicles. The successful slowing of progressive

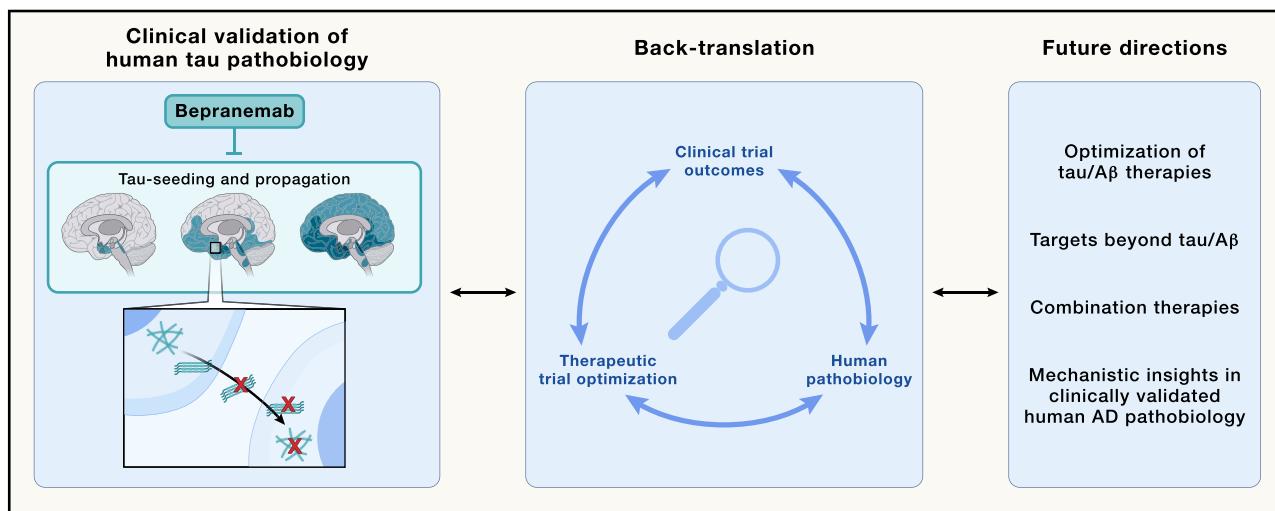


Figure 4. Back-translation to clinically validated insights in human tau pathobiology and future directions

Back-translation enables clinically validated human tau pathobiology insights useful for future development of tau-targeting therapies. Back-translation of clinical outcomes to human tau pathobiology and its mechanistic insights, is enabled by the specific, well-known and well-defined targeting epitope and MoA of passive immunotherapy. In general, back-translation of human AD and tauopathy pathology enable development of mechanism-based AD and tauopathy therapies toward increasingly effective trials for AD and tauopathies. Future trials extend to optimized trials for A β and tau, combination trials of A β /tau, and targets beyond A β and tau, including neuroinflammation and the neurodegenerative process (i.e., its hallmarks¹⁹ in AD and tauopathies (N)). This optimization process is anticipated to lead to increasing insights into human pathobiology and increasingly effective trials with clinical efficacy. (Schematic presentation of tau propagation, based on and modified from Parra Bravo et al.²⁸).

tau pathology in these trials suggests that a considerable portion of seed-competent tau is freely available in the brain parenchyma.^{28,105,115} It is noteworthy that mechanisms of antibody internalization into neurons, and subsequent transfer to the cytosol, have been identified, and enhanced intracellular delivery is being therapeutically pursued, representing interesting alternative or additional approaches to target non-freely available seeds, or tau intracellularly.^{104,115,130,149,150} For bepranemab however, neuronal cytosol levels depend on its uptake and subsequent lysosomal membrane rupture and are anticipated to be low compared with extracellular antibody levels.

The fact that these antibodies both target and bind the MTBR region, shown as necessary for tau seeding in preclinical models, suggests that targeting MTBR-encompassing tau forms and interfering with tau seeding presents a promising therapeutic strategy. However, consolidation of these insights will require confirmation of the disease-modifying effects in future trials. That no complete halting is yet obtained also indicates the need for further optimization.

Recent studies have identified similarities and differences in structure of mature tau aggregates in different tauopathies.^{71,72,151} Therefore, tau antibodies that target different tau strains may need to be developed for different tauopathies.^{71,72,151} Cryogenic electron microscopy studies have shown that the core structure of tau aggregates is composed of a subsegment of the MTBR domain (R3 and R4), while specific conformations vary according to the tauopathy type (Figure 4).^{124,125} However, the common involvement of the MTBR tau domain again underscores its crucial role in tau aggregation. While progressive tau pathology strongly correlates with disease progression,^{18–24} the executive tau polymorphs of the neurodegenerative process, driving clinical symptoms, still

remain unidentified. Fully mature tau aggregates may not represent the main culprits, but rather oligomeric and intermediate soluble tau aggregates could be the toxic species.^{81,82} Upcoming trial results from second-generation tau immunotherapies have the potential to shed light on specific tau forms and their respective contribution to the neurodegenerative process.

Despite the emerging insights, several questions regarding tau pathobiology remain. Although the bepranemab data provide clinical validation and insights into tau propagation, detailed mechanisms of tau uptake, clearance, and tau toxicity are still unclear. Furthermore, within the bepranemab's AD trial, APOE4 carriers displayed differential outcomes, which can suggest more aggressive biological processes, although this remains mechanistically poorly understood. Future trials may provide more insights into the differing tau pathogenetic processes dependent on APOE genotype. Similarly, differing mechanisms of tau pathogenetic processes in AD vs. primary tauopathies need to be clarified, as well as mechanisms of the progression and interaction of A β and tau co-pathologies in AD.

Taken together, the effects of bepranemab and E2814 on tau pathology (PET), cognitive endpoints (ADAS-Cog14), and some biofluid-based biomarkers for tau pathophysiology, provide the first proof of concept for an effect of tau antibodies in the clinic. They also support the concept of tau pathology propagation and spreading to different brain regions (Figure 4). Furthermore, these antibodies mostly have access to freely available extracellular tau, so their clinical effect suggests the contribution of this fraction to tau pathology propagation, while other pools (vesicles, nanotubes, and intracellular tau) may also contribute. Extracellular tau, and the epitope near the MTBR, targeted by these antibodies are considered to participate in the observed effects. Finally, the exact identity of the tau seeding and toxic tau

polymorphs remains unclear; in this respect, the cautiously promising results of bepranemab provide interesting insights, suggesting the antibody may target at least some seeding and toxic species. However, outcomes of ongoing and future tau trials—including postmortem analysis—will be critical to confirm and corroborate these early successes, as well as to gain in-depth insights into optimal targeting strategies and tau pathobiology in human subjects, to back translate the findings to research. This will be essential for the future success of tau therapies and their development into new treatment options for patients.

COMBINATION THERAPIES

With the commercial availability of anti-A β immunotherapies, therapies combining anti-tau and anti-A β strategies can now be explored in clinical trials. AD therapies targeting both initiation and execution mechanisms combined might provide more effective therapies. Currently, E2814 is being tested in combination with lecanemab in the presymptomatic DIAN-TU cohort, with a readout expected in 2028. Another combination trial with E2814 and lecanemab in early symptomatic MCI patients was launched in 2024, with readout expected in 2027. As different stages of pathology are present in various brain regions, combination of A β and tau therapies could target distinct stages of the disease process concomitantly. Combination therapies present an attractive perspective, although they raise additional inherent questions or challenges: are synergistic or complementary effects achieved? What is the impact of combination trials on adverse effects (ARIA, neuroinflammation)? How should these trials be designed and evaluated? What is the ideal timing for each treatment? Will efficacy of the respective targets be evaluated first based on biomarker efficacy, and how will their respective contribution on functional/clinical outcome be assessed? While this is not the aim of this perspective, combination trial designs and evaluation represent highly interesting prospects that will require in-depth and critically constructive consideration and optimization.

Furthermore, beyond A β (A) and tau (T), and their combination therapies, alternative targets are intensively pursued for AD therapies,^{70,83,84,132} as crucial components of the biological framework. Neuroinflammation (involving TREM2, APOE, and others), as an essential component of the pathogenic process,^{152–156} is now incorporated within the biological definition of AD, represented by the A(|)TN framework.^{11,19,152–156} Within this framework “I” represents inflammation and is intensively pursued as target for AD and primary tauopathies, for developing increasingly effective disease-halting therapies. Finally, also other hallmark processes that drive the neurodegenerative process¹⁹—represented by N in the ATN framework—are intensively investigated for therapy development, including synaptic and neuronal network dysfunction, aberrant proteostasis (autophagy/lysosomal dysfunction), cytoskeletal abnormalities (e.g., neurofilaments), altered energy homeostasis, DNA/RNA defects, and neuronal cell death.^{70,83,84,132,157}

LEARNINGS, CHALLENGES, AND FUTURE DIRECTIONS

We highlighted recent clinical advances that marked a turning point in AD therapeutics. Anti-A β therapies, lecanemab and do-

nanemab, showed the first proof of disease modification based on A β clearance (PET based) associated with significant, moderate clinical benefit, and are regulatory agency approved. Trials with the second-generation tau antibody bepranemab and the tau-directed ASO BIIB080 followed up, delivering the first clinical evidence that slowing tau pathology is feasible, as shown by PET imaging, with early signs of cognitive benefit (bepranemab). These latter results represent proof of principle that tau-directed interventions have disease-modifying potential in humans. While corroboration of disease-modifying outcomes in multiple, large future tau trials will be critical and many challenges remain, these studies provide a basis for tau trial optimization, toward increasingly effective trials.

Such progress was built on decades of iterative A β trials, which pioneered the regulatory and biological framework for disease-modifying trials. This framework is built on the biological definition of AD, combined with imaging and biomarkers innovations, enabling validated biomarker endpoints, target engagement as a prerequisite for efficacy, as well as acceptable clinical outcomes (CDR-SB and iADRS). The A β experience also clarified key parameters for successful biologics: appropriate epitope targeting, antibody isotype and affinity, and pharmacokinetics. Furthermore, the concept of threshold of biological change required to achieve clinical benefit was set and critical for success. These insights catalyzed more rational trial designs, including biomarker-based patient selection/exclusion and disease staging.

The tau field has now capitalized on these foundations. Learnings from first-generation tau antibodies guided the refinement of second-generation candidates toward epitopes closer to the MTBR—a node for tau aggregation and propagation. Understanding of affinity, isotype, and pharmacokinetic properties ensures sufficient target engagement in the brain (or its closely related measurable compartment—the CSF), resulting in measurable biological effects on tau pathology (tau PET). Importantly, the absence of ARIA with tau-targeted therapies contrasts favorably with A β immunotherapy, supporting broader safety margins. Various tau-targeting trials are ongoing and will be critical to yield increasing insights and efficacy.

Clinically, these findings enable valuable back-translation to human tau biology. The effects of bepranemab/E2814 on tau pathology (tau PET/MTBR tau) and cognition (bepranemab) suggest that trans-synaptic propagation of extracellular free tau—outside nanotubes or vesicles—occurs in humans, contributes to tau pathology progression and is therapeutically tractable. Current outcomes support the involvement of the MTBR-region—encompassing forms, and efficacy of an IgG4 antibody supports the concept shown preclinically that strong effector function may not be necessary for tau immunotherapy. Together, the present data validate preclinical screening cascades in pre-clinical models as used for bepranemab and BIIB080 as more predictive of clinical translatability. Noteworthy, the recent outcomes indicate safety and early efficacy of recent trials, despite incompletely understood mechanisms. Future trials will be critical for confirmation as well as optimization toward increasingly effective tau therapies. While mechanistic understanding is not essential for effective therapies, it can guide innovative, more effective therapies.

Future development will build upon these mechanistic and trial design insights. Third-generation tau therapies will benefit from refined patient stratification, selecting individuals with low-to-moderate tau burden, before irreversible neurodegeneration (high tau pathology), and considering APOE4 status (APOE4 homozygotes responded less to bepranemab). Trial optimization will rely on quantitative tau PET and fluid biomarkers to monitor target engagement, tau pathology progression, spreading, or clearance. Dosing strategies must balance efficacy with manufacturability and global accessibility, given the scale of the AD population (low doses are needed). Thus far, safety outcomes remain favorable, providing a solid basis for long-term treatment.

Several challenges remain. The impact of co-pathologies, timing of intervention, and threshold of biological effect required for clinical benefit are not yet fully defined. The interplay of tau and A β also raises critical questions regarding sequence and combination of therapies, particularly given ARIA risk with high-effector A β antibodies. Moreover, the exact toxic tau species and mechanisms of neuronal injury remain incompletely characterized, but their elucidation may open routes to small-molecule therapeutics, which are potentially more scalable and cost-effective. Corroboration of the current disease-modifying outcomes in future tau trials will be crucial. And optimization of A β and tau trials are under development. Additional pathways, including neuroinflammation, APOE/TREM2 signaling, and pathways driving the neurodegenerative process (i.e., its hallmarks), represent promising targets. Together these efforts contribute to development of increasingly effective disease halting therapies, as complete halting is not yet obtained.

The Alzheimer's field has thus moved beyond the binary view of "A β versus tau." Tau and A β are interacting drivers within a broader neurodegenerative cascade of events, demanding multi-modal and mechanism driven, disease-modifying therapies. Optimization of mono- and multitarget therapies targeting A β and tau as well as other targets will lead to increasingly effective AD therapies. Advances in biomarkers, adaptive trial design, and back-translation from clinical to molecular insights are accelerating the trajectory toward true disease modification.

Despite the cautious enthusiasm, we acknowledge that no current treatment can yet sufficiently halt disease progression for patients, and that many important challenges remain. However, the convergence of current developments may provide a hopeful path for AD and related tauopathies for transitioning from an inexorable disorder affecting millions of individuals daily to future treatable conditions.

ACKNOWLEDGMENTS

H.Z. is a Wallenberg Scholar and a Distinguished Professor at the Swedish Research Council supported by grants from the Swedish Research Council (#2023-00356, #2022-01018, and #2019-02397); the European Union's Horizon Europe Research and Innovation Program under grant agreement no. 101053962; Swedish State Support for Clinical Research (#ALFGBG-71320); the Alzheimer Drug Discovery Foundation (ADDF), USA (#201809-2016862); the AD Strategic Fund and the Alzheimer's Association (#ADSF-21-831376-C, #ADSF-21-831381-C, #ADSF-21-831377-C, and #ADSF-24-1284328-C); the European Partnership on Metrology, co-financed from the European Union's Horizon Europe Research and Innovation Program and the Participating States (NEuroBioStand, #22HLT07); the Bluefield Project; Cure Alzheimer's Fund; the Olav Thon Foundation; the Erling-Persson Family Founda-

tion; Familjen Rönströms Stiftelse; Familjen Beiglers Stiftelse; Stiftelsen för Gamla Tjänarinnor, Hjärnfonden, Sweden (#FO2022-0270); the European Union's Horizon 2020 Research and Innovation Program under the Marie Skłodowska-Curie Grant Agreement no. 860197 (MIRIADE); the European Union Joint Program—Neurodegenerative Disease Research (JPND2021-00694); the National Institute for Health and Care Research University College; London Hospitals Biomedical Research Centre; the UK Dementia Research Institute at UCL (UKDRI-1003); and an anonymous donor. G.U.H. was funded by the Deutsche Forschungsgemeinschaft (DFG, German Research Foundation) under Germany's Excellence Strategy within the framework of the Munich Cluster for Systems Neurology (EXC 2145 SyNergy—ID 390857198), JPND Consortium, SynOD, "alpha-synuclein OMICS to identify drug-targets" (funded by the Federal Ministry of Education and Research, BMBF, 01ED2405A). I.D. received funding from Stichting Alzheimer Onderzoek Belgium (standard grants, SAO-FRA20240028 and SAO-FRA20220006), Methusalem Fund UHasselt (BOF Methusalem Fase I – BOF22M02), and BOF25IU03-iBOF/25/028: 2025 iBOF: The Interuniversity BOF projects program. Many elegant studies have directed the field; however, selections needed to be made due to limitations in the number of references. We apologize for omitted references. The manuscript was accepted before the 18th Clinical Trials on Alzheimer's Disease Conference (CTAD, December 1–4, 2025), precluding inclusion of data presented at the conference. This article is dedicated to all that have been or are affected by AD and related tauopathies and to their caregivers, who are combating tauopathies daily and hourly. This article aims to join their fight against AD and tauopathies.

DECLARATION OF INTERESTS

G.U.H. participated in industry-sponsored research projects from AbbVie, Biogen, Biohaven, Novartis, Roche, Sanofi, and UCB; received research support from GE Health; has ongoing research collaborations with Roche, UCB, AbbVie, and Ferrer; served as a consultant for AbbVie, Alzprotect, Amylyx, Aprinoia, Asceneuron, Bayer, Bial, Biogen, Biohaven, Epidarex, Ferrer, Kyowa Kirin, Lundbeck, Novartis, Retropode, Roche, Sanofi, Servier, Takeda, Teva, and UCB; and received honoraria for scientific presentations from AbbVie, Bayer, Bial, Biogen, Bristol Myers Squibb, Esteve, Kyowa Kirin, Pfizer, Roche, Teva, UCB, and Zambon. His institution received research support from AbbVie. H.Z. has served on scientific advisory boards and/or as a consultant for AbbVie, Acumen, Alector, Alzinova, ALZpath, Amylyx, Annexon, Apellis, Artery Therapeutics, AZTherapies, Cognito Therapeutics, CogRx, Denali, Eisai, Enigma, LabCorp, Merck Sharp & Dohme, Merry Life, Nervgen, Novo Nordisk, Optoceutics, Passage Bio, Pinteon Therapeutics, Prothena, Quanterix, Red Abbey Labs, reMYND, Roche, Samumed, ScandiBio Therapeutics AB, Siemens Healthineers, Triplet Therapeutics, and Wave; has given lectures sponsored by Alzecure, BioArctic, Biogen, Cellecronic, Fujirebio, LabCorp, Lilly, Novo Nordisk, Oy Medix Biochemica AB, Roche, and WebMD; is a co-founder of Brain Biomarker Solutions in Gothenburg AB (BBS), which is a part of the GU Ventures Incubator Program; and is a shareholder of CERimmune Therapeutics (outside the submitted work). J.-P.C. is one of the inventors of bepranemab and a stock owner of Discoveric bio alpha.

DECLARATION OF GENERATIVE AI AND AI-ASSISTED TECHNOLOGIES IN THE WRITING PROCESS

During the preparation of this work, the authors used ChatGPT (GPT-5) OpenAI, 2025, as a language assistant in order to improve written text. After using this tool/service, the authors reviewed and edited the content as needed and take full responsibility for the content of the publication.

REFERENCES

1. Gustavsson, A., Norton, N., Fast, T., Frölich, L., Georges, J., Holzapfel, D., Kirabali, T., Krolak-Salmon, P., Rossini, P.M., Ferretti, M.T., et al. (2023). Global estimates on the number of persons across the Alzheimer's disease continuum. *Alzheimers Dement.* 19, 658–670. <https://doi.org/10.1002/alz.12694>.

2. GBD 2019 Dementia Forecasting Collaborators (2022). Estimation of the global prevalence of dementia in 2019 and forecasted prevalence in 2050: an analysis for the Global Burden of Disease Study 2019. *Lancet Public Health* 7, e105–e125. [https://doi.org/10.1016/S2468-2667\(21\)00249-8](https://doi.org/10.1016/S2468-2667(21)00249-8).
3. Masters, C.L., Bateman, R., Blennow, K., Rowe, C.C., Sperling, R.A., and Cummings, J.L. (2015). Alzheimer's disease. *Nat. Rev. Dis. Primers* 1, 15056. <https://doi.org/10.1038/nrdp.2015.56>.
4. Long, J.M., and Holtzman, D.M. (2019). Alzheimer Disease: An Update on Pathobiology and Treatment Strategies. *Cell* 179, 312–339. <https://doi.org/10.1016/j.cell.2019.09.001>.
5. Scheltens, P., De Strooper, B., Kivipelto, M., Holstege, H., Chételat, G., Teunissen, C.E., Cummings, J., and van der Flier, W.M. (2021). Alzheimer's disease. *Lancet* 397, 1577–1590. [https://doi.org/10.1016/S0140-6736\(20\)32205-4](https://doi.org/10.1016/S0140-6736(20)32205-4).
6. Serrano-Pozo, A., Frosch, M.P., Masliah, E., and Hyman, B.T. (2011). Neuropathological alterations in Alzheimer disease. *Cold Spring Harb. Perspect. Med.* 1, a006189. <https://doi.org/10.1101/cshperspect.a006189>.
7. Jack, C.R., Jr., Bennett, D.A., Blennow, K., Carrillo, M.C., Dunn, B., Haeberlein, S.B., Holtzman, D.M., Jagust, W., Jessen, F., Karlawish, J., et al. (2018). NIA-AA Research Framework: Toward a biological definition of Alzheimer's disease. *Alzheimers Dement.* 14, 535–562. <https://doi.org/10.1016/j.jalz.2018.02.018>.
8. Jack, C.R., Jr., Bennett, D.A., Blennow, K., Carrillo, M.C., Feldman, H.H., Frisoni, G.B., Hampel, H., Jagust, W.J., Johnson, K.A., Knopman, D.S., et al. (2016). A/T/N: An unbiased descriptive classification scheme for Alzheimer disease biomarkers. *Neurology* 87, 539–547. <https://doi.org/10.1212/WNL.0000000000002923>.
9. Jack, C.R., Jr., Knopman, D.S., Jagust, W.J., Petersen, R.C., Weiner, M.W., Aisen, P.S., Shaw, L.M., Vemuri, P., Wiste, H.J., Weigand, S.D., et al. (2013). Tracking pathophysiological processes in Alzheimer's disease: an updated hypothetical model of dynamic biomarkers. *Lancet Neurol.* 12, 207–216. [https://doi.org/10.1016/S1474-4422\(12\)70291-0](https://doi.org/10.1016/S1474-4422(12)70291-0).
10. Dubois, B., Villain, N., Schneider, L., Fox, N., Campbell, N., Galasko, D., Kivipelto, M., Jessen, F., Hanseeuw, B., Boada, M., et al. (2024). Alzheimer Disease as a Clinical-Biological Construct—An International Working Group Recommendation. *JAMA Neurol.* 81, 1304–1311. <https://doi.org/10.1001/jamaneurol.2024.3770>.
11. Jack, C.R., Jr., Andrews, J.S., Beach, T.G., Buracchio, T., Dunn, B., Graf, A., Hansson, O., Ho, C., Jagust, W., McDade, E., et al. (2024). Revised criteria for diagnosis and staging of Alzheimer's disease: Alzheimer's Association Workgroup. *Alzheimers Dement.* 20, 5143–5169. <https://doi.org/10.1002/alz.13859>.
12. Jucker, M., and Walker, L.C. (2013). Self-propagation of pathogenic protein aggregates in neurodegenerative diseases. *Nature* 501, 45–51. <https://doi.org/10.1038/nature12481>.
13. Hanseeuw, B.J., Betensky, R.A., Jacobs, H.I.L., Schultz, A.P., Sepulcre, J., Becker, J.A., Cosio, D.M.O., Farrell, M., Quiroz, Y.T., Mormino, E.C., et al. (2019). Association of Amyloid and Tau With Cognition in Preclinical Alzheimer Disease: A Longitudinal Study. *JAMA Neurol.* 76, 915–924. <https://doi.org/10.1001/jamaneurol.2019.1424>.
14. Hardy, J., and Selkoe, D.J. (2002). The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. *Science* 297, 353–356. <https://doi.org/10.1126/science.1072994>.
15. Brunden, K.R., Trojanowski, J.Q., and Lee, V.M.Y. (2009). Advances in tau-focused drug discovery for Alzheimer's disease and related tauopathies. *Nat. Rev. Drug Discov.* 8, 783–793. <https://doi.org/10.1038/nrd2959>.
16. Lee, V.M., Goedert, M., and Trojanowski, J.Q. (2001). Neurodegenerative tauopathies. *Annu. Rev. Neurosci.* 24, 1121–1159. <https://doi.org/10.1146/annurev.neuro.24.1.1121>.
17. Li, C., and Götz, J. (2017). Tau-based therapies in neurodegeneration: opportunities and challenges. *Nat. Rev. Drug Discov.* 16, 863–883. <https://doi.org/10.1038/nrd.2017.155>.
18. Hyman, B.T., Phelps, C.H., Beach, T.G., Bigio, E.H., Cairns, N.J., Carrillo, M.C., Dickson, D.W., Duyckaerts, C., Frosch, M.P., Masliah, E., et al. (2012). National Institute on Aging-Alzheimer's Association guidelines for the neuropathologic assessment of Alzheimer's disease. *Alzheimers Dement.* 8, 1–13. <https://doi.org/10.1016/j.jalz.2011.10.007>.
19. Wilson, D.M., 3rd, Cookson, M.R., Van Den Bosch, L., Zetterberg, H., Holtzman, D.M., and Dewachter, I. (2023). Hallmarks of neurodegenerative diseases. *Cell* 186, 693–714. <https://doi.org/10.1016/j.cell.2022.12.032>.
20. Arriagada, P.V., Growdon, J.H., Hedley-Whyte, E.T., and Hyman, B.T. (1992). Neurofibrillary tangles but not senile plaques parallel duration and severity of Alzheimer's disease. *Neurology* 42, 631–639. <https://doi.org/10.1212/WNL.42.3.631>.
21. Braak, H., and Braak, E. (1991). Neuropathological staging of Alzheimer-related changes. *Acta Neuropathol.* 82, 239–259. <https://doi.org/10.1007/BF00308809>.
22. Ossenkoppele, R., Schonhaut, D.R., Schöll, M., Lockhart, S.N., Ayakta, N., Baker, S.L., O'Neil, J.P., Janabi, M., Lazaris, A., Cantwell, A., et al. (2016). Tau PET patterns mirror clinical and neuroanatomical variability in Alzheimer's disease. *Brain* 139, 1551–1567. <https://doi.org/10.1093/brain/aww027>.
23. Therriault, J., Schindler, S.E., Salvadó, G., Pascoal, T.A., Benedet, A.L., Ashton, N.J., Karikari, T.K., Apostolova, L., Murray, M.E., Verberk, I., et al. (2024). Biomarker-based staging of Alzheimer disease: rationale and clinical applications. *Nat. Rev. Neurol.* 20, 232–244. <https://doi.org/10.1038/s41582-024-00942-2>.
24. Villemagne, V.L., Doré, V., Burnham, S.C., Masters, C.L., and Rowe, C.C. (2018). Imaging tau and amyloid-beta proteinopathies in Alzheimer disease and other conditions. *Nat. Rev. Neurol.* 14, 225–236. <https://doi.org/10.1038/nrneuro.2018.9>.
25. Stamelou, M., Respondek, G., Giagkou, N., Whitwell, J.L., Kovacs, G.G., and Högländer, G.U. (2021). Evolving concepts in progressive supranuclear palsy and other 4-repeat tauopathies. *Nat. Rev. Neurol.* 17, 601–620. <https://doi.org/10.1038/s41582-021-00541-5>.
26. Zhang, Y., Wu, K.M., Yang, L., Dong, Q., and Yu, J.T. (2022). Tauopathies: new perspectives and challenges. *Mol. Neurodegener.* 17, 28. <https://doi.org/10.1186/s13024-022-00533-z>.
27. Congdon, E.E., Ji, C., Tetlow, A.M., Jiang, Y., and Sigurdsson, E.M. (2023). Tau-targeting therapies for Alzheimer disease: current status and future directions. *Nat. Rev. Neurol.* 19, 715–736. <https://doi.org/10.1038/s41582-023-00883-2>.
28. Parra Bravo, C., Naguib, S.A., and Gan, L. (2024). Cellular and pathological functions of tau. *Nat. Rev. Mol. Cell Biol.* 25, 845–864. <https://doi.org/10.1038/s41580-024-00753-9>.
29. Wang, Y., and Mandelkow, E. (2016). Tau in physiology and pathology. *Nat. Rev. Neurosci.* 17, 5–21. <https://doi.org/10.1038/nrn.2015.1>.
30. Franzmeier, N., Brendel, M., Beyer, L., Slemann, L., Kovacs, G.G., Arzberger, T., Kurz, C., Respondek, G., Lukic, M.J., Biel, D., et al. (2022). Tau deposition patterns are associated with functional connectivity in primary tauopathies. *Nat. Commun.* 13, 1362. <https://doi.org/10.1038/s41467-022-28896-3>.
31. Kovacs, G.G., Lukic, M.J., Irwin, D.J., Arzberger, T., Respondek, G., Lee, E.B., Coughlin, D., Giese, A., Grossman, M., Kurz, C., et al. (2020). Distribution patterns of tau pathology in progressive supranuclear palsy. *Acta Neuropathol.* 140, 99–119. <https://doi.org/10.1007/s00401-020-02158-2>.
32. Haass, C., and Selkoe, D. (2022). If amyloid drives Alzheimer disease, why have anti-amyloid therapies not yet slowed cognitive decline? *PLoS Biol.* 20, e3001694. <https://doi.org/10.1371/journal.pbio.3001694>.

33. Boxer, A.L., and Sperling, R. (2023). Accelerating Alzheimer's therapeutic development: The past and future of clinical trials. *Cell* 186, 4757–4772. <https://doi.org/10.1016/j.cell.2023.09.023>.
34. Jucker, M., and Walker, L.C. (2023). Alzheimer's disease: From immunotherapy to immunoprevention. *Cell* 186, 4260–4270. <https://doi.org/10.1016/j.cell.2023.08.021>.
35. Benilova, I., Karan, E., and De Strooper, B. (2012). The toxic A β oligomer and Alzheimer's disease: an emperor in need of clothes. *Nat. Neurosci.* 15, 349–357. <https://doi.org/10.1038/nn.3028>.
36. Karan, E., and De Strooper, B. (2022). The amyloid hypothesis in Alzheimer disease: new insights from new therapeutics. *Nat. Rev. Drug Discov.* 21, 306–318. <https://doi.org/10.1038/s41573-022-00391-w>.
37. St George-Hyslop, P.H., and Morris, J.C. (2008). Will anti-amyloid therapies work for Alzheimer's disease? *Lancet* 372, 180–182. [https://doi.org/10.1016/S0140-6736\(08\)61047-8](https://doi.org/10.1016/S0140-6736(08)61047-8).
38. De Strooper, B., Vassar, R., and Golde, T. (2010). The secretases: enzymes with therapeutic potential in Alzheimer disease. *Nat. Rev. Neurol.* 6, 99–107. <https://doi.org/10.1038/nrneurol.2009.218>.
39. Holmes, C., Boche, D., Wilkinson, D., Yadegarfar, G., Hopkins, V., Bayer, A., Jones, R.W., Bullock, R., Love, S., Neal, J.W., et al. (2008). Long-term effects of Abeta42 immunisation in Alzheimer's disease: follow-up of a randomised, placebo-controlled phase I trial. *Lancet* 372, 216–223. [https://doi.org/10.1016/S0140-6736\(08\)61075-2](https://doi.org/10.1016/S0140-6736(08)61075-2).
40. Sims, J.R., Zimmer, J.A., Evans, C.D., Lu, M., Ardayfio, P., Sparks, J., Wessels, A.M., Shcherbinin, S., Wang, H., Monkul Nery, E.S., et al. (2023). Donanemab in Early Symptomatic Alzheimer Disease: The TRAILBLAZER-ALZ 2 Randomized Clinical Trial. *JAMA* 330, 512–527. <https://doi.org/10.1001/jama.2023.13239>.
41. Van Dyck, C.H., Swanson, C.J., Aisen, P., Bateman, R.J., Chen, C., Gee, M., Kanekyo, M., Li, D., Reyderman, L., Cohen, S., et al. (2023). Lecanemab in early Alzheimer's disease. *N. Engl. J. Med.* 388, 9–21. <https://doi.org/10.1056/NEJMoa2212948>.
42. Sevigny, J., Chiao, P., Bussière, T., Weinreb, P.H., Williams, L., Maier, M., Dunstan, R., Salloway, S., Chen, T., Ling, Y., et al. (2016). The antibody aducanumab reduces Abeta plaques in Alzheimer's disease. *Nature* 537, 50–56. <https://doi.org/10.1038/nature19323>.
43. Schenk, D., Barbour, R., Dunn, W., Gordon, G., Grajeda, H., Guido, T., Hu, K., Huang, J., Johnson-Wood, K., Khan, K., et al. (1999). Immunization with amyloid-beta attenuates Alzheimer-disease-like pathology in the PDAPP mouse. *Nature* 400, 173–177. <https://doi.org/10.1038/22124>.
44. Doody, R.S., Thomas, R.G., Farlow, M., Iwatsubo, T., Vellas, B., Joffe, S., Kieburtz, K., Raman, R., Sun, X., Aisen, P.S., et al. (2014). Phase 3 trials of solanezumab for mild-to-moderate Alzheimer's disease. *N. Engl. J. Med.* 370, 311–321. <https://doi.org/10.1056/NEJMoa1312889>.
45. Salloway, S., Sperling, R., and Brashears, H.R. (2014). Phase 3 trials of solanezumab and bapineuzumab for Alzheimer's disease. *N. Engl. J. Med.* 370, 1460. <https://doi.org/10.1056/NEJMc1402193>.
46. Orgogozo, J.M., Gilman, S., Dartigues, J.F., Laurent, B., Puel, M., Kirby, L.C., Jouanny, P., Dubois, B., Eisner, L., Flitman, S., et al. (2003). Subacute meningoencephalitis in a subset of patients with AD after Abeta42 immunization. *Neurology* 61, 46–54. <https://doi.org/10.1212/01.wnl.0000073623.84147.a8>.
47. Hock, C., Konietzko, U., Streffer, J.R., Tracy, J., Signorell, A., Müller-Tillmanns, B., Lemke, U., Henke, K., Moritz, E., Garcia, E., et al. (2003). Antibodies against beta-amyloid slow cognitive decline in Alzheimer's disease. *Neuron* 38, 547–554. [https://doi.org/10.1016/s0896-6273\(03\)00294-0](https://doi.org/10.1016/s0896-6273(03)00294-0).
48. Brody, D.L., and Holtzman, D.M. (2008). Active and passive immunotherapy for neurodegenerative disorders. *Annu. Rev. Neurosci.* 31, 175–193. <https://doi.org/10.1146/annurev.neuro.31.060407.125529>.
49. Blennow, K., Chen, C., Cicognola, C., Wildsmith, K.R., Manser, P.T., Borhorquez, S.M.S., Zhang, Z., Xie, B., Peng, J., Hansson, O., et al. (2020). Cerebrospinal fluid tau fragment correlates with tau PET: a candidate biomarker for tangle pathology. *Brain* 143, 650–660. <https://doi.org/10.1093/brain/awz346>.
50. Jack, C.R., Jr., Wiste, H.J., Schwarz, C.G., Lowe, V.J., Senjem, M.L., Vemuri, P., Weigand, S.D., Therneau, T.M., Knopman, D.S., Gunter, J.L., et al. (2018). Longitudinal tau PET in ageing and Alzheimer's disease. *Brain* 141, 1517–1528. <https://doi.org/10.1093/brain/awy059>.
51. Schoonhoven, D.N., Coomans, E.M., Millán, A.P., van Nifterick, A.M., Visser, D., Ossenkoppele, R., Tuncel, H., van der Flier, W.M., Golla, S.S.V., Scheltens, P., et al. (2023). Tau protein spreads through functionally connected neurons in Alzheimer's disease: a combined MEG/PET study. *Brain* 146, 4040–4054. <https://doi.org/10.1093/brain/awad189>.
52. Schwarz, A.J., Yu, P., Miller, B.B., Shcherbinin, S., Dickson, J., Navitsky, M., Joshi, A.D., Devous, M.D., and Mintun, M.S. (2016). Regional profiles of the candidate tau PET ligand 18F-AV-1451 recapitulate key features of Braak histopathological stages. *Brain* 139, 1539–1550. <https://doi.org/10.1093/brain/aww023>.
53. Olsson, B., Lautner, R., Andreasson, U., Öhrfelt, A., Portelius, E., Bjerke, M., Hölttä, M., Rosén, C., Olsson, C., Strobel, G., et al. (2016). CSF and blood biomarkers for the diagnosis of Alzheimer's disease: a systematic review and meta-analysis. *Lancet Neurol.* 15, 673–684. [https://doi.org/10.1016/S1474-4422\(16\)00070-3](https://doi.org/10.1016/S1474-4422(16)00070-3).
54. Ossenkoppele, R., van der Kant, R., and Hansson, O. (2022). Tau biomarkers in Alzheimer's disease: towards implementation in clinical practice and trials. *Lancet Neurol.* 21, 726–734. [https://doi.org/10.1016/S1474-4422\(22\)00168-5](https://doi.org/10.1016/S1474-4422(22)00168-5).
55. Zetterberg, H., and Blennow, K. (2021). Moving fluid biomarkers for Alzheimer's disease from research tools to routine clinical diagnostics. *Mol. Neurodegener.* 16, 10. <https://doi.org/10.1186/s13024-021-00430-x>.
56. Shaw, L.M., Vanderstichele, H., Knapik-Czajka, M., Clark, C.M., Aisen, P.S., Petersen, R.C., Blennow, K., Soares, H., Simon, A., Lewczuk, P., et al. (2009). Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. *Ann. Neurol.* 65, 403–413. <https://doi.org/10.1002/ana.21610>.
57. Clark, C.M., Schneider, J.A., Bedell, B.J., Beach, T.G., Bilker, W.B., Minutun, M.A., Pontecorvo, M.J., Hefti, F., Carpenter, A.P., Flitter, M.L., et al. (2011). Use of florbetapir-PET for imaging beta-amyloid pathology. *JAMA* 305, 275–283. <https://doi.org/10.1001/jama.2010.2008>.
58. Horie, K., Salvadó, G., Koppisetti, R.K., Janelidze, S., Barthélémy, N.R., He, Y., Sato, C., Gordon, B.A., Jiang, H., Benzinger, T.L.S., et al. (2025). Plasma MTBR-tau243 biomarker identifies tau tangle pathology in Alzheimer's disease. *Nat. Med.* 31, 2044–2053. <https://doi.org/10.1038/s41591-025-03617-7>.
59. Teunissen, C.E., Kolster, R., Triana-Baltzer, G., Janelidze, S., Zetterberg, H., and Kolb, H.C. (2025). Plasma p-tau immunoassays in clinical research for Alzheimer's disease. *Alzheimers Dement.* 21, e14397. <https://doi.org/10.1002/alz.14397>.
60. Bateman, R.J., Xiong, C., Benzinger, T.L.S., Fagan, A.M., Goate, A., Fox, N.C., Marcus, D.S., Cairns, N.J., Xie, X., Blazey, T.M., et al. (2012). Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N. Engl. J. Med.* 367, 795–804. <https://doi.org/10.1056/NEJMoa1202753>.
61. Frost, B., Rowe, J.B., Akinyemi, R.O., Abisambra, J.F., Ashton, N.J., Brendel, M., Buée, L., Butler, D., Carrillo, M.C., Chung, P., et al. (2025). Insights into pathophysiology, biomarkers, and therapeutics in tauopathies: proceedings of the Tau2024 Global Conference. *Alzheimers Dement.* 21, e70078. <https://doi.org/10.1002/alz.70078>.
62. Horie, K., Barthélémy, N.R., Sato, C., and Bateman, R.J. (2021). CSF tau microtubule binding region identifies tau tangle and clinical stages of Alzheimer's disease. *Brain* 144, 515–527. <https://doi.org/10.1093/brain/awaa373>.
63. Horie, K., Salvadó, G., Barthélémy, N.R., Janelidze, S., Li, Y., He, Y., Saef, B., Chen, C.D., Jiang, H., Strandberg, O., et al. (2023). CSF MTBR-tau243 is a specific biomarker of tau tangle pathology in

Alzheimer's disease. *Nat. Med.* 29, 1954–1963. <https://doi.org/10.1038/s41591-023-02443-z>.

64. Goedert, M., Clavaguera, F., and Tolnay, M. (2010). The propagation of prion-like protein inclusions in neurodegenerative diseases. *Trends Neurosci.* 33, 317–325. <https://doi.org/10.1016/j.tins.2010.04.003>.
65. Goedert, M., Eisenberg, D.S., and Crowther, R.A. (2017). Propagation of Tau Aggregates and Neurodegeneration. *Annu. Rev. Neurosci.* 40, 189–210. <https://doi.org/10.1146/annurev-neuro-072116-031153>.
66. Jucker, M., and Walker, L.C. (2018). Propagation and spread of pathogenic protein assemblies in neurodegenerative diseases. *Nat. Neurosci.* 21, 1341–1349. <https://doi.org/10.1038/s41593-018-0238-6>.
67. Ballatore, C., Lee, V.M.Y., and Trojanowski, J.Q. (2007). Tau-mediated neurodegeneration in Alzheimer's disease and related disorders. *Nat. Rev. Neurosci.* 8, 663–672. <https://doi.org/10.1038/nrn2194>.
68. Colin, M., Dujardin, S., Schraen-Maschke, S., Meno-Tetang, G., Duyckaerts, C., Courade, J.P., and Buée, L. (2020). From the prion-like propagation hypothesis to therapeutic strategies of anti-tau immunotherapy. *Acta Neuropathol.* 139, 3–25. <https://doi.org/10.1007/s00401-019-02087-9>.
69. Shi, Y., Zhang, W., Yang, Y., Murzin, A.G., Falcon, B., Kotecha, A., van Beers, M., Tarutani, A., Kometani, F., Garringer, H.J., et al. (2021). Structure-based classification of tauopathies. *Nature* 598, 359–363. <https://doi.org/10.1038/s41586-021-03911-7>.
70. Cummings, J.L., Zhou, Y., Lee, G., Zhong, K., Fonseca, J., Leisgang-Osse, A.M., and Cheng, F. (2025). Alzheimer's disease drug development pipeline: 2025. *Alzheimers Dement. (N Y)* 11, e70098. <https://doi.org/10.1002/trc2.70098>.
71. Qi, C., Lövestam, S., Murzin, A.G., Peak-Chew, S., Franco, C., Bogdani, M., Latimer, C., Murrell, J.R., Cullinane, P.W., Jaunmuktane, Z., et al. (2025). Tau filaments with the Alzheimer fold in human MAPT mutants V337M and R406W. *Nat. Struct. Mol. Biol.* 32, 1297–1304. <https://doi.org/10.1038/s41594-025-01498-5>.
72. Scheres, S.H.W., Ryskeldi-Falcon, B., and Goedert, M. (2023). Molecular pathology of neurodegenerative diseases by cryo-EM of amyloids. *Nature* 621, 701–710. <https://doi.org/10.1038/s41586-023-06437-2>.
73. Goedert, M. (2015). NEURODEGENERATION. Alzheimer's and Parkinson's diseases: The prion concept in relation to assembled Abeta, tau, and alpha-synuclein. *Science* 349, 1255555. <https://doi.org/10.1126/science.1255555>.
74. Stancu, I.C., Vasconcelos, B., Ris, L., Wang, P., Villers, A., Peeraer, E., Buist, A., Terwel, D., Baatsen, P., Oyelami, T., et al. (2015). Templated misfolding of Tau by prion-like seeding along neuronal connections impairs neuronal network function and associated behavioral outcomes in Tau transgenic mice. *Acta Neuropathol.* 129, 875–894. <https://doi.org/10.1007/s00401-015-1413-4>.
75. Clavaguera, F., Bolmont, T., Crowther, R.A., Abramowski, D., Frank, S., Probst, A., Fraser, G., Stalder, A.K., Beibel, M., Staufenbiel, M., et al. (2009). Transmission and spreading of tauopathy in transgenic mouse brain. *Nat. Cell Biol.* 11, 909–913. <https://doi.org/10.1038/ncb1901>.
76. Kaufman, S.K., Sanders, D.W., Thomas, T.L., Ruchinskas, A.J., Vaquer-Alicea, J., Sharma, A.M., Miller, T.M., and Diamond, M.I. (2016). Tau Prion Strains Dictate Patterns of Cell Pathology, Progression Rate, and Regional Vulnerability In Vivo. *Neuron* 92, 796–812. <https://doi.org/10.1016/j.neuron.2016.09.055>.
77. Sanders, D.W., Kaufman, S.K., DeVos, S.L., Sharma, A.M., Mirbaha, H., Li, A., Barker, S.J., Foley, A.C., Thorpe, J.R., Serpell, L.C., et al. (2014). Distinct tau prion strains propagate in cells and mice and define different tauopathies. *Neuron* 82, 1271–1288. <https://doi.org/10.1016/j.neuron.2014.04.047>.
78. Dujardin, S., Commins, C., Lathuiliere, A., Beerepoot, P., Fernandes, A.R., Kamath, T.V., De Los Santos, M.B., Klickstein, N., Corjuc, D.L., Corjuc, B.T., et al. (2020). Tau molecular diversity contributes to clinical heterogeneity in Alzheimer's disease. *Nat. Med.* 26, 1256–1263. <https://doi.org/10.1038/s41591-020-0938-9>.
79. Mudher, A., Colin, M., Dujardin, S., Medina, M., Dewachter, I., Alavi Naini, S.M., Mandelkow, E.M., Mandelkow, E., Buée, L., Goedert, M., et al. (2017). What is the evidence that tau pathology spreads through prion-like propagation? *Acta Neuropathol. Commun.* 5, 99. <https://doi.org/10.1186/s40478-017-0488-7>.
80. Sato, C., Barthélémy, N.R., Mawuenyega, K.G., Patterson, B.W., Gordon, B.A., Jockel-Balsarotti, J., Sullivan, M., Crisp, M.J., Kasten, T., Kirmess, K.M., et al. (2018). Tau Kinetics in Neurons and the Human Central Nervous System. *Neuron* 97, 1284–1298.e7. <https://doi.org/10.1016/j.neuron.2018.02.015>.
81. Lasagna-Reeves, C.A., Castillo-Carranza, D.L., Guerrero-Muoz, M.J., Jackson, G.R., and Kayed, R. (2010). Preparation and characterization of neurotoxic tau oligomers. *Biochemistry* 49, 10039–10041. <https://doi.org/10.1021/bi1016233>.
82. Lasagna-Reeves, C.A., Castillo-Carranza, D.L., SenGupta, U., Clos, A.L., Jackson, G.R., and Kayed, R. (2011). Tau oligomers impair memory and induce synaptic and mitochondrial dysfunction in wild-type mice. *Mol. Neurodegener.* 6, 39. <https://doi.org/10.1186/1750-1326-6-39>.
83. Cummings, J., Lee, G., Mortsdorf, T., Ritter, A., and Zhong, K. (2017). Alzheimer's disease drug development pipeline: 2017. *Alzheimers Dement. (N Y)* 3, 367–384. <https://doi.org/10.1016/j.trci.2017.05.002>.
84. Cummings, J., Zhou, Y., Lee, G., Zhong, K., Fonseca, J., and Cheng, F. (2023). Alzheimer's disease drug development pipeline: 2023. *Alzheimers Dement. (N Y)* 9, e12385. <https://doi.org/10.1002/trc2.12385>.
85. Jabbari, E., and Duff, K.E. (2021). Tau-targeting antibody therapies: too late, wrong epitope or wrong target? *Nat. Med.* 27, 1341–1342. <https://doi.org/10.1038/s41591-021-01465-9>.
86. Congdon, E.E., and Sigurdsson, E.M. (2018). Tau-targeting therapies for Alzheimer disease. *Nat. Rev. Neurol.* 14, 399–415. <https://doi.org/10.1038/s41582-018-0013-z>.
87. DeVos, S.L., Miller, R.L., Schoch, K.M., Holmes, B.B., Kebodeaux, C.S., Wegener, A.J., Chen, G., Shen, T., Tran, H., Nichols, B., et al. (2017). Tau reduction prevents neuronal loss and reverses pathological tau deposition and seeding in mice with tauopathy. *Sci. Transl. Med.* 9, eaag0481. <https://doi.org/10.1126/scitranslmed.aag0481>.
88. DeVos, S.L., Goncharoff, D.K., Chen, G., Kebodeaux, C.S., Yamada, K., Stewart, F.R., Schuler, D.R., Maloney, S.E., Wozniak, D.F., Rigo, F., et al. (2013). Antisense reduction of tau in adult mice protects against seizures. *J. Neurosci.* 33, 12887–12897. <https://doi.org/10.1523/JNEUROSCI.2107-13.2013>.
89. Xu, H., Rösler, T.W., Carlsson, T., de Andrade, A., Fiala, O., Hollerage, M., Oertel, W.H., Goedert, M., Aigner, A., and Höglinder, G.U. (2014). Tau silencing by siRNA in the P301S mouse model of tauopathy. *Curr. Gene Ther.* 14, 343–351. <https://doi.org/10.2174/156652321405140926160602>.
90. Smith, S.E., McCoy-Gross, K., Malcolm, A., Oranski, J., Markway, J.W., Miller, T.M., and Bucelli, R.C. (2025). Tofersen treatment leads to sustained stabilization of disease in SOD1 ALS in a "real-world" setting. *Ann. Clin. Transl. Neurol.* 12, 311–319. <https://doi.org/10.1002/acn3.52264>.
91. FDA. Tofersen. <https://www.fda.gov/drugs/news-events-human-drugs/fda-approves-treatment-amyotrophic-lateral-sclerosis-associated-mutation-sod1-gene>.
92. Kaiser, J. (2025). Kaiser J. In a first, a gene therapy seems to slow Huntington disease. *Science* 390, 15. <https://doi.org/10.1126/science.aec7429>.
93. Wadman, M. (2016). Antisense rescues babies from killer disease. *Science* 354, 1359–1360. <https://doi.org/10.1126/science.354.6318.1359>.
94. Finkel, R.S., Mercuri, E., Darras, B.T., Connolly, A.M., Kuntz, N.L., Kirschner, J., Chiriboga, C.A., Saito, K., Servais, L., Tizzano, E., et al. (2017). Nusinersen versus Sham Control in Infantile-Onset Spinal Muscular

Atrophy. *N. Engl. J. Med.* 377, 1723–1732. <https://doi.org/10.1056/NEJMoa1702752>.

95. Edwards, A.L., Collins, J.A., Junge, C., Kordasiewicz, H., Mignon, L., Wu, S., Li, Y., Lin, L., DuBois, J., Hutchison, R.M., et al. (2023). Exploratory Tau Biomarker Results From a Multiple Ascending-Dose Study of BIIB080 in Alzheimer Disease: A Randomized Clinical Trial. *JAMA Neurol.* 80, 1344–1352. <https://doi.org/10.1001/jamaneurol.2023.3861>.

96. Mummery, C.J., Börjesson-Hanson, A., Blackburn, D.J., Vijverberg, E.G.B., De Deyn, P.P., Ducharme, S., Jonsson, M., Schneider, A., Rinne, J.O., Ludolph, A.C., et al. (2023). Tau-targeting antisense oligonucleotide MAPT(Rx) in mild Alzheimer's disease: a phase 1b, randomized, placebo-controlled trial. *Nat. Med.* 29, 1437–1447. <https://doi.org/10.1038/s41591-023-02326-3>.

97. Harris, G.A., and Hirschfeld, L.R. (2025). Antisense oligonucleotides provide optimism to the therapeutic landscape for tauopathies. *Neural Regen. Res.* 20, 803–804. <https://doi.org/10.4103/NRR.NRR-D-23-02057>.

98. Mullan, A. (2024). Anti-tau antibody stumbles in phase II Alzheimer trial. *Nat. Rev. Drug Discov.* 23, 883. <https://doi.org/10.1038/d41573-024-00180-7>.

99. Alzforum (2024). Finally, Therapeutic Antibodies Start to Reduce Tangles. <https://www.alzforum.org/news/conference-coverage/finally-therapeutic-antibodies-start-reduce-tangles>.

100. Alzforum. Bepranemab 1. <https://www.alzforum.org/therapeutics/bepranemab>.

101. UCB (2024). UCB Presents Encouraging Data on Bepranemab in Early Alzheimer's Disease in Phase 2a Study at CTAD 2024. <https://www.ucb.com/newsroom/press-releases/article/ucb-presents-encouraging-data-on-bepranemab-in-early-alzheimer-s-disease-in-phase-2a-study-at-ctad-2024>.

102. Asumi, A.A., Boutajangout, A., Quartermain, D., and Sigurdsson, E.M. (2007). Immunotherapy targeting pathological tau conformers in a tangle mouse model reduces brain pathology with associated functional improvements. *J. Neurosci.* 27, 9115–9129. <https://doi.org/10.1523/JNEUROSCI.2361-07.2007>.

103. Courade, J.P., Angers, R., Mairet-Coello, G., Pacico, N., Tyson, K., Lightwood, D., Munro, R., McMillan, D., Griffin, R., Baker, T., et al. (2018). Epitope determines efficacy of therapeutic anti-Tau antibodies in a functional assay with human Alzheimer Tau. *Acta Neuropathol.* 136, 729–745. <https://doi.org/10.1007/s00401-018-1911-2>.

104. Gaikwad, S., Puangmalai, N., Sonawane, M., Montalbano, M., Price, R., Iyer, M.S., Ray, A., Moreno, S., and Kayed, R. (2024). Nasal tau immunotherapy clears intracellular tau pathology and improves cognitive functions in aged tauopathy mice. *Sci. Transl. Med.* 16, eadj5958. <https://doi.org/10.1126/scitranslmed.adj5958>.

105. Ayalon, G., Lee, S.H., Adolfsson, O., Foo-Atkins, C., Atwal, J.K., Blendstrup, M., Booler, H., Bravo, J., Brendza, R., Brunstein, F., et al. (2021). Antibody semorinemab reduces tau pathology in a transgenic mouse model and engages tau in patients with Alzheimer's disease. *Sci. Transl. Med.* 13, eabb2639. <https://doi.org/10.1126/scitranslmed.eabb2639>.

106. Boutajangout, A., Ingadottir, J., Davies, P., and Sigurdsson, E.M. (2011). Passive immunization targeting pathological phospho-tau protein in a mouse model reduces functional decline and clears tau aggregates from the brain. *J. Neurochem.* 118, 658–667. <https://doi.org/10.1111/j.1471-4159.2011.07337.x>.

107. Albert, M., Mairet-Coello, G., Danis, C., Lieger, S., Caillierez, R., Carrier, S., Skrobala, E., Landrieu, I., Michel, A., Schmitt, M., et al. (2019). Prevention of tau seeding and propagation by immunotherapy with a central tau epitope antibody. *Brain* 142, 1736–1750. <https://doi.org/10.1093/brain/awz100>.

108. Bittar, A., Bhatt, N., and Kayed, R. (2020). Advances and considerations in AD tau-targeted immunotherapy. *Neurobiol. Dis.* 134, 104707. <https://doi.org/10.1016/j.nbd.2019.104707>.

109. Yanamandra, K., Kfouri, N., Jiang, H., Mahan, T.E., Ma, S., Maloney, S.E., Wozniak, D.F., Diamond, M.I., and Holtzman, D.M. (2013). Anti-tau antibodies that block tau aggregate seeding in vitro markedly decrease pathology and improve cognition in vivo. *Neuron* 80, 402–414. <https://doi.org/10.1016/j.neuron.2013.07.046>.

110. Geerts, H., Bergeler, S., Walker, M., van der Graaf, P.H., and Courade, J.P. (2023). Analysis of clinical failure of anti-tau and anti-synuclein antibodies in neurodegeneration using a quantitative systems pharmacology model. *Sci. Rep.* 13, 14342. <https://doi.org/10.1038/s41598-023-41382-0>.

111. Dam, T., Boxer, A.L., Golbe, L.I., Höglinder, G.U., Morris, H.R., Litvan, I., Lang, A.E., Corvol, J.C., Aiba, I., Grundman, M., et al. (2021). Safety and efficacy of anti-tau monoclonal antibody gosuranemab in progressive supranuclear palsy: a phase 2, randomized, placebo-controlled trial. *Nat. Med.* 27, 1451–1457. <https://doi.org/10.1038/s41591-021-01455-x>.

112. Teng, E., Manser, P.T., Pickthorn, K., Brunstein, F., Blendstrup, M., Sanabria Bohorquez, S., Wildsmith, K.R., Toth, B., Dolton, M., Ramakrishnan, V., et al. (2022). Safety and Efficacy of Semorinemab in Individuals With Prodromal to Mild Alzheimer Disease: A Randomized Clinical Trial. *JAMA Neurol.* 79, 758–767. <https://doi.org/10.1001/jamaneurol.2022.1375>.

113. Höglinder, G.U., Litvan, I., Mendonca, N., Wang, D., Zheng, H., Rendebach-Mueller, B., Lon, H.K., Jin, Z., Fisseha, N., Budur, K., et al. (2021). Safety and efficacy of tilavonemab in progressive supranuclear palsy: a phase 2, randomised, placebo-controlled trial. *Lancet Neurol.* 20, 182–192. [https://doi.org/10.1016/S1474-4422\(20\)30489-0](https://doi.org/10.1016/S1474-4422(20)30489-0).

114. Lee, S.H., Le Pichon, C.E., Adolfsson, O., Gafner, V., Pihlgren, M., Lin, H., Solanoy, H., Brendza, R., Ngu, H., Foreman, O., et al. (2016). Antibody-Mediated Targeting of Tau In Vivo Does Not Require Effector Function and Microglial Engagement. *Cell Rep.* 16, 1690–1700. <https://doi.org/10.1016/j.celrep.2016.06.099>.

115. Congdon, E.E., Jiang, Y., and Sigurdsson, E.M. (2022). Targeting tau only extracellularly is likely to be less efficacious than targeting it both intra- and extracellularly. *Semin. Cell Dev. Biol.* 126, 125–137. <https://doi.org/10.1016/j.semcdb.2021.12.002>.

116. Qureshi, I.A., Tirucherai, G., Ahlijanian, M.K., Kolaitis, G., Bechtold, C., and Grundman, M. (2018). A randomized, single ascending dose study of intravenous BIIB092 in healthy participants. *Alzheimers Dement. (N Y)* 4, 746–755. <https://doi.org/10.1016/j.jtrc.2018.10.007>.

117. Alam, R., Driver, D., Wu, S., Lozano, E., Key, S.L., Hole, J.T., Hayashi, M.L., and Lu, J. (2017). O2-14-05: PRECLINICAL CHARACTERIZATION OF AN ANTIBODY [LY3303560] TARGETING AGGREGATED TAU. *Alzheimers Dement.* 13, P592–P593. <https://doi.org/10.1016/j.jalz.2017.07.227>.

118. Fleisher, A.S., Munsie, L.M., Perahia, D.G.S., Andersen, S.W., Higgins, I.A., Hauck, P.M., Lo, A.C., Sims, J.R., Brys, M., Minturn, M., et al. (2024). Assessment of Efficacy and Safety of Zagotenemab: Results From PERISCOPE-ALZ, a Phase 2 Study in Early Symptomatic Alzheimer Disease. *Neurology* 102, e208061. <https://doi.org/10.1212/WNL.00000000208061>.

119. Barthélémy, N.R., Gabelle, A., Hirtz, C., Fenaille, F., Sergeant, N., Schraen-Maschke, S., Vialaret, J., Buée, L., Junot, C., Becher, F., et al. (2016). Differential Mass Spectrometry Profiles of Tau Protein in the Cerebrospinal Fluid of Patients with Alzheimer's Disease, Progressive Supranuclear Palsy, and Dementia with Lewy Bodies. *J. Alzheimers Dis.* 51, 1033–1043. <https://doi.org/10.3233/JAD-150962>.

120. Amadoro, G., Latina, V., Corsetti, V., and Calissano, P. (2020). N-terminal tau truncation in the pathogenesis of Alzheimer's disease (AD): Developing a novel diagnostic and therapeutic approach. *Biochim. Biophys. Acta Mol. Basis Dis.* 1866, 165584. <https://doi.org/10.1016/j.bbadi.2019.165584>.

121. Quinn, J.P., Corbett, N.J., Kellett, K.A.B., and Hooper, N.M. (2018). Tau Proteolysis in the Pathogenesis of Tauopathies: Neurotoxic Fragments and Novel Biomarkers. *J. Alzheimers Dis.* 63, 13–33. <https://doi.org/10.3233/JAD-170959>.

122. Guo, J.L., and Lee, V.M.Y. (2011). Seeding of normal Tau by pathological Tau conformers drives pathogenesis of Alzheimer-like tangles. *J. Biol. Chem.* 286, 15317–15331. <https://doi.org/10.1074/jbc.M110.209296>.

123. Iba, M., Guo, J.L., McBride, J.D., Zhang, B., Trojanowski, J.Q., and Lee, V.M.Y. (2013). Synthetic tau fibrils mediate transmission of neurofibrillary tangles in a transgenic mouse model of Alzheimer's-like tauopathy. *J. Neurosci.* 33, 1024–1037. <https://doi.org/10.1523/JNEUROSCI.2642-12.2013>.

124. Fitzpatrick, A.W.P., Falcon, B., He, S., Murzin, A.G., Murshudov, G., Garginer, H.J., Crowther, R.A., Ghetti, B., Goedert, M., and Scheres, S.H.W. (2017). Cryo-EM structures of tau filaments from Alzheimer's disease. *Nature* 547, 185–190. <https://doi.org/10.1038/nature23002>.

125. Falcon, B., Cavallini, A., Angers, R., Glover, S., Murray, T.K., Barnham, L., Jackson, S., O'Neill, M.J., Isaacs, A.M., Hutton, M.L., et al. (2015). Conformation determines the seeding potencies of native and recombinant Tau aggregates. *J. Biol. Chem.* 290, 1049–1065. <https://doi.org/10.1074/jbc.M114.589309>.

126. Nobuhara, C.K., DeVos, S.L., Commins, C., Wegmann, S., Moore, B.D., Roe, A.D., Costantino, I., Frosch, M.P., Pitstick, R., Carlson, G.A., et al. (2017). Tau Antibody Targeting Pathological Species Blocks Neuronal Uptake and Interneuron Propagation of Tau in Vitro. *Am. J. Pathol.* 187, 1399–1412. <https://doi.org/10.1016/j.ajpath.2017.01.022>.

127. Fukumoto, H., Kao, T.H., Tai, C.Y., Jang, M.K., and Miyamoto, M. (2024). High-molecular-weight oligomer tau (HMW_oTau) species are dramatically increased in Braak-stage dependent manner in the frontal lobe of human brains, demonstrated by a novel oligomer Tau ELISA with a mouse monoclonal antibody (APNmAb005). *FASEB J.* 38, e70160. <https://doi.org/10.1096/fj.202401704R>.

128. Czernowicz, J., Chen, W., Wang, Q., Shen, C., Wager, C., Stone, I., Stebbins, C., Lamb, M., Setser, J., Cantone, G., et al. (2017). Pan-tau antibody BIIB076 exhibits promising safety and biomarker profile in cynomolgus monkey toxicity study. *Alzheimers Dement.* 13. <https://www.alzforum.org/papers/pan-tau-antibody-biib076-exhibits-promising-safety-and-biomarker-profile-cynomolgus-monkey>.

129. Grimm, H.P., Schumacher, V., Schäfer, M., Imhof-Jung, S., Freskgård, P.O., Brady, K., Hofmann, C., Rüger, P., Schlothauer, T., Göpfert, U., et al. (2023). Delivery of the Brainshuttle™ amyloid-beta antibody fusion trintinemab to non-human primate brain and projected efficacious dose regimens in humans. *MAbs* 15, 2261509. <https://doi.org/10.1080/19420862.2023.2261509>.

130. Benn, J., Cheng, S., Keeling, S., Smith, A.E., Vaysburd, M.J., Böken, D., Miller, L.V.C., Katsinelos, T., Franco, C., Dupré, E., et al. (2024). Aggregate-selective removal of pathological tau by clustering-activated degraders. *Science* 385, 1009–1016. <https://doi.org/10.1126/science.adp5186>.

131. Danis, C., Dupré, E., Bouillet, T., Denéchaud, M., Lefebvre, C., Nguyen, M., Mortelecque, J., Cantrelle, F.X., Rain, J.C., Hanouille, X., et al. (2025). Inhibition of tau neuronal internalization using anti-tau single domain antibodies. *Nat. Commun.* 16, 3162. <https://doi.org/10.1038/s41467-025-58383-4>.

132. Cummings, J.L., Teunissen, C.E., Fiske, B.K., Le Ber, I., Wildsmith, K.R., Schöll, M., Dunn, B., and Scheltens, P. (2025). Biomarker-guided decision making in clinical drug development for neurodegenerative disorders. *Nat. Rev. Drug Discov.* 24, 589–609. <https://doi.org/10.1038/s41573-025-01165-w>.

133. Robinson, J.L., Lee, E.B., Xie, S.X., Rennert, L., Suh, E., Bredenberg, C., Caswell, C., Van Deerlin, V.M., Yan, N., Yousef, A., et al. (2018). Neurodegenerative disease concomitant proteinopathies are prevalent, age-related and APOE4-associated. *Brain* 141, 2181–2193. <https://doi.org/10.1093/brain/awy146>.

134. Franzmeier, N., Roemer-Cassiano, S.N., Bernhardt, A.M., Dehsarvi, A., Dewenter, A., Steward, A., Biel, D., Frontzkowski, L., Zhu, Z., Gnörich, J., et al. (2025). Alpha synuclein co-pathology is associated with accelerated amyloid-driven tau accumulation in Alzheimer's disease. *Mol. Neurodegener.* 20, 31. <https://doi.org/10.1186/s13024-025-00822-3>.

135. Brettschneider, J., Del Tredici, K., Lee, V.M.Y., and Trojanowski, J.Q. (2015). Spreading of pathology in neurodegenerative diseases: a focus on human studies. *Nat. Rev. Neurosci.* 16, 109–120. <https://doi.org/10.1038/nrn3887>.

136. Stopchinski, B.E., and Diamond, M.I. (2017). The prion model for progression and diversity of neurodegenerative diseases. *Lancet Neurol.* 16, 323–332. [https://doi.org/10.1016/S1474-4422\(17\)30037-6](https://doi.org/10.1016/S1474-4422(17)30037-6).

137. Stancu, I.C., Cremers, N., Vanrusselt, H., Couturier, J., Vanoosthuyse, A., Kessels, S., Lodder, C., Brône, B., Huaux, F., Octave, J.N., et al. (2019). Aggregated Tau activates NLRP3-ASC inflammasome exacerbating exogenously seeded and non-exogenously seeded Tau pathology in vivo. *Acta Neuropathol.* 137, 599–617. <https://doi.org/10.1007/s00401-018-01957-y>.

138. Vasconcelos, B., Stancu, I.C., Buist, A., Bird, M., Wang, P., Vanoosthuyse, A., Van Kolen, K., Verheyen, A., Kienlen-Campard, P., Octave, J.N., et al. (2016). Heterotypic seeding of Tau fibrillization by pre-aggregated Abeta provides potent seeds for prion-like seeding and propagation of Tau-pathology in vivo. *Acta Neuropathol.* 131, 549–569. <https://doi.org/10.1007/s00401-015-1525-x>.

139. Wang, P., Joberty, G., Buist, A., Vanoosthuyse, A., Stancu, I.C., Vasconcelos, B., Pierrot, N., Faelth-Savitski, M., Kienlen-Campard, P., Octave, J.N., et al. (2017). Tau interactome mapping based identification of Otub1 as Tau deubiquitinase involved in accumulation of pathological Tau forms in vitro and in vivo. *Acta Neuropathol.* 133, 731–749. <https://doi.org/10.1007/s00401-016-1663-9>.

140. Clavaguera, F., Akatsu, H., Fraser, G., Crowther, R.A., Frank, S., Hench, J., Probst, A., Winkler, D.T., Reichwald, J., Staufenbiel, M., et al. (2013). Brain homogenates from human tauopathies induce tau inclusions in mouse brain. *Proc. Natl. Acad. Sci. USA.* 110, 9535–9540. <https://doi.org/10.1073/pnas.1301175110>.

141. Vogel, J.W., Iturria-Medina, Y., Strandberg, O.T., Smith, R., Levitis, E., Evans, A.C., and Hansson, O.; Alzheimer's Disease Neuroimaging Initiative; Swedish BioFinder Study (2020). Spread of pathological tau proteins through communicating neurons in human Alzheimer's disease. *Nat. Commun.* 11, 2612. <https://doi.org/10.1038/s41467-020-15701-2>.

142. Holmes, B.B., Furman, J.L., Mahan, T.E., Yamasaki, T.R., Mirbaha, H., Eades, W.C., Belaygorod, L., Cairns, N.J., Holtzman, D.M., and Diamond, M.I. (2014). Proteopathic tau seeding predicts tauopathy in vivo. *Proc. Natl. Acad. Sci. USA.* 111, E4376–E4385. <https://doi.org/10.1073/pnas.1411649111>.

143. Kaufman, S.K., Del Tredici, K., Thomas, T.L., Braak, H., and Diamond, M.I. (2018). Tau seeding activity begins in the transentorhinal/entorhinal regions and anticipates phospho-tau pathology in Alzheimer's disease and PART. *Acta Neuropathol.* 136, 57–67. <https://doi.org/10.1007/s00401-018-1855-6>.

144. Gibbons, G.S., Lee, V.M.Y., and Trojanowski, J.Q. (2019). Mechanisms of Cell-to-Cell Transmission of Pathological Tau: A Review. *JAMA Neurol.* 76, 101–108. <https://doi.org/10.1001/jamaneurol.2018.2505>.

145. Brunello, C.A., Merezhko, M., Uronen, R.L., and Huttunen, H.J. (2020). Mechanisms of secretion and spreading of pathological tau protein. *Cell. Mol. Life Sci.* 77, 1721–1744. <https://doi.org/10.1007/s00018-019-03349-1>.

146. Fowler, S.L., Behr, T.S., Turkes, E., O'Brien, D.P., Cauhy, P.M., Rawlinson, I., Edmonds, M., Foiani, M.S., Schaler, A., Crowley, G., et al. (2025). Tau filaments are tethered within brain extracellular vesicles in Alzheimer's disease. *Nat. Neurosci.* 28, 40–48. <https://doi.org/10.1038/s41593-024-01801-5>.

147. Rauch, J.N., Luna, G., Guzman, E., Audouard, M., Challis, C., Sibih, Y.E., Leshuk, C., Hernandez, I., Wegmann, S., Hyman, B.T., et al. (2020). LRP1 is a master regulator of tau uptake and spread. *Nature* 580, 381–385. <https://doi.org/10.1038/s41586-020-2156-5>.

148. Holmes, B.B., DeVos, S.L., Kfoury, N., Li, M., Jacks, R., Yanamandra, K., Ouidja, M.O., Brodsky, F.M., Marasa, J., Bagchi, D.P., et al. (2013). Heparan sulfate proteoglycans mediate internalization and propagation of specific proteopathic seeds. *Proc. Natl. Acad. Sci. USA.* **110**, E3138–E3147. <https://doi.org/10.1073/pnas.1301440110>.

149. Gallardo, G., Wong, C.H., Ricardez, S.M., Mann, C.N., Lin, K.H., Leyns, C.E.G., Jiang, H., and Holtzman, D.M. (2019). Targeting tauopathy with engineered tau-degrading intrabodies. *Mol. Neurodegener.* **14**, 38. <https://doi.org/10.1186/s13024-019-0340-6>.

150. Wongsodirdjo, P., Caruso, A.C., Yong, A.K., Lester, M.A., Vella, L.J., Hung, Y.H., and Nisbet, R.M. (2024). Messenger RNA-encoded antibody approach for targeting extracellular and intracellular tau. *Brain Commun.* **6**, fcae100. <https://doi.org/10.1093/braincomms/fcae100>.

151. Lövestam, S., Li, D., Wagstaff, J.L., Kotecha, A., Kimanis, D., McLaughlin, S.H., Murzin, A.G., Freund, S.M.V., Goedert, M., and Scheres, S.H.W. (2024). Disease-specific tau filaments assemble via polymorphic intermediates. *Nature* **625**, 119–125. <https://doi.org/10.1038/s41586-023-06788-w>.

152. Heneka, M.T., van der Flier, W.M., Jessen, F., Hoozemans, J., Thal, D.R., Boche, D., Brosseron, F., Teunissen, C., Zetterberg, H., Jacobs, A.H., et al. (2025). Neuroinflammation in Alzheimer disease. *Nat. Rev. Immunol.* **25**, 321–352. <https://doi.org/10.1038/s41577-024-01104-7>.

153. Schlepckow, K., Morenas-Rodríguez, E., Hong, S., and Haass, C. (2023). Stimulation of TREM2 with agonistic antibodies—an emerging therapeutic option for Alzheimer's disease. *Lancet Neurol.* **22**, 1048–1060. [https://doi.org/10.1016/S1474-4422\(23\)00247-8](https://doi.org/10.1016/S1474-4422(23)00247-8).

154. Serrano-Pozo, A., Das, S., and Hyman, B.T. (2021). APOE and Alzheimer's disease: advances in genetics, pathophysiology, and therapeutic approaches. *Lancet Neurol.* **20**, 68–80. [https://doi.org/10.1016/S1474-4422\(20\)30412-9](https://doi.org/10.1016/S1474-4422(20)30412-9).

155. Shi, Y., and Holtzman, D.M. (2018). Interplay between innate immunity and Alzheimer disease: APOE and TREM2 in the spotlight. *Nat. Rev. Immunol.* **18**, 759–772. <https://doi.org/10.1038/s41577-018-0051-1>.

156. Martens, Y.A., Zhao, N., Liu, C.C., Kanekiyo, T., Yang, A.J., Goate, A.M., Holtzman, D.M., and Bu, G. (2022). ApoE Cascade Hypothesis in the pathogenesis of Alzheimer's disease and related dementias. *Neuron* **110**, 1304–1317. <https://doi.org/10.1016/j.neuron.2022.03.004>.

157. Princen, K., Van Dooren, T., van Gorsel, M., Louros, N., Yang, X., Dumbacher, M., Bastiaens, I., Coupet, K., Dupont, S., Cuveliers, E., et al. (2024). Pharmacological modulation of septins restores calcium homeostasis and is neuroprotective in models of Alzheimer's disease. *Science* **384**, eadd6260. <https://doi.org/10.1126/science.add6260>.