

Functional Characterization of Cultured Keratinocytes after Acute Cutaneous Burn Injury

Gerd G. Gauglitz^{1*}, Siegfried Zedler², Felix v. Spiegel³, Jasmin Fuhr³, Guido Henkel v. Donnersmarck³, Eugen Faist²

1 Department of Dermatology and Allergy, Ludwig-Maximilian University, Munich, Germany, **2** Department of Surgery, Ludwig-Maximilian University, Munich, Germany, **3** Burn Unit, Department of Plastic and Reconstructive Surgery, Community Hospital Bogenhausen, Munich, Germany

Abstract

Background: In addition to forming the epithelial barrier against the outside environment keratinocytes are immunologically active cells. In the treatment of severely burned skin, cryoconserved keratinocyte allografts gain in importance. It has been proposed that these allografts accelerate wound healing also due to the expression of a favourable - keratinocyte-derived - cytokine and growth factor milieu.

Methods: In this study the morphology and cytokine expression profile of keratinocytes from skin after acute burn injury was compared to non-burned skin. Skin samples were obtained from patients after severe burn injury and healthy controls. Cells were cultured and secretion of selected inflammatory mediators was quantified using Bioplex Immunoassays. Immunohistochemistry was performed to analyse further functional and morphologic parameters.

Results: Histology revealed increased terminal differentiation of keratinocytes (CK10, CK11) in allografts from non-burned skin compared to a higher portion of proliferative cells (CK5, vimentin) in acute burn injury. Increased levels of IL-1 α , IL-2, IL-4, IL-10, IFN- γ and TNF α could be detected in culture media of burn injury skin cultures. Both culture groups contained large amounts of IL-1RA. IL-6 and GM-CSF were increased during the first 15 days of culture of burned skin compared to control skin. Levels of VEGF, FGF-basic, TGF- β und G-CSF were high in both but not significantly different. Cryoconservation led to a diminished mediator synthesis except for higher levels of intracellular IL-1 α and IL-1 β .

Conclusion: Skin allografts from non-burned skin show a different secretion pattern of keratinocyte-derived cytokines and inflammatory mediators compared to keratinocytes after burn injury. As these secreted molecules exert auto- and paracrine effects and subsequently contribute to healing and barrier restoration after acute burn injury therapies affecting this specific cytokine/growth factor micromilieu could be beneficial in burned patients.

Citation: Gauglitz GG, Zedler S, v. Spiegel F, Fuhr J, v. Donnersmarck GH, et al. (2012) Functional Characterization of Cultured Keratinocytes after Acute Cutaneous Burn Injury. PLoS ONE 7(2): e29942. doi:10.1371/journal.pone.0029942

Editor: Jürgen Schaubert, Ludwig-Maximilian-University, Germany

Received: December 4, 2011; **Accepted:** December 8, 2011; **Published:** February 16, 2012

Copyright: © 2012 Gauglitz et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The authors have no support or funding to report.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: gerd.gauglitz@med.uni-muenchen.de

Introduction

Loss of the integrity of large portions of the skin as a result of burn injury may lead to major disability or even death. As the skin forms an active barrier protecting our organism from the outside environment rapid restoration of the epidermal barrier is of vital relevance after acute burn injury. Thus, appropriate wound care is mandatory and various treatment modalities have been utilized to improve and accelerate wound healing. In the past decades, the increasing knowledge of the molecular and cellular mechanisms underlying wound repair and regeneration has led to extensive usage of growth factors in wound care [1–8]. Growth factors and cytokines play major roles in the well-orchestrated integration of the complex biological and molecular events underlying cutaneous wound healing, including cell migration and proliferation, extracellular matrix deposition, angiogenesis and tissue remodeling [9]. However, the clinical effects of the topical application of single growth factors to accelerate wound healing have been discouraging: On the one hand due to the complexity of the wound healing

cascade and on the other due physical inhibition and biological degradation of topically applied factors [10]. The development of gene transfer technology promised to overcome the limitations associated with the (topical) application of recombinant proteins by delivering the respective growth factor directly to the wound bed [11]. Also, stem cells - due to their pluripotency and their growth potential - make them a potentially useful vehicle for gene delivery to injury site [10]. Still, the use of stem cell technology is far from a therapeutic application to date.

The epidermal skin barrier is formed by keratinocytes which secrete a multitude of biological active molecules contributing transiently to inflammatory responses and wound healing. In particular, keratinocytes have been shown to control the behaviour of fibroblasts during wound healing through the secretion, activation or inhibition of cytokines and growth factors such as TGF- β [12]. In the treatment of severely burned patients autologous keratinocyte-sheets but also (cryoconserved) allografts are used. It is supposed that in comparison to keratinocytes sheets these allografts accelerate wound healing possibly due to the

expression of favourable cytokines and growth factors. However, the mechanisms of burn wound healing after allograft transplantation are not well characterized.

Materials and Methods

Patients

Seventeen severely burned patients, admitted to the Department of Burns, Community Hospital Bogenhausen, Munich, Germany, between 2002 and 2005, were enrolled in this prospective study. Inclusion criteria included: Admission within 24 hours post burn injury, burns covering more than 25% of the total body surface area (TBSA), <70 years of age, written consent to the experimental protocol. The study design was approved by the local ethics committee at the Ludwig-Maximilian University, Munich, Germany. 17 randomly selected female patients who underwent breast reduction surgery served as controls. On admission, the extent and degree of burn was assessed and recorded on a standard Lund and Browder chart. Information recorded at the time of admission included burn related (date and mechanism) as well as demographic data (age and gender). All patients were treated in our burn intensive care unit according to standardized protocols. Sepsis was defined as a positive blood culture or pathologic tissue culture identifying the pathogen during hospitalization or at autopsy, in combination with at least 3 of the following: leucocytosis or leucopenia ($>12,000$ or $<4,000$), hyperthermia or hypothermia (>38.5 or $<36.5^{\circ}\text{C}$), tachycardia (>150 BPM in children), refractory hypotension (systolic BP <90 mmHg), thrombocytopenia (platelets $<50,000/\text{mm}^3$), hyperglycemia (serum glucose >240 mg/dl), and enteral feeding intolerance (residuals >200 cc/hr or diarrhea >1 L/day).

Isolation and culture of human keratinocytes

Tissue specimens were obtained during surgical debridement close (approximately 5 cm) to full-thickness burn wounds. Tissue from randomly selected female patients who underwent breast reduction surgery served as a control. Epidermal keratinocytes were isolated starting immediately after tissue harvest following the method of Rheinwald and Green [13]. In brief, harvested tissue was trypsinized (Trypsin 0,05%/EDT 0,02%, Gibco BRL, Invitrogen GmbH, Karlsruhe, Germany) for 12 to 18 hours and dermis was mechanically removed. Obtained cells were resuspended in Dulbecco's modified Eagle's medium (DMEM)/HAMS-F-12 media (Biochrom AG Seromed, Berlin, Germany), centrifuged at 200 g, resuspended in 20 ml of DMEM/HAMS-F-12 media supplemented with L-Glutamine, NaHCO_3 , EGF, FCS, gentamycin, insulin and hydrocortisone and seeded in cell culture flasks at 2×10^6 each flask. Culture medium was collected and frozen in liquid nitrogen for further analysis after 72 hour incubation periods prior to day 3, 10 and 15 (Figure 1). At day 15 confluent cells were washed with dispase solution (2,5 mg/ml DMEM, Roche Diagnostics GmbH, Mannheim, Germany) for 20 minutes at 37 C and 5% CO_2 . A piece of Grassolind (Paul Hartmann AG, Heidenheim, Germany) measuring 75 cm^2 was applied to the cell layer and attached by hemoclips. 24 hours later the culture media was collected (pre cryo) before cell sheets were frozen in MEM Earle supplemented with 20% FCS and 10% DMSO at -80°C . Later on sheets were thawed and re-cultured in DMEM. 24 hours thereafter culture media was collected and frozen for further processing (post cryo, Figure 1).

Cellular morphology and immunohistochemistry

For routine light microscopy, cytopspins from cultured keratinocytes from normal and burned skin were fixed in 10% formalin

and stained with hematoxylin and eosin (at day 10). Immunohistochemistry was used to detect cytokeratin, vimentin and other proteins in order to allow comparison of cellular composition in allografts prepared from discarded tissue following breast reduction versus autologous cultured keratinocytes. For this, cytopspins were incubated with the respective primary monoclonal antibodies (listed in Table 1). After frequent washings, a biotinylated rabbit anti-rat secondary antibody or a rabbit anti-mouse secondary antibody was applied respectively. After rinsing, enzyme conjugate was applied for 10 min. Final incubation included Substrate-Chromogen Mixture (3-amino-9-ethylcarbazole, AEC) and positive staining revealed a red precipitate.

Cytokine and growth factor measurements

The Bio-Plex Human Cytokine 17-Plex panel was used with the Bio-Plex Suspension Array System (Bio-Rad, Hercules, CA) to profile expression of interleukin (IL)-1 α , IL-1 receptor antagonist (IL-1Ra), IL-1 β , IL-2, IL-4, IL-6, IL-10, interferon-gamma (IFN- γ), tumor necrosis factor alpha (TNF- α), Vascular endothelial growth factor (VEGF), fibroblast growth factor basic (FGF-basic) granulocyte colony-stimulating factor (G-CSF) and granulocyte-macrophage colony-stimulating factor (GM-CSF). The assay was performed according to the manufacturer's instructions. Briefly, cell culture medium was centrifuged at 4500 rpm for 3 minutes at 4°C . Samples were then incubated with microbeads labeled with specific antibodies to one of the aforementioned cytokines for 30 minutes. Following a washing step, the beads were incubated with the detection antibody cocktail, each antibody specific to a single cytokine. After another wash step, beads were incubated with streptavidin-phycoerythrin for 10 minutes, again washed and the concentration of each cytokine was determined using the array reader. Transforming growth factor beta-1 (TGF- β 1) was determined by double-sandwich, enzyme-linked, immunosorbent assays (Biosource Europe SA, B-1400 Nivelles, Belgium) according to the protocol of the manufacturer in a microplate reader (Vmax, Molecular Devices Corporation, Sunnyvale, CA, USA) at 450 nm.

Statistical analysis

Paired and unpaired Student's t-test and Mann-Whitney tests were used where appropriate. Data are expressed as means \pm SD or SEM as indicated. Differences were considered significant at a p value of <0.05 .

Results

Patient characteristics

Characteristics of burn patients and controls at the time of hospitalization are listed in Table 1. Patients were on average 41 years old and suffered from severe burn injury involving 42% of TBSA and third-degree burn involving 28% of TBSA. There were more male patients than female patients in the experimental group. During acute hospitalization, 24% of the patients suffered from inhalation injury, sepsis occurred in 29% and multi-organ failure (MOV) in 18% of patients. 24% of our patients subsequently died from their injuries (Table 2). 17 randomly selected female patients aged 20 to 50 years (mean 37 ± 7 years) who underwent breast reduction surgery served as controls.

H&E stainings and immunohistochemistry

H&E stainings revealed a significantly increased proportion of terminally differentiated cells (white arrows) in allogenic cultured epithelial grafts from control tissue (Fig. 2A) in comparison to cells from autologous cultured epithelial grafts from burn patients (Fig. 2B). In stainings with the monoclonal antibody KL-1 (which

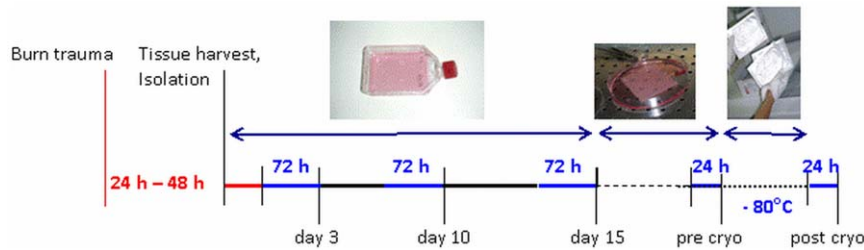


Figure 1. Time line and study design.
doi:10.1371/journal.pone.0029942.g001

marks the cytokeratins 2, 6, 8, 10, 11, 18 and 19 and especially K 6, 10 and 11 are expressed by terminally differentiated keratinocytes) significantly more terminal differentiated keratinocytes in allogenic graft (Fig. 2C) were detected compared with autologous cultured epithelial grafts (Fig. 2D). Vimentin is expressed by mesenchymal cells (e.g. fibroblasts) but also by basaloid keratinocytes. More vimentin-positive cells were present in autologous cultured epithelial grafts (Fig. 2F) when compared to allogenic sheets (Fig. 2E). No CD-209, 1F11 or T-200 (CD45) or MOPC-21 expressing cells could be detected in either group indicating the absence of dermal dendritic cells, macrophages, monocytes, endothelial cells and leukocytes (data not shown).

Cytokine expression profiles

No significant differences between groups could be measured in culture supernatants for IL-1 α within the first 15 days of culture (Fig. 3A). During cell culture, levels of IL-1 β were barely detectable at all time points measured (Fig. 3B). In contrast, IL-1RA was detected at high concentrations in culture media of both patients and controls peaking at day 15 (16690,69 \pm 2258,85 pg/ml vs. 12807,68 \pm 3016,70, respectively) without revealing any significant difference between patients and controls (Fig. 3C). Analyses of keratinocyte culture media revealed significantly elevated levels of IL-2, -4, -6, -10, IFN- γ , TNF- α and GM-CSF (Fig. 3D–I, N) at most of the first three time points studied in the patient group compared to control patients. However, expression of most of the respective cytokines was generally very low. Expression levels of these cytokines did not significantly differ when comparing non-surviving patients with survivors of the burn trauma (data not shown).

IL-6 and GM-CSF were significantly increased during the first 15 days of culture after acute burn injury compared to control skin (Fig. 3F, N) as a result of significant higher expression in non-survivors, Fig. 3O, P). Concentrations of IL-6 increased early after burn injury peaking at 10 days post injury (2874,04 \pm 1352,89 pg/

ml) while culture media analysis of the control group demonstrated basic levels during the 15 day culture period (with 69,35 \pm 21,82 pg/ml; 60,37 \pm 28,19 pg/ml, and 98,28 \pm 35,38 pg/ml, respectively). Noteworthy, when differentiating between survivors and non-survivors, keratinocytes of patients that did not survive the burn trauma revealed significantly increased secretion of IL-6 at the first three time points measured when compared to the survivor group. The expression profile of GM-CSF revealed a similar pattern however, differences between groups were, although statistically significant, not as pronounced as with IL-6.

Levels of VEGF, TGF- β , FGF-basic und G-CSF could be detected at relatively high concentrations, however only moderate differences could be detected comparing the two collectives (Fig. 3J–M). Interestingly, at 3 and 10 days post burn, keratinocytes isolated close to a burn wound did secrete significantly elevated levels of TGF- β 1 compared to normal controls (e.g. 526,43 \pm 61,16 pg/ml and 362,59 \pm 38,50 pg/ml at day 10, respectively). No differences could be detected comparing the secretion profile of keratinocytes from patients that did not survive the thermal trauma when compared to surviving patients.

When grafts of keratinocytes were prepared after 15 days and then frozen a diminished mediator synthesis from grafts from burn patients and controls after a freeze/thaw cycle was measured. In contrast, significant higher levels of IL-1 α , IL-1 β , IL-1RA were observed in cultured keratinocytes from burn injury patients after a freeze/thaw cycle and further culture for 24 hours as compared to control keratinocytes (Fig. 3A–C).

Discussion

Tissue-engineered skin substitutes constitute a promising alternative for the permanent closure of burn wounds and in the meantime protocols have been established for the culture of large numbers of human epidermal keratinocytes [3,14]. In particular,

Table 1. Antibodies used for immunohistochemical analyses.

Antigen	Anti-	Isotype	Product #	Company	conc.
Control	MOPC21	IgG1	M9269	Sigma	1 mg/ml
Control	UPC-10	IgG2a	M9144	Sigma	1 mg/ml
Cytokeratin 2,6,8,10,11,18,19	KI-1	IgG1	1918	Immunotech, Marseille	126,5 ng/ml
Vimentin	Vim 3B4	IgG2a	M7020	Sigma	60 μ g/ml
CD 45	T 200	IgG1	M0701	Sigma	0,45 mg/ml
PECAM-1/CD 31	1F11	IgG1	2052 Klon 1F11	Immunotech, Marseille	200 ng/ml
DC-Sign/ CD 209	DCN46	IgG2b	551249	BD Biosciences	31,25 μ g/ml

doi:10.1371/journal.pone.0029942.t001

Table 2. Patient characteristics (n = 17).

Patient characteristics	Mean \pm SEM
Age (years)	41 \pm 4
Gender (F/M)	3/14
TBSA (%)	42 \pm 2
3 rd degree (%)	28 \pm 4
MOV (%)	18
Inhalation injury (%)	24
Sepsis (%)	29
Non-survivors (%)	24

Randomly selected female patients who underwent breast reduction surgery served as a control population (n = 17; Age 37 \pm 7 years). doi:10.1371/journal.pone.0029942.t002

autologous epithelial cells grown from a single full-thickness skin biopsy have been available for nearly three decades [15]. These keratinocytes were considered solely as reconstituting elements of the skin barrier. Meanwhile it is well known that these cells secrete a multitude of biological relevant molecules which contribute to inflammatory and immunological responses and wound healing [12]. Particularly, release of IL-1 from various cells at the wound site seems to represent the initial trigger for the inflammatory reaction and serves as an autocrine, activating signal to fibroblasts and endothelial cells [16,17].

The characterization of the secreted factors from keratinocytes could improve our understanding of their role in wound healing and – in the case of skin graft transplantation for burn injuries – improve treatments. When comparing the cellular morphology of keratinocytes in allografts made of breast reductions with autologous keratinocytes from burns removed close to the wound we observed differences in proliferative activity and differentiation as indicated by increased expression of vimentin in autologous cultured epithelial grafts and elevated KL-1 expression by keratinocytes isolated from breast reduction tissue.

Comparing the *in vitro* cytokine secretion profile of keratinocytes obtained from an area close to a full-thickness burn wound after longterm culture with normal keratinocytes significantly elevated levels of IL-2, -4, -6, -10, IFN- γ , TNF- α , TGF- β 1 and GM-CSF in culture media of keratinocytes from burn patients were observed. In contrast, only minimal concentrations of IL-1 β and no significant differences in IL-1 α expression could be detected. While keratinocytes express the inactive pro-IL-1 β , its active form is secreted [18]. Still, IL-1 β was barely detectable in our experiments. IL-1 α on the other hand, is barely detectable under physiologic conditions, but may be secreted by keratinocytes after tissue damage [18]. In our analyses IL-1 α was detected in cultures from burn patients and controls, however no significant difference was observed after 15 days of culture.

IL-6, TNF- α , and TGF- β 1 are further mediators known to exert pro-inflammatory (IL-6, TNF- α [19,20]) and keratinocyte and fibroblast proliferating (IL-6 [21], TNF- α [22,23], and TGF- β [24]) properties and may thus significantly contribute to the physiologic wound healing process. Keratinocytes in healthy skin do rarely express IL-6 [25]. However, cutaneous injury or physiological processes such as UVB irradiation induce increased IL-6 expression in different cell types of the dermis and in keratinocytes [26]. The significantly enhanced *in vitro* release of the potent mitogen and pro-inflammatory cytokine IL-6 from keratinocytes after severe burns may thus be important in the

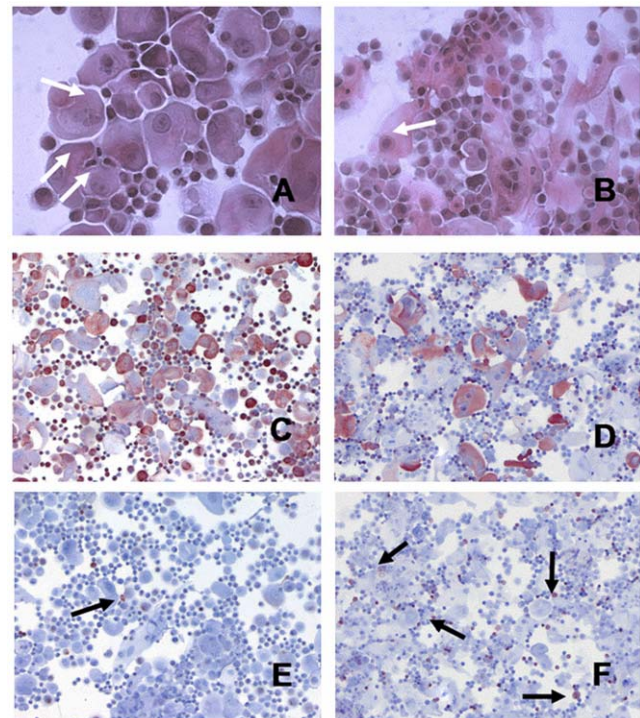


Figure 2. Morphological characterization of keratinocytes from acute burn injury. H&E stainings revealed a significantly increased proportion of terminally differentiated cells (white arrows) in allogenic cultured epithelial grafts (Fig. 2A) from breast reduction material in comparison to cells from autologous cultured epithelial grafts from burn patients (Fig. 2B). In stainings with the monoclonal antibody KL-1 significantly more terminal differentiated keratinocytes in allogenic grafts (Fig. 2C) were detected compared with autologous cultured epithelial grafts (Fig. 2D). More vimentin-positive cells are present in autologous cultured epithelial grafts (Fig. 2F) when compared to allogenic sheets (black arrows, Fig. 2E). doi:10.1371/journal.pone.0029942.g002

inflammatory event and the increased activity of the thermally affected epithelial cells with auto- and paracrine effects.

In addition, genes and proteins of IL-6 and GM-CSF show a high sequence homology and their receptor binding and signal transduction structures are uniform. This suggests common signal transduction and activation pathways [26] and may explain their comparable expression profiles in our study. Indeed, increased GM-CSF expression in human and murine skin with damaged barrier function has been demonstrated in various studies [27–29].

TNF- α is predominantly expressed in keratinocytes and has a profound autocrine effect on keratinocytes in wound healing. Expression of this growth factor is upregulated in keratinocytes after skin injury [30]. Surprisingly, wound healing experiments in TNF- α -deficient mice showed no abnormal wound healing phenotype [31,32]. TNF- α acts as strong mitogen for fibroblasts [33]. However, the lack of an obvious mesenchymal phenotype in TNF- α -deficient mice suggests that this growth factor is dispensable for fibroblast proliferation and that its loss can be compensated by other EGFR ligands.

TGF- β plays an important role in wound healing as demonstrated in various *in vitro* and *in vivo* studies. The TGF- β family consists of at least five highly conserved polypeptides, with TGF- β 1, -2 and -3 being the principal mammalian forms. TGF- β 1 and -2 are one of the most important stimulators of collagen and proteoglycan synthesis, and affect the ECM not only by stimulating collagen synthesis but also by preventing its breakdown

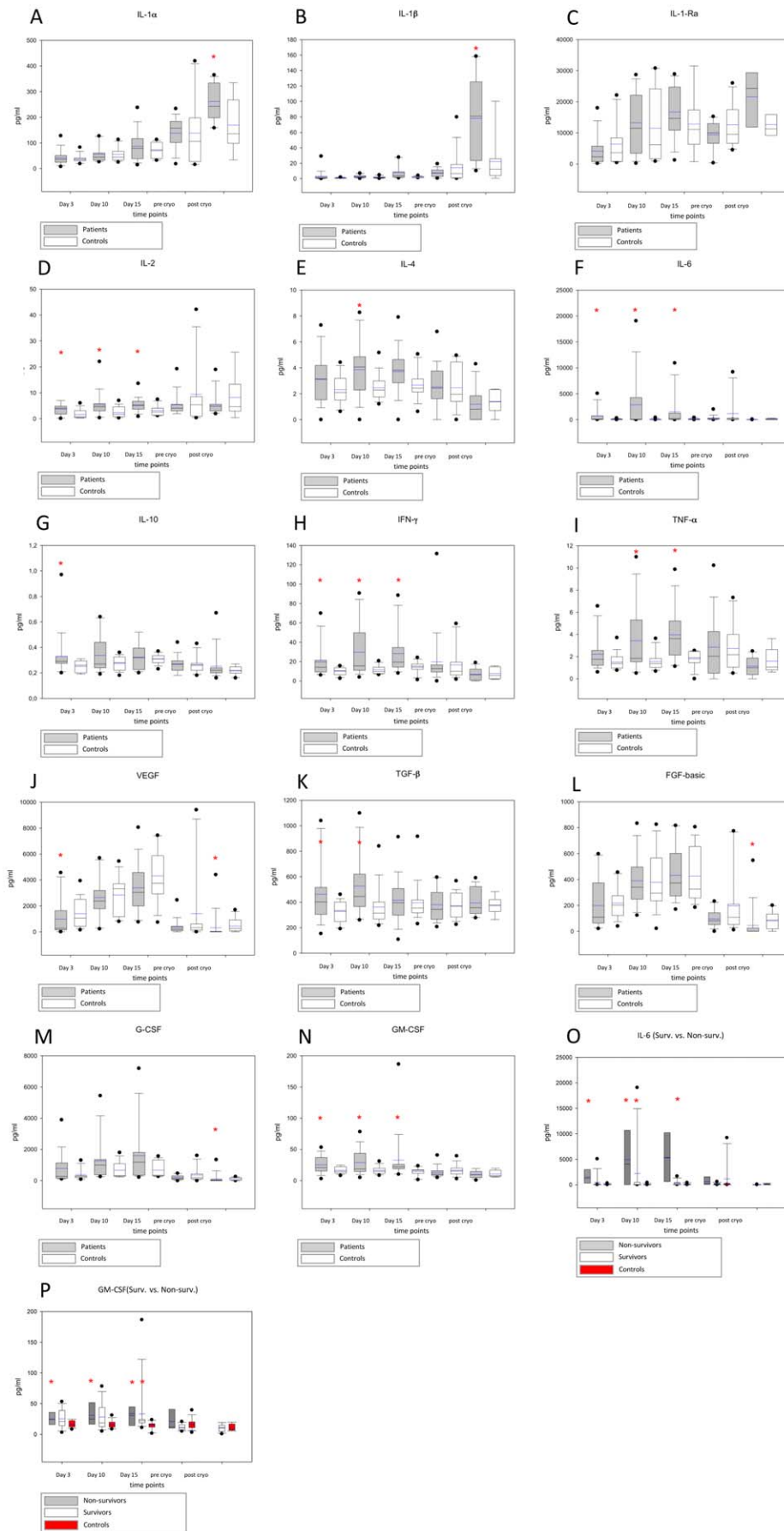


Figure 3. Cytokine profiles in keratinocytes cultures from burn injuries. Fourteen cytokines secreted by keratinocytes and measured in this study were significantly altered in keratinocytes cultures from sites of burn injury. Particularly levels of IL-2, -4, IL-6, -10, IFN- γ , TGF- β 1 and TNF α were significantly altered in response to burn injury as compared to controls. Histograms depict culture media concentrations of the respective cytokine at the time of analysis. Bars represent means; error bars correspond to SEM. Asterisks denote statistical difference between cultures from burned skin vs. control skin. (* $p < 0.05$).
doi:10.1371/journal.pone.0029942.g003

[34,35]. In contrast, TGF- β 3, which is predominantly induced in the later stages of wound healing, has been found to reduce connective tissue deposition [36]. Hence, overexpression of TGF- β 1 and β 2 has been linked to excessive scar formation [37,38], while increased levels of TGF- β 3 appear to prevent excessive scar formation. Specifically, beyond one-week, differential expression of TGF- β isoforms, receptors and activity modulators, rather than the mere presence or absence of TGF- β , may have a major role in the development of keloids and hypertrophic scarring [39]. Although, within this study only TGF- β 1 was quantified, its significantly increased secretion by keratinocytes obtained from burn wounds may indeed stimulate underlying fibroblasts to produce elevated levels of ECM and by this significantly contributing to hypertrophic scar formation, commonly observed in this patient population.

Noteworthy, VEGF, FGF-basic, and G-CSF were produced in relatively high quantities with no significant differences between groups. VEGF represents one of the most important pro-angiogenic factors. It accelerates wound healing by increasing neovascularization during the early period of wound healing [10,40]. FGF-basic is produced by keratinocytes and has chemotactic effects on fibroblasts, smooth muscle cells, endothelial cells and keratinocytes [41]. Like VEGF it has pro-angiogenic properties and is essential for wound healing [42]. Clinically, local application of FGF-basic led to improved healing of diabetic ulcers [43].

However, the application of cultured keratinocytes no matter if autologous or allogenic requires the presence of a dermal component since keratinocytes do not stimulate granulation tissue formation or dermal remodelling in full-thickness wounds [44,45]. Therefore, autologous or allogenic keratinocyte sheets are less suited for application onto inert, poorly granulating wounds (e.g.,

ulcers). They are more suited for the application onto well-granulating wounds in which the dermal component is already present. E.g. cultured keratinocyte sheets are applied onto large burns in a two-step procedure in which a dermal compartment (dead donor dermis) is initially applied and allowed to stabilize (develop capillaries) before applying the cultured keratinocytes [46]. Based on the delay between taking a biopsy specimen from the patient and the keratinocyte autograft becoming available, sheets of allogenic origin from unrelated donors have been used as keratinocyte allografts. Their proposed mechanisms of actions of on wound healing may indeed be mostly exerted via the secretion of the respective growth factors. However, in our study, freezing led to a diminished mediator synthesis, except for significant higher levels of IL-1 α and IL-1 β , probably as a result of physical cell destruction after thawing. It thus remains questionable if cytokines and growth factors of interest survive deep-freezing.

In conclusion, epidermal keratinocytes from burn patients show an increased proliferative activity with augmented mediator release compared to keratinocytes isolated from control donors. In the future, the substitution of cellular growth-enhancing cytokines and mediators (and maybe additional cell types besides keratinocytes) in the bioengineered skin substitutes may prove promising to generate skin grafts with improved wound-healing function after burn injury.

Author Contributions

Conceived and designed the experiments: GG SZ FS JF GHD EF. Performed the experiments: GG SZ FS JF GHD EF. Analyzed the data: GG SZ FS JF GHD EF. Contributed reagents/materials/analysis tools: GG SZ FS JF GHD EF. Wrote the paper: GG.

References

- Blome-Eberwein S, Jester A, Kuentscher M, Raff T, Germann G, et al. (2002) Clinical practice of glycerol preserved allograft skin coverage. *Burns* 28 Suppl 1: S10–12.
- Garfein ES, Orgill DP, Pribaz JJ (2003) Clinical applications of tissue engineered constructs. *Clin Plast Surg* 30: 485–498.
- Munster AM (1996) Cultured skin for massive burns. A prospective, controlled trial. *Ann Surg* 224: 372–375; discussion 375–377.
- Barret JP, Wolf SE, Desai MH, Herndon DN (2000) Cost-efficacy of cultured epidermal autografts in massive pediatric burns. *Ann Surg* 231: 869–876.
- Bannasch H, Fohn M, Unterberg T, Bach AD, Weyand B, et al. (2003) Skin tissue engineering. *Clin Plast Surg* 30: 573–579.
- Pellegrini G, Ranno R, Stracuzzi G, Bondanza S, Guerra L, et al. (1999) The control of epidermal stem cells (holoclones) in the treatment of massive full-thickness burns with autologous keratinocytes cultured on fibrin. *Transplantation* 68: 868–879.
- Hohlfeld J, de Buys Roessingh A, Hirt-Burri N, Chaubert P, Gerber S, et al. (2005) Tissue engineered fetal skin constructs for paediatric burns. *Lancet* 366: 840–842.
- Muller M, Gahankari D, Herndon DN (2007) Operative Wound Management. In: Herndon DN, ed. *Total Burn Care*. 3 ed. New York City: Saunders. pp 177–195.
- Gauglitz GG, Korting HC, Pavicic T, Ruzicka T, Jeschke MG (2011) Hypertrophic scarring and keloids: pathomechanisms and current and emerging treatment strategies. *Mol Med* 17: 113–125.
- Gauglitz GG, Jeschke MG (2011) Combined gene and stem cell therapy for cutaneous wound healing. *Mol Pharm* 8: 1471–1479.
- Branski LK, Gauglitz GG, Herndon DN, Jeschke MG (2009) A review of gene and stem cell therapy in cutaneous wound healing. *Burns* 35: 171–180.
- Armour A, Scott PG, Tredget EE (2007) Cellular and molecular pathology of HTS: basis for treatment. *Wound Repair Regen* 15 Suppl 1: S6–17.
- Rheinwald JG, Green H (1975) Serial cultivation of strains of human epidermal keratinocytes: the formation of keratinizing colonies from single cells. *Cell* 6: 331–343.
- Boyce ST (1996) Cultured skin substitutes: a review. *Tissue Eng* 2: 255–266.
- (1981) Grafting of burns with cultured epithelium prepared from autologous epidermal cells. *Lancet* 1: 75–78.
- Niessen FB, Schalkwijk J, Vos H, Timens W (2004) Hypertrophic scar formation is associated with an increased number of epidermal Langerhans cells. *J Pathol* 202: 121–129.
- Andriessen MP, Niessen FB, Van de Kerkhof PC, Schalkwijk J (1998) Hypertrophic scarring is associated with epidermal abnormalities: an immunohistochemical study. *J Pathol* 186: 192–200.
- Nashan D, Luger TA (1999) [Interleukin 1. 1: Basic principles and pathophysiology]. *Hautarzt* 50: 680–688.
- Spiekstra SW, Toebak MJ, Sampat-Sardjoeapersad S, van Beek PJ, Boersma DM, et al. (2005) Induction of cytokine (interleukin-1 α and tumor necrosis factor- α) and chemokine (CCL20, CCL27, and CXCL8) alarm signals after allergen and irritant exposure. *Exp Dermatol* 14: 109–116.
- Nakae S, Komiyama Y, Narumi S, Sudo K, Horai R, et al. (2003) IL-1-induced tumor necrosis factor- α elicits inflammatory cell infiltration in the skin by inducing IFN- γ -inducible protein 10 in the elicitation phase of the contact hypersensitivity response. *Int Immunol* 15: 251–260.
- Zhang JZ, Maruyama K, Iwatsuki K, Ono I, Kaneko F (1994) Effects of prostaglandin E1 on human keratinocytes and dermal fibroblasts: a possible mechanism for the healing of skin ulcers. *Exp Dermatol* 3: 164–170.
- Ono I, Gunji H, Suda K, Iwatsuki K, Kaneko F (1994) Evaluation of cytokines in donor site wound fluids. *Scand J Plast Reconstr Surg Hand Surg* 28: 269–273.
- Schultz GS, White M, Mitchell R, Brown G, Lynch J, et al. (1987) Epithelial wound healing enhanced by transforming growth factor- α and vaccinia growth factor. *Science* 235: 350–352.

24. Pierce GF, Mustoe TA, Lingelbach J, Masakowski VR, Griffin GL, et al. (1989) Platelet-derived growth factor and transforming growth factor-beta enhance tissue repair activities by unique mechanisms. *J Cell Biol* 109: 429–440.
25. Ohta Y, Nishiyama S, Nishioka K (1994) In situ expression of interleukin-6 in psoriatic epidermis during treatment. *J Dermatol* 21: 301–307.
26. Kishimoto T, Akira S, Tani T (1992) Interleukin-6 and its receptor: a paradigm for cytokines. *Science* 258: 593–597.
27. Nickoloff BJ, Naidu Y (1994) Perturbation of epidermal barrier function correlates with initiation of cytokine cascade in human skin. *J Am Acad Dermatol* 30: 535–546.
28. Wood LC, Jackson SM, Elias PM, Grunfeld C, Feingold KR (1992) Cutaneous barrier perturbation stimulates cytokine production in the epidermis of mice. *J Clin Invest* 90: 482–487.
29. Wood LC, Elias PM, Calhoun C, Tsai JC, Grunfeld C, et al. (1996) Barrier disruption stimulates interleukin-1 alpha expression and release from a preformed pool in murine epidermis. *J Invest Dermatol* 106: 397–403.
30. Antoniadis HN, Galanopoulos T, Neville-Golden J, Kiritsy CP, Lynch SE (1993) Expression of growth factor and receptor mRNAs in skin epithelial cells following acute cutaneous injury. *Am J Pathol* 142: 1099–1110.
31. Luetke NC, Qiu TH, Peiffer RL, Oliver P, Smithies O, et al. (1993) TGF alpha deficiency results in hair follicle and eye abnormalities in targeted and waved-1 mice. *Cell* 73: 263–278.
32. Mann GB, Fowler KJ, Gabriel A, Nice EC, Williams RL, et al. (1993) Mice with a null mutation of the TGF alpha gene have abnormal skin architecture, wavy hair, and curly whiskers and often develop corneal inflammation. *Cell* 73: 249–261.
33. Rosenthal A, Lindquist PB, Bringman TS, Goeddel DV, Derynck R (1986) Expression in rat fibroblasts of a human transforming growth factor-alpha cDNA results in transformation. *Cell* 46: 301–309.
34. Szulgit G, Rudolph R, Wandel A, Tenenhaus M, Panos R, et al. (2002) Alterations in fibroblast alpha1beta1 integrin collagen receptor expression in keloids and hypertrophic scars. *J Invest Dermatol* 118: 409–415.
35. Kose O, Waseem A (2008) Keloids and hypertrophic scars: are they two different sides of the same coin? *Dermatol Surg* 34: 336–346.
36. Bock O, Yu H, Zitron S, Bayat A, Ferguson MW, et al. (2005) Studies of transforming growth factors beta 1–3 and their receptors I and II in fibroblast of keloids and hypertrophic scars. *Acta Derm Venereol* 85: 216–220.
37. Lee TY, Chin GS, Kim WJ, Chau D, Gittes GK, et al. (1999) Expression of transforming growth factor beta 1, 2, and 3 proteins in keloids. *Ann Plast Surg* 43: 179–184.
38. Xia W, Phan TT, Lim IJ, Longaker MT, Yang GP (2004) Complex epithelial-mesenchymal interactions modulate transforming growth factor-beta expression in keloid-derived cells. *Wound Repair Regen* 12: 546–556.
39. Lu L, Saulis AS, Liu WR, Roy NK, Chao JD, et al. (2005) The temporal effects of anti-TGF-beta1, 2, and 3 monoclonal antibody on wound healing and hypertrophic scar formation. *J Am Coll Surg* 201: 391–397.
40. Singer AJ, Clark RA (1999) Cutaneous wound healing. *N Engl J Med* 341: 738–746.
41. Aktas GK, R. (2000) Ultrastructural immunolocalization of basic fibroblast growth factor in fibroblasts and extracellular matrix. *Histochemistry & Cell Biology*. pp 227–233.
42. Corral CJ, Siddiqui A, Wu L, Farrell CL, Lyons D, et al. (1999) Vascular endothelial growth factor is more important than basic fibroblastic growth factor during ischemic wound healing. *Arch Surg* 134: 200–205.
43. Richard JL, Parer-Richard C, Daures JP, Clouet S, Vannereau D, et al. (1995) Effect of topical basic fibroblast growth factor on the healing of chronic diabetic neuropathic ulcer of the foot. A pilot, randomized, double-blind, placebo-controlled study. *Diabetes Care* 18: 64–69.
44. Atiyeh BS, Gunn SW, Hayek SN (2005) State of the art in burn treatment. *World J Surg* 29: 131–148.
45. Wood FM, Kolybaba ML, Allen P (2006) The use of cultured epithelial autograft in the treatment of major burn wounds: eleven years of clinical experience. *Burns* 32: 538–544.
46. Pellegrini G, Bondanza S, Guerra L, De Luca M (1998) Cultivation of human keratinocyte stem cells: current and future clinical applications. *Med Biol Eng Comput* 36: 778–790.