

Receptor Tyrosine Kinase and p16/CDKN2 Expression in a Case of Tripe Palms Associated with Non-Small-Cell Lung Cancer

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Key Words

Tripe palms · Paraneoplastic disease · Epidermal growth factor receptor · HER2 · HER3 · SRC · p16/CDKN2 · *c-myc* oncogene · Reverse-transcription polymerase chain reaction · Non-small-cell lung cancer

Abstract

Background: Tripe palms is a descriptive term for a cutaneous paraneoplastic keratoderma. Tripe palms are frequently associated with gastric and pulmonary carcinoma. The pathogenetic mechanism remains unknown.

Objective: To determine the influence of receptor tyrosine kinases, which are both expressed in pulmonary carcinomas and in human skin, we performed expression studies on epidermal growth factor receptor (EGFR), HER2, HER3 in a skin sample of tripe palms obtained from a patient with non-small-cell lung cancer with lymph node involvement. Two months after diagnosis, the patient had developed palmoplantar 'tripe palms'. Additionally, the expression of SRC, *c-myc* and p16/CDKN2 were studied. **Method:** Conventional reverse-transcription polymerase chain reaction was performed on a tissue sample of tripe palms. **Results:** Weak expression of HER2 and of p16/CDKN2 was found. EGFR, HER3, *c-myc* and SRC were not expressed. **Conclusion:** Receptor tyrosine kinases of subclass I, the tyrosine kinase SRC and the oncogene *c-myc* play no major role in the pathogenesis of this case of tripe palms.

Tripe palms is a descriptive term for a rare cutaneous paraneoplastic keratoderma referring to the rugose surface of bovine foregut, which when prepared as food is known as tripe. The term was first introduced in 1977 by J. Clark [1]. The disease is characterized by thickening of the epidermis leading to a velvety texture and the accentuation of the normal dermatoglyphic ridges and sulci of palms and soles. Tripe palms are most frequently associated with gastric cancer (27%) and, as in our case, pulmonary carcinoma (22%) [2, 3]. Approximately 80 cases of tripe palms [1–8] have been described in the literature, not only in association with gastric and pulmonary carcinoma but also with e.g. melanomas, lymphomas, carcinomas of the entire gastrointestinal tract and urogenital tract, mammary carcinomas and carcinomas of the central nervous system. The isolated occurrence of tripe palms is more frequently found in pulmonary carcinoma, whereas 55% of patients with the combination of tripe palms and acanthosis nigricans suffer from gastric carcinoma [9]. In contrast to our patient, tripe palms in most cases develop prior to the diagnosis of the malignant disease [10]. Tripe palms are known to be a paraneoplastic disease, the pathogenesis, however, remains obscure. It has been speculated that the human growth hormone or related proteins, epidermal growth factor (EGF) or tissue growth factor α (TGF- α) may cause tripe palms [1, 11–13]. In order to elucidate the pathogenetic role of receptor tyrosine kinases (RTKs), which represent binding sites for EGF and TGF- α , we studied the expression of EGF receptor (EGFR), HER2, HER3 as well as SRC, a tyrosine kinase activated by RTKs in the signal transduction process. The oncogene

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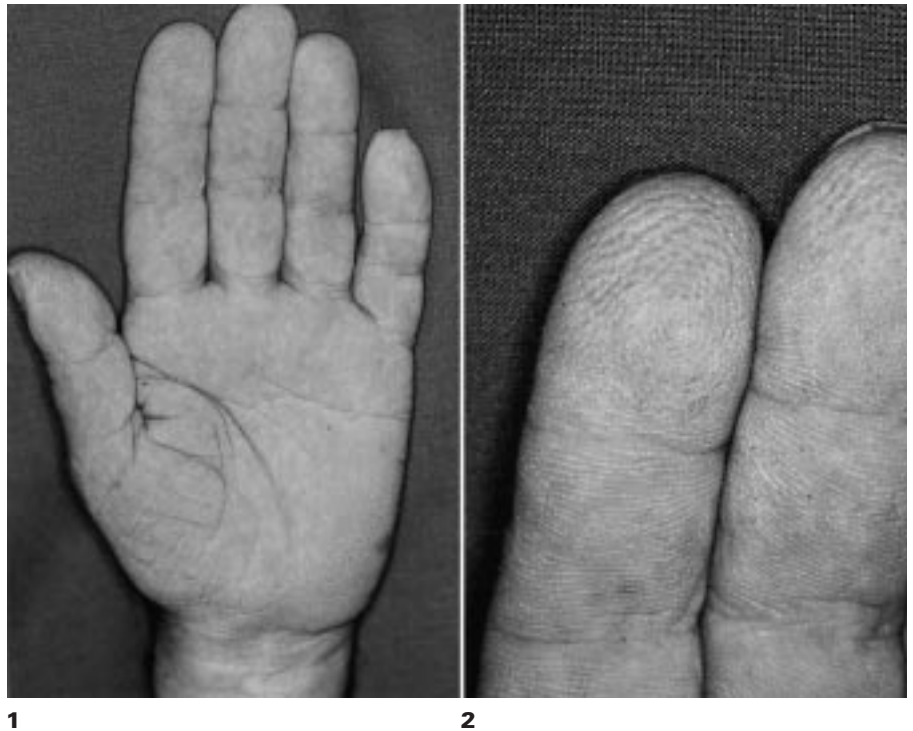


Fig. 1. Tripe palms, palmar aspect.

Fig. 2. Accentuation of the normal dermatoglyphic ridges and sulci.

c-myc involved in the regulation of apoptosis as well as the expression of p16/CDKN2, a tumour suppressor gene, were investigated.

Patient and Methods

Case Report

A 67-year-old Caucasian presented with chronic obstructive pulmonary disease, due to nicotine abuse and a mediastinal tumour, diagnosed as a non-small-cell lung cancer (NSCLC) with infiltration of the ganglion cervicale superius as well as cervical and mediastinal lymph nodes. Two months after diagnosis the patient developed hyperkeratotic, light yellow, velvety, non-pruritic skin changes starting on the palms followed by similar less severe skin alterations on the soles. No surgical therapy on the primary tumour or the lymph nodes was performed. Both chemotherapy (carboplatin and paclitaxel) and radiation therapy were performed, leading to a reduction of tumour size of 25% and a 50% reduction of lymph node involvement. Despite this partial remission the hyperkeratotic skin alterations remained unchanged, arguing for a continuing stimulus which – in this case – was independent of tumour size. Routine laboratory findings and human growth hormone were within the normal range. There were no endocrinological disorders such as acromegaly, Cushing's syndrome or diabetes mellitus.

On the palms and, to a lesser degree, the soles, a yellowish, velvety hyperkeratosis with accentuation of the dermatoglyphic lines with isolated hyperkeratotic papules and 'pits' was seen. Clubbing of the distal fingers, symmetric onychodystrophy and palmar hyperhidrosis were diagnosed (fig. 1, 2). There were no signs of acanthosis nigricans on the trunk.

Dermatopathology revealed regular acanthosis and psoriasiform papillomatosis with orthohyperkeratosis. PAS staining was negative.

Considering the poor prognosis of lung cancer (fig. 3), we did not administer etretinate [14, 15]. There was considerable improvement using a topical application of salicylic acid and urea-containing ointments.

Reverse-Transcription Polymerase Chain Reaction Analysis of *EGFR*, *HER2*, *HER3*, *C-myc*, *SRC* and *p16/CDKN2*

To elucidate the role of RTKs as well as *SRC*, *c-myc* and *p16/CDKN2* in this case of tripe palms, we performed conventional reverse-transcription polymerase chain reaction (RT-PCR) on a punch biopsy from the left palm (fig. 4).

Material

An N₂-frozen tissue specimen taken from the left palm of this male Caucasian suffering from tripe palms was studied. Since no surgery was performed on the NSCLC, an expression study on the underlying tumour tissue was impossible.

Methods

The tissue sample was shock frozen in liquid nitrogen immediately after excision. The tissue was minced, total mRNAs were isolated from homogenates using the RNA-Clean™ System (Angewandte Gentechologie Systeme GmbH, Germany). Using oligo (dt) primers, the extracted mRNA was transcribed with the Reverse Transcriptase System (Promega) to cDNA. The cDNAs were then phenol/chloroform extracted, precipitated by ethanol extraction and redissolved in ddH₂O. The cDNA concentration was measured by absorbance at 260 nm. In subsequent PCR reactions (25 µl), 100 ng cDNA template from above was used to investigate the expression of the following genes. The



Fig. 3. Chest X-ray, NSCLC (mediastinal).

sense and antisense strand primers and cycling parameters used to detect transcripts were as follows:

EGFR: 5'-ACT AGC CAG GAA CTA CTT CC-3' and 5'-GGC CTT CTT GGA TCT TTA GT-3', predicted product size: 398 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 60 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min); HER2: 5'-CGG GAG ATC CCT GAC CTG CTG GAA-3' and 5'-CTG CTG GGG TAC CAG ATA CTC CTC-3', predicted product size: 300 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 60 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min); HER3: 5'-CUA CUA CUA CUA GAG GCT GAG CTC CAG GAG AA-3' and 5'-CAU CAU CAU CAU CTG GGA CCT GGG AGA GAG AG-3', predicted product size: 814 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 56 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min); SRC: 5'-CAG ACC TGT CCT TCA AGA AA-3' and 5'-TCA GCC TGG ATG GAG TCG GA-3', predicted product size: 162 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 59 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min); *c-myc*: 5'-CCA GCA GCG ACT CTG AGG-3' and 5'-CCA AGA CGT TGT GTG TTC-3', predicted product size: 345 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 60 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min); p16/CDKN2: 5'-ATC CTG ACT GCT GTC ATG GC-3' and 5'-ACT CTT GGA AGT CCA CCT CG-3', predicted product size: for cDNA 238 bp and genomic DNA 959 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 60 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min).

The PCR products had been sequenced.

The PCR mixtures contained 100 ng of template cDNA, 1.5 mM MgCl₂, 1 μM/25 μl sense and antisense primer, 0.2 mM dNTPs (Promega) and *Taq* polymerase 1 unit (Boehringer, Mannheim, Germany). To circumvent false-negative results, we used primers detecting the GAPDH gene (*Stratagene*); sense 5'-CCA CCC ATG GCA AAT TCC

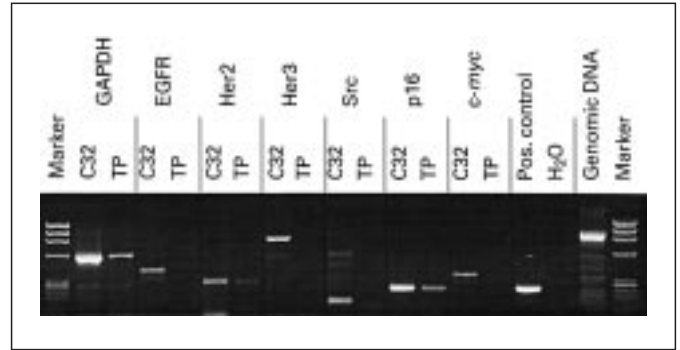


Fig. 4. PCR products. TP = Tripe palms.

ATG GCA-3'; antisense 5'-TCT AGA CGG CAG GTC AGG TCC ACC-3', predicted product size: 598 bp, cycling parameters: a first heating step (94 °C, 4 min), followed by 93 °C for 35 s, 60 °C for 35 s, 72 °C for 35 s, 35 cycles and one final extension step (68 °C, 10 min). The DNA molecular-weight standard (*PhiX 174RF/HaeIII*) was purchased from Boehringer Mannheim. RNA from the melanoma cell line C32, obtained from the American Type Culture Collection, was amplified as a positive control. The cell line was cultured as recommended by the American Type Culture Collection. Water instead of cDNA was applied as a negative control. In parallel genomic DNA was amplified. Following PCR, identical aliquots (5 μl) were loaded on 2% agarose gels and the products were visualized by ethidium bromide staining. The optical density of bands was analysed by a computerized densitometric analysis device (*Image master VDS*, Pharmacia). The expression level of the genomic DNA was taken as a level for determining 'weakly positive' or 'positive' expression. Optical density in the ethidium bromide gel lower than the genomic DNA band was regarded as 'weak', similar or higher as 'positive'. Genomic contamination was ruled out by the use of p16/CDKN2 primers with different product sizes for cDNA (238 bp) and genomic DNA (959 bp).

Results

The housekeeping gene GAPDH was strongly expressed. Weak expression of HER2 was detectable in the tripe palms sample, while EGFR and HER3 expression was not found. In 105 previously tested samples of normal skin, naevi and malignant melanoma, HER2 expression was detectable in all samples regardless of the dignity of the tissue (data not shown).

SRC, a tyrosine kinase involved in signal transduction downstream of RTKs [16], was not detectable either.

c-myc, an oncogene involved in a variety of malignant tumours [17], was not expressed. p16/CDKN2 [18, 19], a tumour suppressor gene [20], was moderately expressed. Since different product sizes for cDNA and genomic DNA are produced with these applied primers, genomic contamination could be ruled out.

Discussion

It has been speculated that the human growth hormone or related proteins may cause tripe palms [1]. In 2 cases, high levels of immunoreactive human growth hormone were detected. The elevated level of this hormone may act as a mediator for the hypertrophic processes. Elevated levels of immunoreactive human growth hormone have also been described in patients with lung cancer. Since elevated human growth hormone as well as plasma growth-hormone-releasing hormone have been detected in serum of patients both with small-cell and non-small-cell lung carcinoma [21] without tripe palms, the development of tripe palms is not necessarily caused by the human growth hormone.

Based on the observation of an elevated EGF level in a patient with tripe palms in association with bronchial carcinoma, the role of tumour growth factors such as EGF has been discussed [11]. The overexpression of EGF in the serum of this published patient [11] could be attributable to the underlying malignant process, since elevated EGF levels were also detected in patients with lung carcinoma [22] without tripe palms.

Publications attributing a pathogenetic influence of the TGF- α to the development of acanthosis nigricans, a paraneoplastic skin disease with similarity to tripe palms, are based on the detection of TGF- α in tumour tissue but not in the skin samples [12, 13]. A case report on a patient with systemic mastocytosis and tripe palms describes elevated serum levels of TGF- α and mast cells in the skin [8].

A recent concept [16] assumes that the activation of membrane-based receptor tyrosine kinases by binding of ligand such as EGF or TGF- α leads to the induction of signal transduction processes resulting in cell proliferation and differentiation. An elevated level of EGF or TGF- α should thus result in the expression of corresponding receptors such as the EGFR, expressed in a variety of malignant tumours including pulmonary carcinoma [23] and melanoma [24], or HER2, an important oncogene in mammary carcinoma [25, 26], or HER3 [27, 28], a closely related member of this gene family [29], expressed among others in head and neck tumours. These RTKs of subclass I are also designated erbB1–3. A further member, HER4 [30], has not yet been extensively studied in human carcinomas in vivo.

Based on these data, we investigated the role of RTKs of subclass I, which could be putative receptors for ligands such as EGF in this case of tripe palms.

Recent studies provide evidence that the family I of RTKs can associate with each other to form an array of heterodimer complexes [31–36]. Therefore, the activation of one receptor could modulate the activity of the other mem-

bers of this group of RTKs when co-expressed in a cell and extend the repertoire of downstream signalling events. HER2 can – in the absence of any HER2-specific ligand – be transmodulated within a heterodimer complex by associating with other HER family members. Tzahar et al. [31] came to the conclusion that HER2 might serve as the preferred heterodimerizing partner of the three other ERBB proteins.

In this case of tripe palms EGFR and HER3 are not involved in this signal transduction process. HER2 is expressed in this case of tripe palms. Isolated expression of HER2 could theoretically – by homodimerization – be sufficient to activate signal transduction downstream of the RTKs. HER4 [30], a further RTK of subclass I which we did not test, as well as other yet unidentified RTKs might be possible candidates for further partners for heterodimerization with HER2. However, HER4 has not yet been described in adult skin or any skin cancer [37]. The lack of SRC expression hints at another signal transduction pathway activated by RTKs than the one via SRC.

HER2 expression both in vitro and in vivo has been described in NSCLC [38–42]. We found weak to strong HER2 expression in more than 100 skin samples of varying dignity (data not shown), and frequent HER2 protein expression has been described both in benign and malignant cutaneous tissue [43–45]. Despite the known HER2 expression in NSCLC, but due to the frequent HER2 expression in cutaneous tissue and its weak expression in this particular case, HER2 might not be a causative factor for NSCLC-associated paraneoplastic skin diseases, such as tripe palms.

c-myc, a transcription factor that is believed to regulate the expression of genes governing cellular growth, differentiation and apoptosis [17, 46, 47] is – due to a lack of its expression – presumably not involved in regulatory mechanisms in this case of tripe palms.

Interestingly, p16/CDKN2, a cell cycle regulator with tumour suppressor activity [18–20], among others in malignant melanoma, was expressed in this case of tripe palms. The role of further cell cycle regulators with tumour suppressor gene potential such as p53 need to be elucidated in further studies.

Our data demonstrate that EGFR, HER3, *c-myc* and SRC are not involved in the pathogenesis of this case of tripe palms. Whether HER2 homodimer formation or the possible involvement of further RTKs [32] is responsible for tripe palms will have to be studied. Except for ligands such as EGF or TGF- α , which have been detected in patients with tripe palms, yet uninvestigated RTK ligands such as heregulin [48] might furthermore play a role in the pathogenesis of tripe palms.

Although the pathogenetic mechanisms causing tripe palms remain unclear, further studies on gene expression of growth factors and their ligands, with special respect to those involved in the underlying malignant neoplasias, may help to understand the intriguing pathways leading to paraneoplastic skin diseases.

From a clinical point of view the knowledge of tripe palms is crucial, since in 40% of patients, tripe palms are the presenting feature of a previously undiagnosed malignant

neoplasia [10]. The observation of tripe palms should thus always prompt a meticulous search for an underlying malignancy.

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