

Oxidation of an Oil Rich in Docosahexaenoic Acid Compared to Linoleic Acid in Lactating Women

N. Fidler T. Sauerwald H. Demmelmair A. Pohl B. Koletzko

Division of Metabolic Disorders and Nutrition, Kinderklinik and Kinderpoliklinik,
Dr. von Haunersches Kinderspital, Ludwig Maximilians University, Munich, Germany

Key Words

Oxidation · Docosahexaenoic acid ·
Linoleic acid · ^{13}C -labeled fatty acids ·
Isotopic ratio mass spectrometry · Lactation

Abstract

Background: We studied the oxidation of an oil rich in docosahexaenoic acid (DHA; DHASCO[®]) in lactating mothers receiving a dietary DHA supplement or a placebo. The results were compared with the oxidation of linoleic acid. **Methods:** Breast-feeding mothers received a dietary supplement (DHASCO; 200 mg DHA/day, n = 5) or a placebo (n = 5) for 14 days. Six weeks post partum all 10 mothers received a single dose of 2 mg/kg body weight uniformly ^{13}C -labeled DHASCO. In a previously reported study 6 mothers received 1 mg/kg body weight uniformly ^{13}C -labeled linoleic acid. Breath samples were collected over 48 h after tracer application.

The total CO_2 production was measured by indirect calorimetry and the ^{13}C isotopic enrichment of labeled CO_2 by isotopic ratio mass spectrometry. **Results:** The oxidation of ^{13}C -labeled DHASCO in the supplemented and placebo groups was similar. Maximal ^{13}C enrichment was reached earlier in the group receiving ^{13}C -DHASCO (median 1.0 vs. 3.0 h in the linoleic acid group). The cumulative ^{13}C recovery in breath was higher in the DHASCO versus the linoleic acid group until 10 h after tracer application and comparable thereafter. **Conclusions:** The difference in oxidation of DHASCO versus linoleic acid after tracer ingestion might be partly due to a faster absorption and oxidation of shorter chain saturated fatty acids contained in DHASCO. The cumulative oxidation of DHASCO and linoleic acid 24 and 48 h after tracer ingestion is similar.

Copyright © 2000 S. Karger AG, Basel

KARGER

Fax +41 61 306 12 34
E-Mail karger@karger.ch
www.karger.com

© 2000 S. Karger AG, Basel
0250-6807/99/0436-0339\$17.50/0

Accessible online at:
www.karger.com/journals/anm

Berthold Koletzko, MD, Division of Metabolic Diseases and Nutrition
Kinderklinik and Kinderpoliklinik, Dr. von Haunersches Kinderspital
Ludwig Maximilians University, Lindwurmstrasse 4
D-80337 München (Germany), Fax +49 89 5160 3336
E-Mail Berthold.Koletzko@kk-i.med.uni-muenchen.de

Introduction

The supply of essential fatty acids and long-chain polyunsaturated fatty acids (LCP) is important for normal growth and development of infants. Their contents in human milk depend on maternal nutrition, utilization, and composition of depot fats and endogenous LCP synthesis from precursors [1–3]. Essential fatty acids and LCP may also be used as an energy source and thus be oxidized to CO₂. Methods for measuring the *in vivo* oxidation of substrates labeled with the non-radioactive isotope ¹³C have been increasingly used for diagnostic and research purposes [4–6]. Small excesses of exogenous ¹³CO₂ arising from oxidation of the labeled substrate can be measured by sensitive mass spectrometry techniques [5, 6].

The first objective of this study was to investigate the oxidation of an orally applied ¹³C-labeled oil rich in docosahexaenoic acid (DHA; DHASCO®, Martek Biosciences, Columbia, Md., USA) in a group of lactating mothers supplemented either with DHA or a placebo oil. The second objective was to compare the oxidation of this fatty acid mixture containing about 40% DHA with the oxidation of a single polyunsaturated fatty acid (linoleic acid) in lactation.

Subjects and Methods

Ten healthy breast-feeding mothers were recruited after term delivery at the Department of Gynecology and Obstetrics, University of Munich. The study was approved by the Ethical Committee of the Medical Faculty of the University of Munich, and written consent was obtained from the participating women.

The women were 31 ± (SD) 5 years old and had a weight of 71.4 ± 4.6 kg (at 4 weeks post partum) and a height of 170 ± 6 cm. All were consuming omnivorous diets which were assessed by 7-day weighted dietary protocols. At 4 weeks post partum (study day 0), they were randomly and blindly assigned to receive either a

Table 1. Fatty acid composition (w%) of the capsules with the dietary supplement DHASCO oil, the ¹³C-labeled DHASCO oil, and the placebo capsules containing a 1:1 mixture of corn and soy oils

Fatty acid	DHASCO capsules ^a	¹³ C-labeled DHASCO ^a	Placebo capsules ^b
C8:0	–	0.0–0.3	–
C10:0	–	1.1–1.8	–
C12:0	5.4	7.8–9.3	–
C14:0	19.40	18.1–19.6	–
C16:0	17.5	9.0–13.2	11.0
C18:0	0.4	0.0–0.3	3.0
C20:0	–	0.1	0.4
C16:1n-7	1.7	0.9–1.0	–
C18:1n-9	9.5	5.1–7.4	23.3
C20:1n-9	–	0.0–0.3	0.3
C18:2n-6	0.6	–	56.7
C18:3n-3	–	–	4.1
C22:5n-3	0.3	0.2–0.4	–
C22:6n-3	45.1	49.3–55.4	–
C24:1n-9	–	–	–

^a Analysis in our laboratory (analysis of ¹³C tracer represents the range from three different lots).

^b Compositional data provided by the manufacturer.

dietary supplement rich in DHA (200 mg DHA/day from DHASCO capsules) or a placebo oil for 14 days. The mothers were asked not to eat fish (rich in n-3 LCP) during the 2-week supplementation period. Further, they were asked not to eat corn-based products the day before tracer application because of an elevated ¹³C content. At 6 weeks post partum mothers from both groups received with their breakfast a single dose of 2 mg/kg of uniformly ¹³C-labeled DHASCO oil (57.1–65.7% ¹³C enrichment). The fatty acid compositions of the DHASCO and placebo capsules as well as the tracer are shown in table 1. In a previously published comparable study [1], 6 breast-feeding mothers (age 31 ± 3 years, weight 68.17 ± 8.2 kg, height 167 ± 3 cm; mean ± SD) received a single dose of uniformly ¹³C-labeled linoleic acid (~98% ¹³C labeling; Martek Biosciences) at 6 weeks post partum.

Exhaled breath samples were collected into 1-liter breath bags. An aliquot of 11 ml was transferred into evacuated glass tubes (Labco, Manchester, UK) and

stored until measurement of the ^{13}C content of the CO_2 . Immediately before tracer application a breath sample was taken in triplicate in order to measure baseline ^{13}C enrichment of CO_2 . Further samples were collected 30 min and 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 12, 24, 36, and 48 h after tracer administration. The total CO_2 production was measured by indirect calorimetry twice in each woman from 30 min preprandial until 30 min postprandial (Deltatrac[®] II MBM-200; Hoyer, Bremen, Germany). The ^{13}C enrichments of breath CO_2 were measured in duplicate by isotopic ratio mass spectrometry (Delta S; Finnigan MAT, Bremen). Carbon isotope ratios were expressed as $\delta^{13}\text{C}$ values (‰) versus the Pee Dee Belemnite standard. The oxidation rates over 48 h were calculated as described elsewhere [1].

Statistical differences between the DHA-supplemented group and the placebo group as well as between the groups combined (supplemented and placebo groups; $n = 10$) and the group given ^{13}C linoleic acid were assessed by a nonparametric test (Mann-Whitney, Monte Carlo) using the Stastical Package for the Social Sciences (version 8.0; SPSS, Chicago, Ill., USA). This test was chosen due to the relatively small number of subjects per group. The significance level was set at $p < 0.05$, and the results are presented as median and interquartile range (IQR).

Results

The times of peak ^{13}C enrichment in breath CO_2 (delta over baseline, DOB) of the DHA-supplemented group and the placebo group were at 1 h (1.0–1.0; median and IQR) and at 2 h (1.0–3.3), respectively (table 2). Forty-eight hours after tracer application ^{13}C enrichment in both groups had almost returned to baseline values [DOB 0.21‰ (0.04–0.59)]. Because of the nearly identical kinetics of the supplemented and the placebo group, both were combined for comparison with the linoleic acid group.

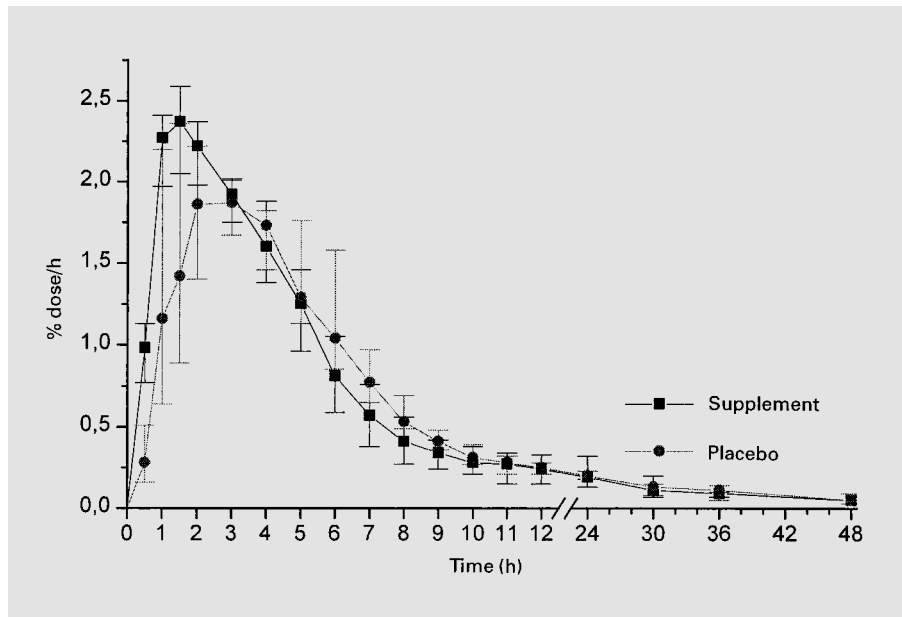
Time of peak ^{13}C enrichment in the combined group was at 1 h (1.0–2.0) after tracer application which was about 3 h earlier than in the linoleic acid group [3.5 h (3.0–4.0); $p = 0.003$].

The percentage of the dose of tracer ^{13}C exhaled in CO_2 per hour (percent dose recovery; PDR/h) tended to be slightly higher during the first 2 h after tracer application in the DHA-supplemented versus the placebo group which was not significantly different due to large interindividual differences (fig. 1). From 3 to 48 h after tracer application, the tracer oxidation did not differ between the supplemented and the placebo groups.

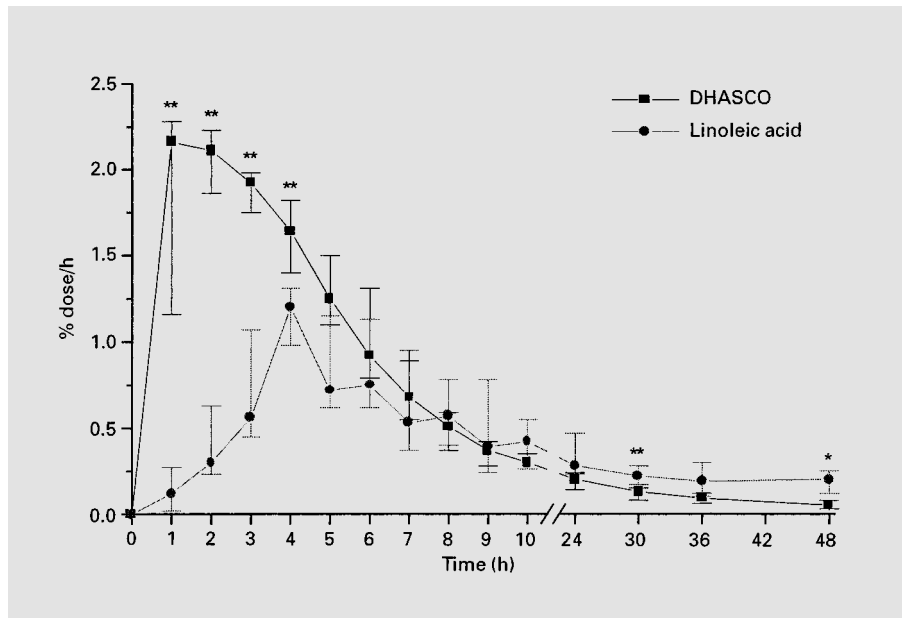
A comparison of the combined group and the linoleic acid group showed a severalfold higher PDR/h in the group receiving labeled DHASCO during the first 4 h after tracer application: 18-fold higher at 1 h after tracer ingestion, 7-fold higher at 2 h, 3.4-fold higher at 3 h, and 1.6-fold higher at 4 h (fig. 2). At the later time points, 5–24 h after tracer application, there were no significant differences between the combined and the linoleic acid group. At 30 and 48 h after tracer application, the PDR/h in the linoleic acid group versus the combined group was higher [30 h: 0.22 (0.17–0.28) vs. 0.13 (0.08–0.15), $p = 0.003$; 48 h: 0.20 (0.12–0.25) vs. 0.05 (0.03–0.08), $p = 0.026$; see figure 2).

The cumulative PDR/h of tracer doses in breath tended to be higher in the DHA-supplemented group than in the placebo group during the first 2 h after tracer application. This difference was significant only at 1 h after the tracer application [1.61% (1.38–1.77) in the supplemented vs. 0.76% (0.40–1.15) in the placebo group, $p = 0.033$].

The cumulative recovery of ^{13}C in the combined group at 1 h after tracer ingestion was 10.7-fold higher than in the linoleic acid group (1.28 vs. 0.12). This tendency was observed during the first 10 h after tracer application. From 24 to 48 h after tracer application, the cumulative recovery of DHASCO and linoleic acid was very similar and reached about 15% of the ingested doses (fig. 3).



1



2

Fig. 1. Time course of ^{13}C in breath CO_2 in breast-feeding mothers receiving DHA supplementation ($n = 5$) or a placebo ($n = 5$) after oral application of 2 mg/kg body weight of U- ^{13}C -labeled DHASCO, expressed as percentage of tracer dose ingested per hour (PDR/h). Median and IQR.

Fig. 2. Time course of ^{13}C in breath CO_2 in breast-feeding mothers after oral tracer application of ^{13}C -DHASCO ($n = 10$) or ^{13}C -linoleic acid ($n = 6$) [1]. Median and IQR. * $p < 0.05$; ** $p < 0.01$.

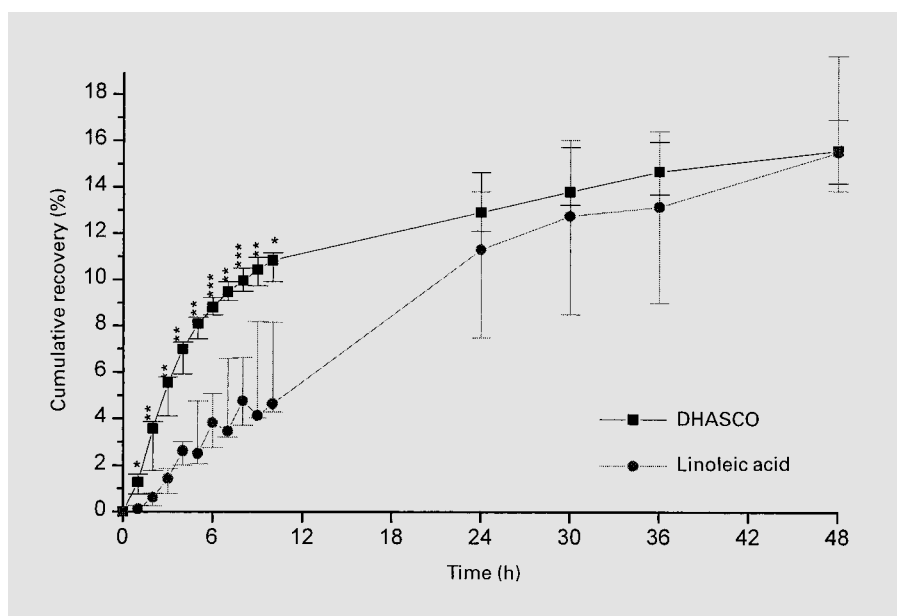


Fig. 3. Time course of cumulative recovery of ^{13}C in breath CO_2 in breast-feeding mothers after oral tracer application of ^{13}C -DHASCO ($n = 10$) or ^{13}C -linoleic acid ($n = 6$) [1]. Median and IQR. * $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

Table 2. Time point of $\delta^{13}\text{C}$ peak and peak $\delta^{13}\text{C}$ values (‰) in breath CO_2 of lactating women after oral administration of ^{13}C -DHASCO and ^{13}C -linoleic acid [1] at 6 weeks post partum

Supplement DHASCO ^{13}C -DHASCO ($n = 5$)		Placebo ^{13}C -DHASCO ($n = 5$)		Regular diet ^{13}C -linoleic acid ($n = 6$)	
subject No. h		subject No. h		subject No. h	
<i>Time point of $\delta^{13}\text{C}$ peak (h after administration)</i>					
1	1	6	1	11	3
2	1	7	1	12	4
3	1	8	2	13	5
4	1	9	3	14	3
5	1	10	4	15	3
<i>Peak $\delta^{13}\text{C}$ value, ‰</i>					
1	22.47	6	22.47	11	9.95
2	29.04	7	19.14	12	8.00
3	24.88	8	17.79	13	5.95
4	22.18	9	17.62	14	8.00
5	20.01	10	15.80	15	10.20
				16	13.10

Discussion

In this study, we investigated the oxidation of an oil rich in DHA and compared these results with data obtained in a previous study on linoleic acid oxidation in lactating women.

DHASCO is a refined single-cell triglyceride oil highly enriched in DHA that is produced by the microalgal strain MK 8805 [7]. Due to positional preference of DHA for sn-1 and sn-3, it is more similar to mammal than to fish oil. It is used as a dietary supplement to elevate the levels of DHA in conventional foods, especially in infant formulas [7, 8]. DHASCO labeled with ^{13}C , a safe nonradioactive isotope that causes no radiation risk to mother or infant [4, 9, 10], was used to measure fatty acid oxidation. Breath tests using stable isotopic labeled substrates are generally considered as safe and noninvasive tests to measure *in vivo* substrate oxidation [6, 11–13].

The baseline ^{13}C content in breath CO_2 of mothers from the combined group was 1.0838 atom percent (1.0837–1.0839; median and IQR) and achieved peak ^{13}C enrichment of 1.1051 atom percent (1.1027–1.1082) which represents <2% enrichment over the baseline. The baseline ^{13}C content in breath CO_2 measured in our study is slightly lower than the natural average ^{13}C abundance reported in Europeans (1.088) [14]. This is a reflection of the local Bavarian diet, with little intake of maize and cane sugar, but high in proportions of potato, cabbage, rice, and beet sugar, as well as the overnight fasting.

Our results show no apparent difference in the kinetics and the cumulative oxidation rate between the groups of women that were supplemented or not supplemented with DHASCO over a period of 14 days. The direct measurement of DHA oxidation was not intended in this study. This would have only

been possible using a pure DHA tracer, instead of a mixture of different fatty acids.

We combined the results of the DHA-supplemented and the placebo groups and compared the oxidation of a single oral dose of ^{13}C -DHASCO with the results of the ^{13}C -linoleic acid oxidation obtained in a previous study [1]. Time points of sampling exhaled air and the measurement of ^{13}C enrichment of breath CO_2 were performed in the same way and using the same instruments as in the current study.

Earlier maximum ^{13}C enrichment in breath CO_2 (DOB) in the combined group versus the linoleic acid group (1.0 vs. 3.5; median) reflects the faster oxidation rate of the DHASCO tracer. Different kinetics of the oxidation of the two tracers can also be seen from the data of the PDR/h (fig. 1) and the cumulative recovery of the tracer in breath (fig. 3).

The oxidation rate of DHASCO tracer versus linoleic acid was faster, especially during the first 4 h after its ingestion. The ^{13}C -labeled DHASCO tracer is a triglyceride-oil mixture that contains twelve different fatty acids, with the main components being lauric (C12:0), myristic (C14:0), and palmitic acid (C16:0) and DHA (C22:6n-3) which represent 84.2–97.5 w% of all the fatty acids (table 1). From the oxidation data alone, it is difficult to speculate which fatty acids are the ones with faster oxidation in DHASCO as compared with linoleic acid. Cumulative ^{13}C enrichments of single fatty acids at 48 h in human milk of mothers from the combined DHASCO group were severalfold higher for C16:0, C16:1n-7, C18:1n-9, and C22:6n-3 than for of C14:0 [unpubl. data]. This could be explained by a greater rate of C14:0 oxidation in comparison with other mono- and polyunsaturated fatty acids contained in DHASCO. This preservation from oxidation is in agreement with the results of a study performed by Metges and

Wolfram [15] who found greater and more rapid oxidation of trioctanoate as compared with trioleate after oral and parenteral administration in healthy adult volunteers.

It is likely that the faster oxidation rate of DHASCO, as compared to linoleic acid, is due to a quicker absorption and oxidation of shorter-chain saturated fatty acids (C8:0, C10:0, C12:0, and C14:0) contained in DHASCO. The cumulative oxidation of DHASCO is about 15% of the dose at 48 h after ingestion. This value is comparable to values obtained in other studies measuring

fatty acid oxidation. Although there were differences in the kinetics of oxidation between the DHASCO group and the linoleic acid group, the cumulative oxidation 1 and 2 days after tracer application did not differ.

Acknowledgements

This study was supported in part by the Deutsche Forschungsgemeinschaft, Bonn, Germany (Ko 912/5-2), and Martek Biosciences, Columbia, Md., USA. Natasa Fidler was the recipient of a scholarship from Deutscher Akademischer Austauschdienst, Bonn.

References

- 1 Demmelmair H, Baumheuer M, Koletzko B, Dokoupil K, Kratl G: Metabolism of U¹³C-labeled linoleic acid in lactating women. *J Lipid Res* 1998;39:1389–1396.
- 2 Hachey DL, Thomas MR, Emken EA, Garza C, Brown-Booth L, Adolf RO, Klein PD: Human lactation: Maternal transfer of dietary triglycerides labeled with stable isotopes. *J Lipid Res* 1987;28:1185–1192.
- 3 Martin JC, Bougnoux P, Fignon A, Theret V, Antonie JM, Lamisse F, Couet C: Dependence of human milk essential fatty acids on adipose stores during lactation. *Am J Clin Nutr* 1993;58:653–659.
- 4 Koletzko B, Sauerwald T, Demmelmair H: Safety of stable isotope use. *Eur J Pediatr* 1997;156(suppl 1):12–17.
- 5 Koletzko B, Demmelmair H, Hartl W, Kindermann A, Koletzko S, Sauerwald T, Szitanyi P: The use of stable isotope techniques for nutritional and metabolic research in paediatrics. *Early Hum Dev* 1998; 53(supl):77–97.
- 6 Rating D, Langhans CD: Breath tests: Concepts, applications and limitations. *Eur J Pediatr* 1997; 156(suppl 1):18–23.
- 7 Kyle DJ, Sicotte VJ, Singer JJ, Reeb SE: Bioproduction of docosahexaenoic acid (DHA) by microalgae; in Kyle DJ, Ratledge C (eds): *Industrial Applications of Single Cell Oils*. Champaign, American Oil Chemists' Society, 1992, pp 287–300.
- 8 Boswell K, Koskelo EK, Carl L, Glaza S, Hensen DJ, Williams KD, Kyle DJ: Preclinical evaluation of single-cell oils that are highly enriched with arachidonic acid and docosahexaenoic acid. *Food Chem Toxicol* 1996;34:585–593.
- 9 Klein PD, Klein ER: Stable isotopes: Origins and safety. *J Clin Pharmacol* 1986;26:378–382.
- 10 Jones PJH, Leatherdale ST: Stable isotopes in clinical research: Safety reaffirmed. *Clin Sci (Colch)* 1991; 80:277–280.
- 11 Heiling VJ, Miles JM, Jensen MD: How valid are isotopic measurements of fatty acid oxidation? *Am J Physiol* 1991;261:E572–E577.
- 12 Schoeller DA, Schneider JF, Solomons NW, Watkins JB, Klein PD: Clinical diagnosis with the stable isotope ¹³C in CO₂ breath tests: Methodology and fundamental considerations. *J Lab Clin Med* 1997; 90:412–421.
- 13 Demmelmair H, Sauerwald T, Koletzko B, Richter T: New insights into lipid and fatty acid metabolism via stable isotopes. *Eur J Pediatr* 1997;156(suppl 1):70–74.
- 14 Krumbiegel P: *Stable Isotope Pharmaceuticals for Clinical Research and Diagnosis*. Jena, Fischer, 1991, pp 70–72.
- 15 Metges CC, Wolfram G: Medium- and long-chain triglycerides labeled with ¹³C: A comparison of oxidation after oral or parenteral administration in humans. *J Nutr* 1991;121: 31–36.