Review Article



Ann Nutr Metab 2013;63:208-215 DOI: 10.1159/000355222

Received: June 10, 2013 Accepted after revision: August 19, 2013 Published online: October 8, 2013

The Placental Exposome: Placental **Determinants of Fetal Adiposity and Postnatal Body Composition**

R.M. Lewis^a H. Demmelmair^b R. Gaillard^c K.M. Godfrey^{a, d} S. Hauguel-de Mouzon^e B. Huppertz^f E. Largue^g R. Saffery^h M.E. Symondsⁱ G. Desoye^f

^aUniversity of Southampton, UK; ^bLudwig Maximillian University Munich, Germany; ^cErasmus Medical Centre, $Amsterdam, The \ Netherlands; {}^{d}MRC \ Life course \ Epidemiology \ Unit \ and \ NIHR \ Southampton \ Biomedical \ Research$ Centre, University of Southampton and University Hospital Southampton NHS Foundation Trust, Southampton, UK; ^eCase Western Reserve University MetroHealth Medical Center, Cleveland, Ohio, USA; ^fMedical University of Graz, Austria; ⁹University of Murcia, Spain; ^hMurdoch Childrens Research Institute and University of Melbourne, Australia; ⁱUniversity of Nottingham, UK

Key Words

Fetal adiposity · Maternal obesity · Nutrient · Obesity · Placenta · Postnatal environment

Abstract

Offspring of obese and diabetic mothers are at increased risk of being born with excess adiposity as a consequence of their intrauterine environment. Excessive fetal fat accretion reflects additional placental nutrient transfer, suggesting an effect of the maternal environment on placental function. High plasma levels of particular nutrients in obese and diabetic mothers are likely to be the important drivers of nutrient transfer to the fetus, resulting in excess fat accretion. However, not all offspring of obese and diabetic mothers are born large for gestational age and the explanation may involve the regulation of placental nutrient transfer required for fetal growth. The placenta integrates maternal and fetal signals across gestation in order to determine nutrient transfer rate. Understanding the nature of these signals and placental responses to them is key to understanding the pathology of both fetal growth restriction and macrosomia. The overall effects of the maternal environment on the placenta are the product of its exposures throughout gestation, the 'placental exposome'. Understanding these environmental influences is important as exposures early in gestation, for instance causing changes in the function of genes involved in nutrient transfer, may determine how the placenta will respond to exposures later in gestation, such as to raised maternal plasma glucose or lipid concentrations. Longitudinal studies are required which allow investigation of the influences on the placenta across gestation. These studies need to make full use of developing technologies characterising placental function, fetal growth and body composition. Understanding these processes will assist in the development of preventive strategies and treatments to optimise prenatal growth in those pregnancies at risk of either excess or insufficient nutrient supply and could also reduce the risk of chronic disease in later life. © 2013 S. Karger AG, Basel

University of Southampton Faculty of Medicine

Tremona Road, Southampton SO16 6YD (UK)

Southampton General Hospital

E-Mail Rohan.Lewis@soton.ac.uk

Introduction

Maternal obesity and diabetes alter the intrauterine environment and increase the risk of large for gestational age (LGA) offspring. A disproportionate increase in fat mass makes an important contribution to the increased birth weight in LGA babies [1, 2], but can also be found in offspring born with weight appropriate for their gestational age [3]. Excess fetal fat accumulation may predispose these individuals to develop obesity in later life and raises the possibility of an intergenerational cycle of obesity [4]. Maternal obesity is associated with a doubling in the rate of LGA births compared to women of normal weight [5]. While shared genetics and environment will contribute to this relationship it is likely that factors specific to the in utero environment are also important. The role of intrauterine environment is illustrated by the observation that babies born to women following bariatric surgery have half the risk of becoming obese relative to their siblings born prior to surgery [6].

While the association between in utero environment and fetal adiposity is clear, with maternal pre-gravid obesity and diabetes being the strongest maternal determinants [1, 7], the mechanisms through which maternal factors increase fetal adiposity are not fully understood. However, as the principal organ through which nutrients are transferred to the fetus, the placenta is likely to be an important mediator of this process. The placenta is not simply a passive conduit for nutrients but responds to both maternal and fetal signals, altering placental transport and metabolic function [8, 9]. At any point in gestation, placental function will reflect a progressive accumulation of external influences experienced during the course of its development. These can be conceptualised as the 'placental exposome'. The exposome of an individual encompasses the product of lifetime environmental exposures starting in utero and varying according to life stage [10]. The in utero environment experienced by the placenta is itself the product of the maternal exposome, for instance effects of maternal obesity on the placenta reflect a historical mismatch between food intake and energy expenditure.

Ideally, prevention of obesity and diabetes should occur prior to pregnancy. However, where this is not possible, the development of successful interventions will be facilitated by a more detailed understanding of those pregnancies in which maternal obesity and diabetes leads to increased placental nutrient transfer and greater fetal adiposity.

This review is based on discussions at the workshop 'The Placenta and Its Role for Fetal and Neonatal Development', which took place in Austria, October 2012, supported by the EU FP7 project EarlyNutrition. Themes emerging from this workshop included: the central role of placental function in determining fetal growth and body composition, how placental function is determined by its exposome, the potential for epigenetics to act as a mediator of environmental influence on placental function, and ways in which new technologies can be incorporated into longitudinal population studies to investigate the role of the placenta in mediating maternal exposures on fetal growth and postnatal health.

Placental Structure, Function and Regulation

Placental nutrient transfer underpins fetal growth and development. The nutrient transfer capacity of a placenta will depend on its structure and fetal maternal blood flow, whereas actual transfer depends on how these factors and transporter density are regulated. This regulation can occur over long (e.g. structural or epigenetic changes in the placenta) and short (e.g. placental responses to maternal hormones or nutrients) time scales, and therefore it is important to understand all the determinants of placental function across gestation.

The growing fetus requires adequate and appropriate placental nutrient transfer but excessive nutrient transfer, or an inappropriate balance of nutrients, may cause the fetus to lay down excess fat and become LGA [1, 2]. In order to identify abnormal fetal growth, it is important to understand placental development, the mechanisms underlying placental function and their regulation.

The placenta has a complex anatomical structure which develops throughout gestation increasing its nutrient transfer capacity to match the requirements of the growing fetus [11]. The placental villi are the sites of nutrient exchange and, in broad terms, they form in the first trimester, branch in the second and assume their final structure during the third trimester. Once its final structure is established, the placenta remains a dynamic organ with continual turnover of villous trophoblasts as well as development and regression of placental vessels (fig. 1). This evolution throughout gestation means that disruptive factors can have persistent effects that will differ depending on their timing. The role of extravillous trophoblast cells, which associate with spiral arteries and uterine glands, is also important in mediating blood flow to the fetus [12]. Placental weight is a major determinant of fetal growth, reflecting a greater nutrient transport capacity. Maternal weight gain in early pregnancy has an influence on fetal weight, which is thought to be partly mediated

Sinusoid **Fig. 1.** Representation of the structures of a first-trimester immature intermediate villus (a, b) and a third-trimester terminal villus (c, d), a, b There is a double-layered epithelium with a thick syncytiotrophoblast (ST) and a complete layer of cytotro-Stroma phoblasts (CT). The distance between a capillary (Ca) and the outer surface of the ST is depicted by an arrow and is in the range of 50-60 µm at the beginning of MVM pregnancy. c, d There is only a thin layer of ST with some CT underneath. Especially in terminal villi, Ca transform into larger sinusoids that are located subjacent to the ST. Hence, the distance between fetal Sinusoid and maternal blood - as depicted by the arrows – is very short (1.3–4 µm). The shortening of the diffusion distance between the maternal and fetal circulation during the course of pregnancy facilitates the transport of nutrients and oxygen paralleling the Stroma increasing needs of the growing fetus. The Stroma MVM and BM of the ST are indicated by

through its effects on placental growth [13]. Consequently, in some obese mothers, enhanced fetal growth could be mediated through raised placental mass [14]. Blood flow through the maternal side of the placenta starts at the end of the first trimester. Prior to the initiation of placental blood flow, the villi are nourished by secretions from uterine glands and plasma filtrate [15]. Maternal blood delivers nutrients and oxygen to the placenta and any disruption of this flow may affect fetal nutrient supply. One of the major causes of impaired maternal blood supply is the failure of spiral artery remodelling [16]. Reduced blood flow through the placenta as a result of failure to remodel the spiral arteries may decrease nutrient availability to the fetus and may affect fetal development in the second and third trimesters [17]. Failure to remodel the

Excess placental transfer of glucose and lipids are prime candidates for promoting fetal adiposity. Transfer of glucose, lipids and amino acids across the placenta requires

spiral arteries early in gestation may therefore limit pla-

cental nutrient transfer capacity throughout gestation.

transport across a series of cell membranes by specific membrane transport proteins. Transport occurs across the maternal-facing microvillus membrane (MVM) and fetal-facing basal membrane (BM) of the placental syncytiotrophoblast (fig. 1). Glucose transfer is primarily through facilitated diffusion and is mediated by one family of transporters, the GLUTs, across both the MVM and the BM. There are multiple transporters for fatty acids and amino acids which are differentially localised to the MVM and BM [18]. Once across the BM, it is likely that glucose and amino acids can diffuse through endothelial junctions into the fetal circulation while fatty acids may also require mediated transport across the endothelium.

Metabolism of nutrients within the syncytiotrophoblast will affect fetal transfer and may be regulatory [19]. A proportion of the glucose taken up by the placenta is converted to lactate and in humans this lactate is primarily transported back to the mother [20]. Fatty acids and amino acids can be incorporated into lipid and protein pools, respectively, as well as being catabolised for energy.

Color version available online

MVM

ВМ

arrows.

Membrane transporter abundance in the placenta is regulated in response to nutritional and endocrine signals from the mother [21]. Both maternal diet and body composition are associated with changes in placental transporter activity that reflect nutritional and endocrine signals [22, 23]. Decreased transporter activity is associated with fetal growth restriction that precedes the onset of intra-uterine growth retardation [24]. Changes in the pattern of expression and methylation of the different GLUT transporters in the placenta point to dynamic regulation across gestation [25].

For amino acid transport, and most likely for other nutrients as well, a change in the activity of one transporter cannot be assumed to correspond to an equivalent change in nutrient transport [26]. Computational modelling of these processes may provide an important tool to better understand these processes and to identify the ratellimiting steps, as these will be the best targets for therapeutic interventions [26, 27].

In some diabetic or obese woman, elevated nutrient levels may drive placental transfer down the concentration gradient thereby promoting fetal growth. However, in many obese and diabetic pregnancies, fetal growth is normal raising the question as to why fetal overgrowth occurs in some pregnancies but not in others. One explanation may be that maternal obesity and diabetes in themselves are not sufficient to induce fetal obesity, but that additional exposures are necessary, which in combination lead to excessive placental nutrient transfer and LGA offspring.

The Placental Exposome and Placental Function

For a fetus to become LGA there must be an underlying placental capacity able to support this growth. In absolute terms, fetal growth is greatest in the last trimester and this is when placental nutrient transfer must be sufficient. As placental function develops progressively, it must acquire this capacity much earlier in gestation. For this reason, environmental exposure affecting the placenta in early gestation may interact with those acting in late gestation in order to determine the fetal growth outcome (fig. 2). For instance, an early exposure leading to a large placenta may, when coupled with a later nutrient-rich environment, increase the likelihood of an LGA baby (but may decrease the likelihood of fetal growth restriction in a nutrient-poor late-gestation environment). Alternatively, an early exposure which causes down-regulation of glucose transporters could reduce the likelihood of an LGA infant when exposed to a later glucose-rich environ-

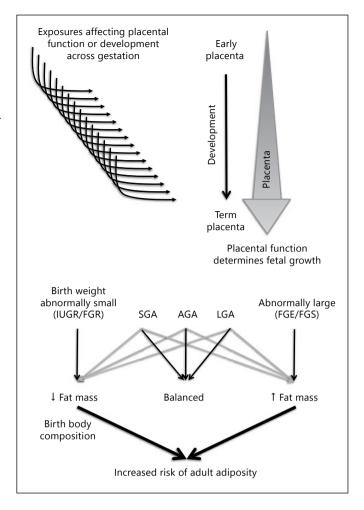


Fig. 2. Placental development and function is determined by the product of its environmental exposures across gestation. This will determine how big the fetus grows and whether its body composition is balanced or imbalanced. Normally grown babies (AGA) as well as some small (SGA) or large (LGA) babies may have a balanced body composition, but most have either inappropriately low or high adiposity, respectively. Abnormally small (IUGR/FGR) infants may have a decreased fat mass, while abnormally large infants with fetal growth excess or fetal growth surplus (FGE/FGS) can have an increased fat mass. Both extremes of fetal growth may lead to adult adiposity.

ment (but might increase the likelihood of fetal growth restriction in a lower glucose environment).

Studies linking maternal body composition to placental function and gene expression suggest a role for longer-term, pre-pregnancy influences of maternal diet on the placenta although it is not clear at which point in gestation these act [22, 23, 28]. Environmental exposures during gametogenesis could influence placental development and thus should be considered as part of the placental exposome [29].

From an evolutionary perspective, co-adaptation suggests that genes will be selected that promote signalling between the mother and offspring to optimise nutrient partitioning in a given environment [30]. In our evolutionary past, under- rather than over-nutrition will have been the predominant selective pressure on the placenta. In this case, signalling between the mother and the fetus via the placenta may be poorly adapted to protect the fetus where there is an overabundance of maternal nutrients such as in maternal obesity or diabetes [31].

A Role for Epigenetics in Pregnancy Adaptation and Placental Function?

Epigenetic mechanisms encompass a range of covalent and other modifications to DNA and associated proteins that together regulate gene activity. The epigenetic profile (epigenome) is dynamic in early life and sensitive to environmental influences, yet the role of epigenetic variation in regulating placental function and pregnancy outcome in humans remains unclear. Epigenetic chance in response to an in utero environmental exposure may manifest as an alteration in placental development potentially as an 'adaptation' of the developing pregnancy to a 'perceived' postnatal environment related to the in utero exposure.

Epigenetic changes are therefore likely to be an important mechanism by which early exposure affects placental function in gestation. There is evidence for increased inter-individual variation in the DNA methylation profile in the third-trimester placenta relative to first and second trimesters, supporting an accumulation of environmentally induced changes in DNA methylation patterns [32]. The mechanisms underlying this variation remain unclear, as does the potential link between altered methylation and adverse pregnancy outcomes. However, parallel changes in methylation and expression in glucose transporters across gestation support a functional role for epigenetic regulation in the placenta [25].

Maternal Determinants of Placental Function

It is currently unclear which aspects of maternal obesity or diabetes predispose to LGA offspring and whether there are common or distinct pathways. Obvious candidates include differences in plasma nutrient and metabolic hormone levels, but other factors such as the systemic inflammatory status could also be important.

The altered energy homeostasis of an obese woman, specifically her higher systemic insulin concentrations and insulin resistance, may also impact on placental function. Even though the placenta is not a typical insulinsensitive tissue with regard to the stimulation of glucose transport, it is equipped with functional insulin receptors and downstream signalling molecules [33]. Leptin is another potentially relevant endocrine signal since plasma leptin is elevated in obese and diabetic women and is synthesized in excess by the placenta [34].

Some diabetic and obese pregnant women have chronic low-grade systemic inflammation, which extends to the placental tissue [35]. The source and nature of the inflammatory stimuli are currently unknown. However, factors originating from the maternal environment are strong candidates to enhance inflammation. Increased systemic lipopolysaccharides in obese pregnant women support a role of innate immune regulation by maternal stimuli in increasing inflammation at the maternal-fetal interface [36]. Lipids, particularly saturated fatty acids, can also trigger immune responses in the placenta increasing in situ inflammation in a manner similar to lipopolysaccharides. The response of the placenta to inflammatory stimuli is demonstrated by enhanced production of cytokines and inflammatory factors released locally and systematically in diabetes and obesity [37].

In addition to dietary factors and hormones, chemicals, such as endocrine disruptors, in the maternal environment can also change the phenotype of offspring by stably altering the epigenome, as has been demonstrated in rodents [38, 39].

Postnatal Outcomes

The long-term consequences of being born with excess fat or LGA are potentially very significant and further research is required to better understand these relationships. Being born large is associated with an increased risk of obesity in childhood although the extent to which this persists into adulthood has not yet been fully defined [40]. If being born LGA is established to be causally associated with adult obesity it might also be expected to predict the risk of metabolic disease and in certain populations LGA may directly be associated with type-2 diabetes. In most populations, the risk of type-2 diabetes decreases with increasing birth weight [41], but in those with a high prevalence of gestational diabetes, the risk of later diabetes is greatest at the highest birth weights [42]. Further research is required to establish whether fetal adiposity contrib-

utes to the risk of adult adiposity and the associated health consequences. This research needs to make use of more detailed analysis of body composition at birth to distinguish those born with a high proportion of body fat from those who are genetically large.

While the focus of this review has been on LGA babies it should not be forgotten that babies who do not reach their growth potential in utero are also prone to develop obesity and other conditions related to the metabolic syndrome in later life which are associated with excess adiposity [43].

If fetal adiposity leads to adult adiposity how might this be mediated? One mechanism may be through effects on the hyperplasia of white, beige or brown adipose tissue [44]. Adipocyte number appears to be a major determinant of adult adiposity and suggests a tight regulation of adipocyte cell number during adult life [45]. Brown fat remains active into adult life [46] and if its abundance was determined in utero this would have implications for energy expenditure and the propensity to develop obesity. While the relationship between early environment and brown fat mass has not been determined it is interesting to note that morbidly obese adults have less brown fat, suggesting a role in the development of obesity [47]. Other mechanisms may involve reprogramming of neural appetite control and there is evidence for reprogramming of the hypothalamus-pituitary-adrenal axis [48]. The relationship between fetal adiposity and early growth should also be investigated as there are strong correlates between early growth and later obesity [49] (fig. 2).

Population Studies and the Power of New Technologies

Population studies are needed to identify the effects of maternal obesity on placental function, fetal growth and postnatal consequences. Developing technologies mean that the level of insight that can be gained from these studies is now substantial. For instance, birth weight is an imprecise marker of fetal growth and while some LGA babies will have a disproportionate increase in adipose tissues others may be proportionately large. If fetal adiposity is key to future health then it is those infants with disproportionately high adiposity rather than those who are proportionately large that need to be identified. The use of neonatal dual-energy X-ray absorptiometry and densitometry to measure body composition allows us to make this distinction and has great potential to enhance the power of these studies.

Furthermore, in order to demonstrate the role of the placenta in mediating maternal environment, it is important that we are able to make better estimates of placental function. It is now possible to make in utero measurements of fetal and placental blood flow and growth [50, 51]. The use of stable isotopes provides an important means to study placental transfer in vivo avoiding the drawbacks of ex vivo systems [52] when coupled with increasingly powerful biochemical and molecular approaches to study plasma and tissues samples [25, 53]. An example of the use of these techniques in a longitudinal study comes from the Southampton Women's Survey, where fetal growth measures have been determined by ultrasound and related to placental gene expression and post-natal bone density [54]. Other studies have related maternal body composition and placental weight to fetal blood flow distribution in the fetus [50, 55]. Increased fetal liver blood flow is associated with fetal macrosomia in the third trimester [51] and greater offspring fat mass measured by dual-energy X-ray absorptiometry, both in the infant at birth and at age 4 years [50]. In the Generation R study, analysis of placental haemodynamics demonstrated associations between maternal lifestyle characteristics and placental resistance indices [56].

This review has focused on the consequences of maternal obesity in humans, however, it should not be forgotten that there is a large body of related work in animal models, which has been reviewed elsewhere [57, 58]. These animal models provide supporting evidence that maternal obesity has long-term effects on the offspring. They also allow study of the underlying processes in ways that are not always possible in humans [59, 60]. As such, animal models provide an important research tool which compliment human studies.

Conclusion

Maternal obesity and diabetes increase the risk of excessive fat accretion in the offspring and are strong risk factors for being born LGA. However, not all such pregnancies lead to fetal overgrowth, and understanding why some fetuses are protected will assist the characterisation of future intervention strategies. Increased nutrient transfer is required to produce an LGA infant, and the factors which determine whether or not placental function mediates this nutrient transfer will determine how the fetus grows. We suggest that to understand this problem will require an appreciation of how exposures across development may affect placental function and that this should be a focus of new and ongoing longitudinal studies.

Acknowledgements

This work was financially supported in part by the Commission of the European Community, specific RTD Programme 'Quality of Life and Management of Living Resources', within the 7th Framework Programme, research grant No. FP7/2007-13 under

grant agreement No. 289346 (Early Nutrition Project). This paper does not necessarily reflect the views of the Commission and in no way anticipates the future policy in this area. K.M.G. is supported by the National Institute for Health Research through the NIHR Southampton Biomedical Research Centre.

References

- 1 Sewell MF, Huston-Presley L, Super DM, Catalano P: Increased neonatal fat mass, not lean body mass, is associated with maternal obesity. Am J Obstet Gynecol 2006;195:1100–1103.
- 2 de Santis MS, Taricco E, Radaelli T, Spada E, Rigano S, Ferrazzi E, Milani S, Cetin I: Growth of fetal lean mass and fetal fat mass in gestational diabetes. Ultrasound Obstet Gynecol 2010;36:328–337.
- 3 Cetin I, Alvino G: Intrauterine growth restriction: implications for placental metabolism and transport. A review. Placenta 2009; 30(suppl A):S77-S82.
- 4 Catalano PM: Obesity and pregnancy the propagation of a viscous cycle? J Clin Endocrinol Metab 2003;88:3505–3506.
- 5 Sebire NJ, Jolly M, Harris JP, Wadsworth J, Joffe M, Beard RW, Regan L, Robinson S: Maternal obesity and pregnancy outcome: a study of 287,213 pregnancies in London. Int J Obes Relat Metab Disord 2001;25:1175–1182.
- 6 Smith J, Cianflone K, Biron S, Hould FS, Lebel S, Marceau S, Lescelleur O, Biertho L, Simard S, Kral JG, Marceau P: Effects of maternal surgical weight loss in mothers on intergenerational transmission of obesity. J Clin Endocrinol Metab 2009;94:4275–4283.
- 7 Catalano PM, Thomas A, Huston-Presley L, Amini SB: Increased fetal adiposity: a very sensitive marker of abnormal in utero development. Am J Obstet Gynecol 2003;189:1698–1704.
- 8 Roos S, Lagerlöf O, Wennergren M, Powell TL, Jansson T: Regulation of amino acid transporters by glucose and growth factors in cultured primary human trophoblast cells is mediated by mTOR signaling. Am J Physiol Cell Physiol 2009;297:C723–C731.
- 9 Constancia M, Hemberger M, Hughes J, Dean W, Ferguson-Smith A, Fundele R, Stewart F, Kelsey G, Fowden A, Sibley C, Reik W: Placental-specific IGF-II is a major modulator of placental and fetal growth. Nature 2002;417: 945–948
- 10 Wild CP: Complementing the genome with an 'exposome': the outstanding challenge of environmental exposure measurement in molecular epidemiology. Cancer Epidemiol Biomarkers Prev 2005;14:1847–1850.
- 11 Benirscke K, Kaufmann P: Pathology of the Human Placenta, ed 3. New York, Springer, 1995
- 12 Moser G, Gauster M, Orendi K, Glasner A, Theuerkauf R, Huppertz B: Endoglandular trophoblast, an alternative route of trophoblast invasion? Analysis with novel confron-

- tation co-culture models. Hum Reprod 2010; 25:1127–1136.
- 13 Diouf I, Botton J, Charles MA, Morel O, Forhan A, Kaminski M, Heude B, The EDEN Study Group: Specific role of maternal weight change in the first trimester of pregnancy on birth size. Matern Child Nutr 2012, E-pub ahead of print.
- 14 Swanson LD, Bewtra C: Increase in normal placental weights related to increase in maternal body mass index. J Matern Fetal Neonatal Med 2008;21:111–113.
- 15 Burton GJ, Watson AL, Hempstock J, Skepper JN, Jauniaux E: Uterine glands provide histiotrophic nutrition for the human fetus during the first trimester of pregnancy. J Clin Endocrinol Metab 2002;87:2954–2959.
- 16 Kaufmann P, Black S, Huppertz B: Endovascular trophoblast invasion: implications for the pathogenesis of intrauterine growth retardation and preeclampsia. Biol Reprod 2003; 69:1–7.
- 17 Burton GJ, Woods AW, Jauniaux E, Kingdom JC: Rheological and physiological consequences of conversion of the maternal spiral arteries for uteroplacental blood flow during human pregnancy. Placenta 2009;30:473–482.
- 18 Cleal JK, Glazier JD, Ntani G, Crozier SR, Day PE, Harvey NC, Robinson SM, Cooper C, Godfrey KM, Hanson MA, Lewis RM: Facilitated transporters mediate net efflux of amino acids to the fetus across the basal membrane of the placental syncytiotrophoblast. J Physiol 2011;589:987–997.
- 19 Zamudio S, Torricos T, Fik E, Oyala M, Echalar L, Pullockaran J, Tutino E, Martin B, Belliappa S, Balanza E, Illsley NP: Hypoglycemia and the origin of hypoxia-induced reduction in human fetal growth. PloS One 2010;5:e8551.
- 20 Osmond DT, Nolan CJ, King RG, Brennecke SP, Gude NM: Effects of gestational diabetes on human placental glucose uptake, transfer, and utilisation. Diabetologia 2000;43:576–582.
- 21 Roos S, Jansson N, Palmberg I, Saljo K, Powell TL, Jansson T: Mammalian target of rapamycin in the human placenta regulates leucine transport and is down-regulated in restricted fetal growth. J Physiol 2007;582:449–459.
- 22 Lewis RM, Greenwood SL, Cleal JK, Crozier SR, Verrall L, Inskip HM, Cameron IT, Cooper C, Sibley CP, Hanson MA, Godfrey KM: Maternal muscle mass may influence system A activity in human placenta. Placenta 2010;31:418–422.
- 23 Radaelli T, Varastehpour A, Catalano P, Hauguel-de Mouzon S: Gestational diabetes in-

- duces placental genes for chronic stress and inflammatory pathways. Diabetes 2003;52: 2951–2958
- 24 Jansson N, Pettersson J, Haafiz A, Ericsson A, Palmberg I, Tranberg M, Ganapathy V, Powell TL, Jansson T: Down-regulation of placental transport of amino acids precedes the development of intrauterine growth restriction in rats fed a low protein diet. J Physiol 2006; 576:935–946.
- 25 Novakovic B, Gordon L, Robinson WP, Desoye G, Saffery R: Glucose as a fetal nutrient: dynamic regulation of several glucose transporter genes by DNA methylation in the human placenta across gestation. J Nutr Biochem 2013;24:282–288.
- 26 Lewis RM, Brooks S, Crocker IP, Glazier J, Hanson M, Johnstone ED, Panitchob N, Please CP, Sibley CP, Widdows KL, Sengers BG: Modelling placental amino acid transfer – from transporters to placental function. Placenta 2013;34:S46–S51.
- 27 Sengers BG, Please CP, Lewis RM: Computational modelling of amino acid transfer interactions in the placenta. Exp Physiol 2010;95: 829–840.
- 28 O'Tierney PF, Lewis RM, McWeeney SK, Hanson MA, Inskip HM, Morgan TK, Barker DJ, Bagby G, Cooper C, Godfrey KM, Thornburg KL: Immune response gene profiles in the term placenta depend upon maternal muscle mass. Reprod Sci 2012;19:1041–1056.
- 29 Fleming TP, Kwong WY, Porter R, Ursell E, Fesenko I, Wilkins A, Miller DJ, Watkins AJ, Eckert JJ: The embryo and its future. Biol Reprod 2004;71:1046–1054.
- 30 Agrawal AF, Brodie ED 3rd, Brown J: Parentoffspring coadaptation and the dual genetic control of maternal care. Science 2001;292: 1710–1712.
- 31 Lewis RM, Cleal JK, Hanson MA: Review: placenta, evolution and lifelong health. Placenta 2012;33:S28–S32.
- 32 Novakovic B, Yuen RK, Gordon L, Penaherrera MS, Sharkey A, Moffett A, Craig JM, Robinson WP, Saffery R: Evidence for widespread changes in promoter methylation profile in human placenta in response to increasing gestational age and environmental/stochastic factors. BMC Genomics 2011;12: 529.
- 33 Desoye G, Hauguel-de Mouzon S: The human placenta in gestational diabetes mellitus. The insulin and cytokine network. Diabetes Care 2007;30(suppl 2):S120–S126.

- 34 Lepercq J, Cauzac M, Lahlou N, Timsit J, Girard J, Auwerx J, Hauguel-de Mouzon S: Overexpression of placental leptin in diabetic pregnancy: a critical role for insulin. Diabetes 1998;47:847–850.
- 35 Catalano PM, Presley L, Minium J, Hauguelde Mouzon S: Fetuses of obese mothers develop insulin resistance in utero. Diabetes Care 2009;32:1076–1080.
- 36 Basu S, Haghiac M, Surace P, Challier JC, Guerre-Millo M, Singh K, Waters T, Minium J, Presley L, Catalano PM, Hauguel-de Mouzon S: Pregravid obesity associates with increased maternal endotoxemia and metabolic inflammation. Obesity 2011;19:476–482.
- 37 Challier JC, Basu S, Bintein T, Minium J, Hotmire K, Catalano PM, Hauguel-de Mouzon S: Obesity in pregnancy stimulates macrophage accumulation and inflammation in the placenta. Placenta 2008;29:274–281.
- 38 Ryan KK, Haller AM, Sorrell JE, Woods SC, Jandacek RJ, Seeley RJ: Perinatal exposure to bisphenol-a and the development of metabolic syndrome in CD-1 mice. Endocrinology 2010;151:2603–2612.
- 39 Dolinoy DC, Huang D, Jirtle RL: Maternal nutrient supplementation counteracts bisphenol a-induced DNA hypomethylation in early development. Proc Natl Acad Sci USA 2007;104:13056–13061.
- 40 Yu ZB, Han SP, Zhu GZ, Zhu C, Wang XJ, Cao XG, Guo XR: Birth weight and subsequent risk of obesity. Obes Rev 2011;12:525–542.
- 41 Whincup PH, Kaye SJ, Owen CG, Huxley R, Cook DG, Anazawa S, Barrett-Connor E, Bhargava SK, Birgisdottir BE, Carlsson S, de Rooij SR, Dyck RF, Eriksson JG, Falkner B, Fall C, Forsen T, Grill V, Gudnason V, Hulman S, Hypponen E, Jeffreys M, Lawlor DA, Leon DA, Minami J, Mishra G, Osmond C, Power C, Rich-Edwards JW, Roseboom TJ, Sachdev HS, Syddall H, Thorsdottir I, Vanhala M, Wadsworth M, Yarbrough DE: Birth weight and risk of type 2 diabetes: a systematic review. JAMA 2008;300:2886–2897.
- 42 Pettitt DJ, Nelson RG, Saad MF, Bennett PH, Knowler WC: Diabetes and obesity in the offspring of Pima Indian women with diabetes during pregnancy. Diabetes Care 1993;16: 310–314.

- 43 Symonds ME, Sebert SP, Hyatt MA, Budge H: Nutritional programming of the metabolic syndrome. Nat Rev Endocrinol 2009;5:604– 610.
- 44 Symonds ME, Pope M, Sharkey D, Budge H: Adipose tissue and fetal programming. Diabetologia 2012;55:1597–1606.
- 45 Spalding KL, Arner E, Westermark PO, Bernard S, Buchholz BA, Bergmann O, Blomqvist L, Hoffstedt J, Naslund E, Britton T, Concha H, Hassan M, Ryden M, Frisen J, Arner P: Dynamics of fat cell turnover in humans. Nature 2008:453:783–787.
- 46 van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, Drossaerts JM, Kemerink GJ, Bouvy ND, Schrauwen P, Teule GJ: Coldactivated brown adipose tissue in healthy men. N Engl J Med 2009;360:1500–1508.
- 47 Vijgen GH, Bouvy ND, Teule GJ, Brans B, Schrauwen P, van Marken Lichtenbelt WD: Brown adipose tissue in morbidly obese subjects. PloS One 2011;6:e17247.
- 48 Reynolds RM: Corticosteroid-mediated programming and the pathogenesis of obesity and diabetes. J Steroid Biochem Mol Biol 2010;122:3–9.
- 49 Monteiro PO, Victora CG: Rapid growth in infancy and childhood and obesity in later life – a systematic review. Obes Rev 2005;6: 143–154.
- 50 Godfrey KM, Haugen G, Kiserud T, Inskip HM, Cooper C, Harvey NC, Crozier SR, Robinson SM, Davies L, Southampton Women's Survey Study Group, Hanson MA: Fetal liver blood flow distribution: role in human developmental strategy to prioritize fat deposition versus brain development. PloS One 2012; 7:e41759.
- 51 Kessler J, Rasmussen S, Godfrey K, Hanson M, Kiserud T: Venous liver blood flow and regulation of human fetal growth: evidence from macrosomic fetuses. Am J Obstet Gynecol 2011;204:429. e421–e427.
- 52 Gil-Sanchez A, Larque E, Demmelmair H, Acien MI, Faber FL, Parrilla JJ, Koletzko B: Maternal-fetal in vivo transfer of [13C]docosahexaenoic and other fatty acids across the human placenta 12 h after maternal oral intake. Am J Clin Nutr 2010;92:115–122.

- 53 Hellmuth C, Weber M, Koletzko B, Peissner W: Nonesterified fatty acid determination for functional lipidomics: comprehensive ultrahigh performance liquid chromatography-tandem mass spectrometry quantitation, qualification, and parameter prediction. Anal Chem 2012;84:1483–1490.
- 54 Lewis RM, Cleal JK, Ntani G, Crozier SR, Mahon PA, Robinson SM, Harvey NC, Cooper C, Inskip HM, Godfrey KM, Hanson MA, Southampton Women's Survey Study Group, John RM: Relationship between placental expression of the imprinted PHLDA2 gene, intrauterine skeletal growth and childhood bone mass. Bone 2012;50:337– 342.
- 55 Haugen G, Hanson M, Kiserud T, Crozier S, Inskip H, Godfrey KM: Fetal liver-sparing cardiovascular adaptations linked to mother's slimness and diet. Circ Res 2005;96:12–14.
- 56 Gaillard R, Arends LR, Steegers EA, Hofman A, Jaddoe VW: Second- and third-trimester placental hemodynamics and the risks of pregnancy complications: the Generation R Study. Am J Epidemiol 2013;177:743–754.
- 57 Nathanielsz PW, Poston L, Taylor PD: In utero exposure to maternal obesity and diabetes: animal models that identify and characterize implications for future health. Clin Perinatol 2007;34:515–526, v.
- 58 Seki Y, Williams L, Vuguin PM, Charron MJ: Minireview: epigenetic programming of diabetes and obesity: animal models. Endocrinology 2012;153:1031–1038.
- 59 Samuelsson AM, Clark J, Rudyk O, Shattock MJ, Bae SE, South T, Pombo J, Redington K, Uppal E, Coen CW, Poston L, Taylor PD: Experimental hyperleptinemia in neonatal rats leads to selective leptin responsiveness, hypertension, and altered myocardial function. Hypertension 2013;62:627–633.
- 60 Nicholas LM, Rattanatray L, Maclaughlin SM, Ozanne SE, Kleemann DO, Walker SK, Morrison JL, Zhang S, Muhlhausler BS, Martin-Gronert MS, McMillen IC: Differential effects of maternal obesity and weight loss in the periconceptional period on the epigenetic regulation of hepatic insulin-signaling pathways in the offspring. FASEB J 2013;27:3786– 3796.