

RESEARCH

Open Access

The cytohesin paralog Sec7 of *Dictyostelium discoideum* is required for phagocytosis and cell motility

Rolf Müller¹, Claudia Herr¹, Salil K Sukumaran¹, Napoleon Nosa Omosigho¹, Markus Plomann², Tanja Y Riyahi^{1*}, Maria Stumpf¹, Karthic Swaminathan¹, Marios Tsangarides¹, Kyriacos Yiannakou¹, Rosemarie Blau-Wasser¹, Christoph Gallinger³, Michael Schleicher³, Waldemar Kolanus⁴ and Angelika A Noegel^{1*}

Abstract

Background: *Dictyostelium* harbors several paralogous Sec7 genes that encode members of three subfamilies of the Sec7 superfamily of guanine nucleotide exchange factors. One of them is the cytohesin family represented by three members in *D. discoideum*, SecG, Sec7 and a further protein distinguished by several transmembrane domains. Cytohesins are characterized by a Sec7-PH tandem domain and have roles in cell adhesion and migration.

Results: We study here Sec7. In vitro its PH domain bound preferentially to phosphatidylinositol 3,4-bisphosphate (PI(3,4)P₂), phosphatidylinositol 4,5-bisphosphate (PI(4,5)P₂) and phosphatidylinositol 3,4,5-trisphosphate (PI(3,4,5)P₃). When following the distribution of GFP-Sec7 in vivo we observed the protein in the cytosol and at the plasma membrane. Strikingly, when cells formed pseudopods, macropinosomes or phagosomes, GFP-Sec7 was conspicuously absent from areas of the plasma membrane which were involved in these processes. Mutant cells lacking Sec7 exhibited an impaired phagocytosis and showed significantly reduced speed and less persistence during migration. Cellular properties associated with mammalian cytohesins like cell-cell and cell-substratum adhesion were not altered. Proteins with roles in membrane trafficking and signal transduction have been identified as putative interaction partners consistent with the data obtained from mutant analysis.

Conclusions: Sec7 is a cytosolic component and is associated with the plasma membrane in a pattern distinctly different from the accumulation of PI(3,4,5)P₃. Mutant analysis reveals that loss of the protein affects cellular processes that involve membrane flow and the actin cytoskeleton.

Keywords: ARFGEF, Cell adhesion, Cell migration, Phagocytosis, Phosphoinositide binding

Background

ADP ribosylation factor (ARF) GTPases have roles in vesicular transport, in the regulation of actin cytoskeleton dynamics, cell adhesion, cell migration and in signal transduction processes. They depend on specific nucleotide exchange factors, the ARFGEFs, which through their conserved Sec7 domain (Sec7d) catalyze the GDP to GTP exchange. Six subfamilies of ARFGEFs exist in eukaryotes, the large ARFGEFs of the GBF and BIG family, and

the small ARFGEFs of the cytohesin, EFA6, BRAG and FBX family. For all ARFGEFs localization to membranes is important for their functions in ARF activation. BIG1 and BIG2 localize to the TGN and endosomes, GBF localizes to the Golgi, and cytohesins are found at the cell periphery where they function in plasma membrane endosomal membrane trafficking and in signal transduction pathways [1].

Cytohesins are composed of a Sec7d domain and a pleckstrin homology (PH) domain of the cytohesin type followed by a polybasic stretch which together with the PH domain is necessary for plasma membrane association and biological function [2]. The PH domain of the cytohesins specifically binds to phosphoinositides, whose

* Correspondence: tanja.riyahi@uni-koeln.de; noegel@uni-koeln.de
¹Institute of Biochemistry I, Medical Faculty, Center for Molecular Medicine Cologne (CMMC) and Cologne Excellence Cluster on Cellular Stress Responses in Aging-Associated Diseases (CECAD), University of Cologne, 50931 Köln, Germany
Full list of author information is available at the end of the article

levels are influenced by PI3-kinases in response to upstream signals. Upon recruitment to the plasma membrane cytohesins activate ARFs, which influence the cortical actin cytoskeleton, endocytic processes and phagocytosis. Cytohesins also regulate signaling pathways by functioning in scaffolding complexes. Such a complex was reported for cytohesin-2 which through its N-terminal sequences could assemble a protein complex that contained the RacGEF DOCK180 and promoted Rac activation and cell migration [3]. Furthermore, in dendritic cells cytohesin-1 regulated migration in vivo by functioning upstream of RhoA activation [4].

The lower eukaryote *D. discoideum* harbors six ARFGEFs belonging to three families, the GBF and BIG family with one and two representatives each, and three members of the cytohesin family harboring the characteristic Sec7-PH tandem [5]. The finding of this class of proteins in a lower eukaryote was surprising as cytohesins had been reported only for metazoans [6]. Instead of the typical coiled coil domain at the N-terminus, the *Dictyostelium* cytohesins have variable domains. SecG (DDB_G0287459) has several ankyrin repeats and DDB0233591 (DDB_G0279241) harbors four predicted transmembrane domains whereas the N-terminus of DDB0233617 (DDB_G0272486) which we designate Sec7 has no putative conserved domains [5]. Instead, homopolymer tracts of asparagine and threonine are present. Such homopolymer tracts are quite frequent in *D. discoideum* proteins [7,8].

In previous work we had analyzed the function of SecG and found that mutant cells lacking SecG had reduced cell-substratum adhesion whereas cell cell adhesion was not affected. In cell migration analysis speed was significantly reduced, persistence and directionality of migration were unaltered. Here we analyze the Sec7 protein and characterize Sec7 deficient cells. We find that Sec7 is a cytosolic component and is associated with the plasma membrane in a pattern distinctly different from the accumulation of PI(3,4,5)P₃. We compare these data with the one reported for known PH sensors and ArfA, the single ARF GTPase of *D. discoideum*.

Results

Sec7 associates with the plasma membrane

The *D. discoideum* cytohesin Sec7 is a 931 amino acids protein of 103.797 kDa and has a pI of 7.97. The rather unstructured N terminal region encompasses ~250 amino acids, the Sec7 domain extends from amino acids 252 to 441 and is separated from the basic PH domain (residues 454–577) through a short linker (Figure 1A). The structure of Sec7-PH has been obtained by homology modeling using 2r0da (Crystal Structure of Autoinhibited Form of Grp1 Arf GTPase Exchange Factor (Cytohesin-3)) as template (Figure 1B). The Sec7

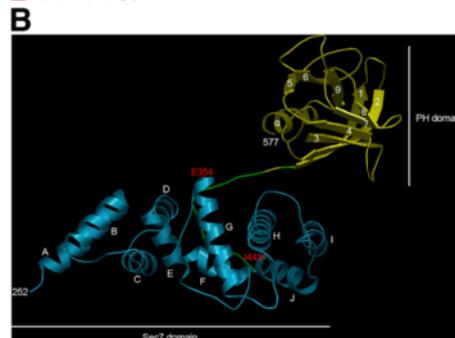
domain of the *D. discoideum* protein contains 10 alpha-helices, A to J. The helix J (position 428-438) is crucial for ARF binding and exchange. The Sec7 domain also includes the key residue glutamate (E) (position 354), which is essential for GDP to GTP exchange, and an isoleucine (I) at position 443 that is part of a hydrophobic cluster involved in ARF interaction (Figure 1B, indicated in red). Thus it corresponds to a typical Sec7 domain [9,10].

The PH domains of cytohesins bind polyphosphoinositides and have different affinities and specificities for PI(3,4)P₂, PI(4,5)P₂ and PI(3,4,5)P₃ depending on their structure [2,13,14]. The signaling lipids PI(3,4)P₂ and PI(3,4,5)P₃ are rare components of the plasma membrane and are produced in response to a stimulus which then recruits the PH domain proteins to the membrane. PH domains have a conserved core fold consisting of a seven strand β -barrel followed by an α -helix [11,15]. The PH domain of Sec7 has these structural elements too, however, when we modeled the Sec7 sequence to the crystal structure of the autoinhibited form of Grp1 Arf GTPase Exchange Factor we identified a nine strand β -barrel (Figure 1B). The signature motif for 3-phosphoinositide binding K X_m KxR X_n Y [11] is modified to I X₁₀ SxK X₁₀ F (m = 5-10; n = 6-13). Changes in these positions are also present in PH domains of other proteins (Figure 1C) [16]. A recently described glutamate in the PH domain of cytohesin-3 (GRP1), a sentry glutamate, appears to be essential for specific PI(3,4,5)P₃ binding by the cytohesins as a charge reversal by mutation to lysine (E345K) enhanced the affinity for PI(4,5)P₂ and yielded constitutive plasma membrane binding [17]. In Sec7 the glutamate is replaced by a glutamine which is considered a neutral residue (Figure 1A,B). A phylogenetic analysis of the Sec7 PH domain placed the *D. discoideum* protein close to GRP1 proteins of several species including the human protein (Additional file 1: Figure S1).

We tested the ability of the PH domain of Sec7 to bind to different phosphoinositides in vitro and used a GST fusion protein encompassing residues 455-931 (GST-PH) in dot blot overlay assays. In this assay GST-PH bound preferentially to PI(3,4,5)P₃ and showed decreased binding to PI(4,5)P₂. GST alone did not show binding (Figure 2A). Although dot blot overlay assays are convenient assays, they need to be supported by different methods as apparent specificities can be distorted and as they do not allow reliable quantification [18]. We therefore carried out liposome binding assays in which we used liposomes containing 65% phosphatidylcholine, 20% phosphatidylethanolamine, 5% phosphatidylserine and 10% individual phosphoinositides and examined the sedimentation of GST-PH with the vesicles. The protein did not show significant binding to the monophosphorylated inositides PI

A
 MSDSQTTTHDNNKEVEFNEPITIVISNNGADKSLPPI S QKQFLN NNNNNNDEDSNTNTN
 TTTT PRSDTHLPT INNNNNNNNGITDEEETSPIGSYPT SWMELKSKMSQVDNSDLKRG
 SILIEHLSEYNKIASSQVLHEEIIQPTNNNNNNNNNNNNNNNNNNNNNNNNNNNNNN
 NNNKQOMTINININTNTI PLSTRI P PPP PVTKTTININNNNT PAATTTTTTTTTPTPTPI
 PTPPTPTNITT TSLTQANLDSIILSGIQLFNEKPKKGI EYFFQHKLEKT PPSVSEFLHE
 CPLLDKKSIGDYLGDAE PFCILVLESLSIRFNFKDLD FMSLRQLLYS FRLPG EAQKI DR
 VVQRFANQYHKDNKEGGFIEDPDTVYTLAFAL ILLNTDLHNANVKPTMTKAKFVKLSKI
 NSGKDLPSFLEDMFDR I L VDEIKLNPSSSTLFF YAVKKGWLNIRVKGKVTDKWSRKWCVL
 SDGTYLFFRKPTDPSVRYLRPDTVITSKKEIKGRKNC EMLNHSSPITLES LIKSTQKN
 KSQMFNPQSLQLQALFGKEKEKVVYFWDSAVEK FYEQQPQLDIQDILDTSVSSIDT
 LSETSSSTNYFDDEYLSPTSSSVSTNSSTNSTTS PTLFDQQSFDQNKKGLFRSKNGQIV
 ECEGMYKNGTQLSRPPNKNTVHYNPNFNTIHP I PQSGSSTNSYNNNNNNNNNNNN
 NNNNNNTNSNANTTTTTTTTTTTTTTNTGNSNNTSAVNHSLTQPRPNYFKTVSSSLIRS
 SSI STFSTPPSSFNNNNNNNNNNNNNNNV PSSLTSSNTSVA SNNQQHNHS PAGPANSSS
 SSS T PVLSSSTSSSSSSSSSRNT PVSHEQNNKSEYDKNYEKNLNAFMKLQEKMIKA
 SKNTNTCILNTDTKREMDSWIRLLILHQFSK*

■ Sec 7 domain
 ■ linker region
 ■ Pleckstrin homology domain
 * E(354) and I(443) highly conserved



C

PH_d4DB_G0272486	1	-----LFPYA VREGWLRV K--GK--VTD KSKK--CML SD-----G TAVFERKPTD	43
PH_haGrp1	1	-----HTFFNP DREGWLRG G--G-RVE TNRGSRHFD ID-----N CAGDFEYTD	42
PH_haDAPP1	1	-----TAPSLG TREGWLRG G--G-LVK TNRGSRHFD HR-----N RQDFEDQMS	42
PH_haPKB1	1	-----DVAI VREGWLRV G--G-RYK TNRGSRHFD KM-----DOT FFGKRPQD	42
PH_haBtk	1	-----AV ILTITLSS GQK-EKESPL HNSG--L EY-----N CAGDFEFD	43
PH_haGAB1	1	-----MSGGEV VREGWLRV PEKRL-KRY ADRG--L EY-----N RSGRLTDDP	53
PH_haGAP1	1	-----DGDGAVP ARALFPSSAI VREGWLRK EEPAGLATRF ADRG--L EY-----E DGSFESPEV	60
PH_haDYN	1	-----EILV IREGWLRV I--GI--MKG GGRVY--MPL TA-----E NGSWYKDDK	42
PH_haSBF1	1	-----GR SYEGWLRG G--G--A-FMK PNRGSRHFD DK-----TEK CAGDFEYTD	40
PH_haARNO	1	-----HTFFNP DREGWLRG G--G-RVK TNRGSRHFD TD-----N CAGDFEYTD	43
PH_haPDK1	1	LEIKQAGNHP NMQVVENHLE EREGWLRK G--L-----FARRGQLLP TE-----G PHSQVDPVH	54
PH_haCyto	1	-----HTFFNP DREGWLRG G--G-RVK TNRGSRHFD ID-----N CAGDFEYTD	43
PH_d4Sec9	1	-----QANQNV EREGWLRK G--G-RIK TNRGSRHFD TA-----N CAGDFEYTD	42
PH_haPKBg	1	-----MSDVTI VREGWLRV G--G-YIK NDRG--L EY-----DGS FFGKRPQD	44
PH_haP1Cd	1	-----LQAL LREGWLRV G--G--KSS SRRRERFD QE-----D CKTIHRSRK	39
Consensus		-----LFPYA VREGWLRV K--GK--VTD KSKK--CML SD-----G TAVFERKPTD	

PH_d4DB_G0272486	44	P-----SRYR LEPDTVITSE KRI--KG-----RKNC LNHSSPPI TLESLIKSTQ KNSGMPFPQ	98
PH_haGrp1	43	K-----RNOY DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	93
PH_haDAPP1	43	P-----RRIE DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	76
PH_haPKB1	44	VDRGAEANN FVAQCQLMK TER--PR-----PH-T IIR-----C-----LQ	77
PH_haBtk	44	RRRRSKKS DVEKTCVY TVVPEKN-----PPP RQFPROGES SEMQISII-ERFFYFPVY	103
PH_haGAB1	54	K-----KLE DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	88
PH_haGAP1	61	Q-----MCS FVSEIYAVE RVD-----EGR DDFPHVQ-----VU-----TQDT	96
PH_haDYN	43	K-----EKYV LVDNLELD VEKGFMS-----SKH GAF-----NT-EGRNRY	81
PH_haSBF1	41	T-----ECKGV DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	77
PH_haARNO	44	K-----RNOY DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	94
PH_haPDK1	55	K-----VLKGE FVNSQLRPE AK-----NFKT F-----VH	80
PH_haCyto	44	K-----RNOY DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	94
PH_d4Sec9	43	H-----RNOY DLENLSIRE VED-PRK-----PH-C PELVFNFSH--KQVVKACK TEA-DGRVVE	98
PH_haPKBg	45	VDL-FYANN FVAQCQLMK TER--PR-----PH-T IIR-----C-----LQ	78
PH_haP1Cd	40	V-MESFESQI FSEDIQVR MSHRTEGLEK FARDIPERC SLV-----FK	84
Consensus		P-----SRYR LEPDTVITSE KRI--KG-----RKNC LNHSSPPI TLESLIKSTQ KNSGMPFPQ	

PH_d4DB_G0272486	99	SLLQQLAEPG KEKKEKVTI DSA-----V EEPYEQD-----	130
PH_haGrp1	94	GHRVYTRER P-PEKKEEY HKAHAISR DFFY-----	126
PH_haDAPP1	76	-----TFVICA K-EGVADH EKILNKLQ I-----	102
PH_haPKB1	78	WTVIERTFH VETPEKREY TAAQTAVDQ LKKQE-----	112
PH_haBtk	104	YDEGPLVYF P-TELRKREY HOLENVIRY NSDLVQYKH CFWIDGQQLC CSQTAKHANG CQILE	167
PH_haGAB1	89	TIDRFYLYA D-SEKQNHV VRC-CDICGF NPTREDPVK P-----	128
PH_haGAP1	97	GALHTTYLQC K-NVPLNMG ESALERKASAP MPKLAACHP GAFESARWT-----	144
PH_haDYN	82	EDYRGLRAC P-EGVVDV EAFERAVV-----FPERV-----	114
PH_haSBF1	78	TRKVTYVPC Q-DVTEAQQ VDSKSCLS GP-----	108
PH_haARNO	95	GHNMYTRISA P-EGVVDV EAFERAVV-----FPERV-----	127
PH_haPDK1	81	TFNRTYIMD P--SONAHV CRKDEWWRQ R--VQSH-----	113
PH_haCyto	95	GHRVYTRER P-PEKKEEY HKAHAISR DFFY-----	127
PH_d4Sec9	89	ANHAAYFAR A-NGLMDS VDSKSCLS NPMP-----	121
PH_haPKBg	79	WTVIERTFH VETPEKREY TAAQTAVDQ LKKQE-----	112
PH_haP1Cd	85	DQRNTLDIHA P-EPADAQK HOGELRIH SSSMDQRK-----	122
Consensus		SLLQQLAEPG KEKKEKVTI DSA-----V EEPYEQD-----	

Figure 1 (See legend on next page.)

(See figure on previous page.)

Figure 1 Domain structure of the Sec7 protein. (A) The amino acid sequence of Sec7 of *D. discoideum* is shown (DDB0233617). The Sec7 domain is highlighted in blue, the linker region in green and the Pleckstrin homology (PH) domain in yellow. The highly conserved amino acids E(354) and I(443) are marked with red. (B) The DDB0233617 Sec7-PH-domain modeled to 2r0dA (Crystal Structure of Autoinhibited Form of Grp1 Arf GTPase Exchange Factor). The Sec7 domain is depicted in blue, the linker region in green and the PH domain in yellow. The highly conserved amino acids E(354) and I(443) are shown in red. Modeling was with SwissModel, visualization with OpenAstexViewer. (C) CLUSTALX alignments of PH domains from *Homo sapiens* and *D. discoideum* according to Lietzke et al. and Ferguson et al. [11,12]. The asterisks in the Consensus line indicate the signature motif as suggested by Lietzke et al. and Ferguson et al. [11,12]. ddDDB_G0272486 corresponds to Sec7.

(3)P, PI(4)P, PI(5)P and to the bisphosphorylated PI(3,5)P₂, whereas it pelleted with PI(3,4)P₂, PI(4,5)P₂ and PI(3,4,5)P₂. To quantitatively study which PIs are preferred, we scanned the band intensities of the Coomassie-stained gels and plotted the sedimented pellet fractions in a diagram. We found that in this assay the PH domain interacted equally well with PI(3,4)P₂ and PI(4,5)P₂ and exhibited less binding to PI(3,4,5)P₂. The GST control showed no binding (Figure 2B,C,D).

A polybasic domain which follows the PH domains in all cytohesins is not present in Sec7. However, the C-terminus of Sec7 encompassing the PH domain and the remaining stretch of amino acids is highly basic with a pI of 9.58 which is presumably due to two poly

asparagine stretches and an abundance of lysine residues near the C-terminus (10 lysine residues out of 56 residues).

To address the localization of Sec7, we expressed the protein as a GFP fusion protein (GFP-Sec7) in a Sec7 deficient strain (see below). In fixed cells we found GFP-Sec7 diffusely distributed throughout the cytosol and present at the plasma membrane where it colocalized with annexin 7 [19] (Figure 3A). In live cell microscopy we observed GFP-Sec7 in association with the plasma membrane and in the cytosol. When cells extended pseudopods, GFP-Sec7 disappeared from the plasma membrane in the region of the newly forming protrusions (Figure 3B; 140, 160 sec). During macropinocytic

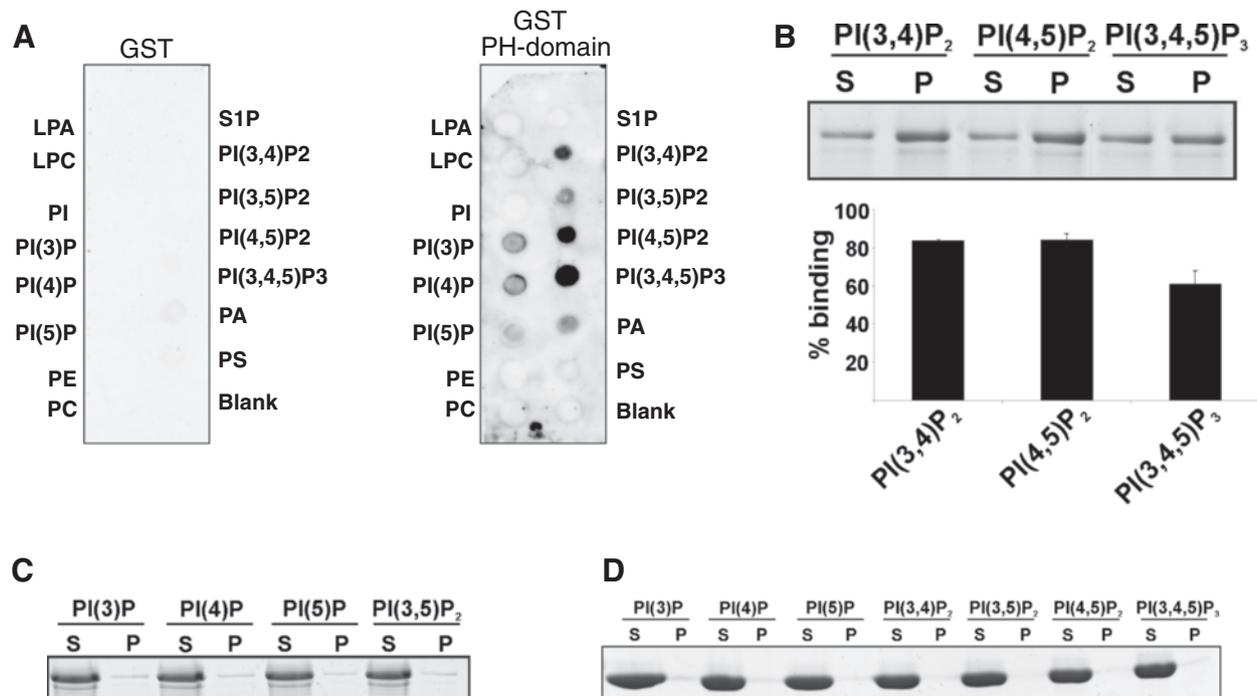


Figure 2 The PH domain binds to phosphoinositides. (A) PIP-Strip-membranes were incubated over night with GST-PH domain (1 µg/ml) and with GST (1 µg/ml) for control. Binding was detected by incubation with polyclonal GST-antibodies. (B) Binding of Sec7-PH domain (residues 454-577) to PIs in a liposome binding assay. Upper panel, 5 µg of the GST-PH fusion protein were incubated with liposomes containing 10% (wt/wt) of the indicated PIs. Liposomes were collected by centrifugation, and bound proteins resolved by SDS-PAGE and detected by Coomassie Blue staining (S = supernatant; P = pellet). Lower panel, for quantification, the bands were scanned and the ratio of the pellet vs. supernatant intensities of each PI fraction after background subtraction calculated. Error bars, SD of liposome binding from two independent experiments. (C) Binding of the Sec7 PH domain to PI(3)P, PI(4)P, PI(5)P and PI(3,5)P₂ was negligible. (D) Liposome binding assay using GST for control.

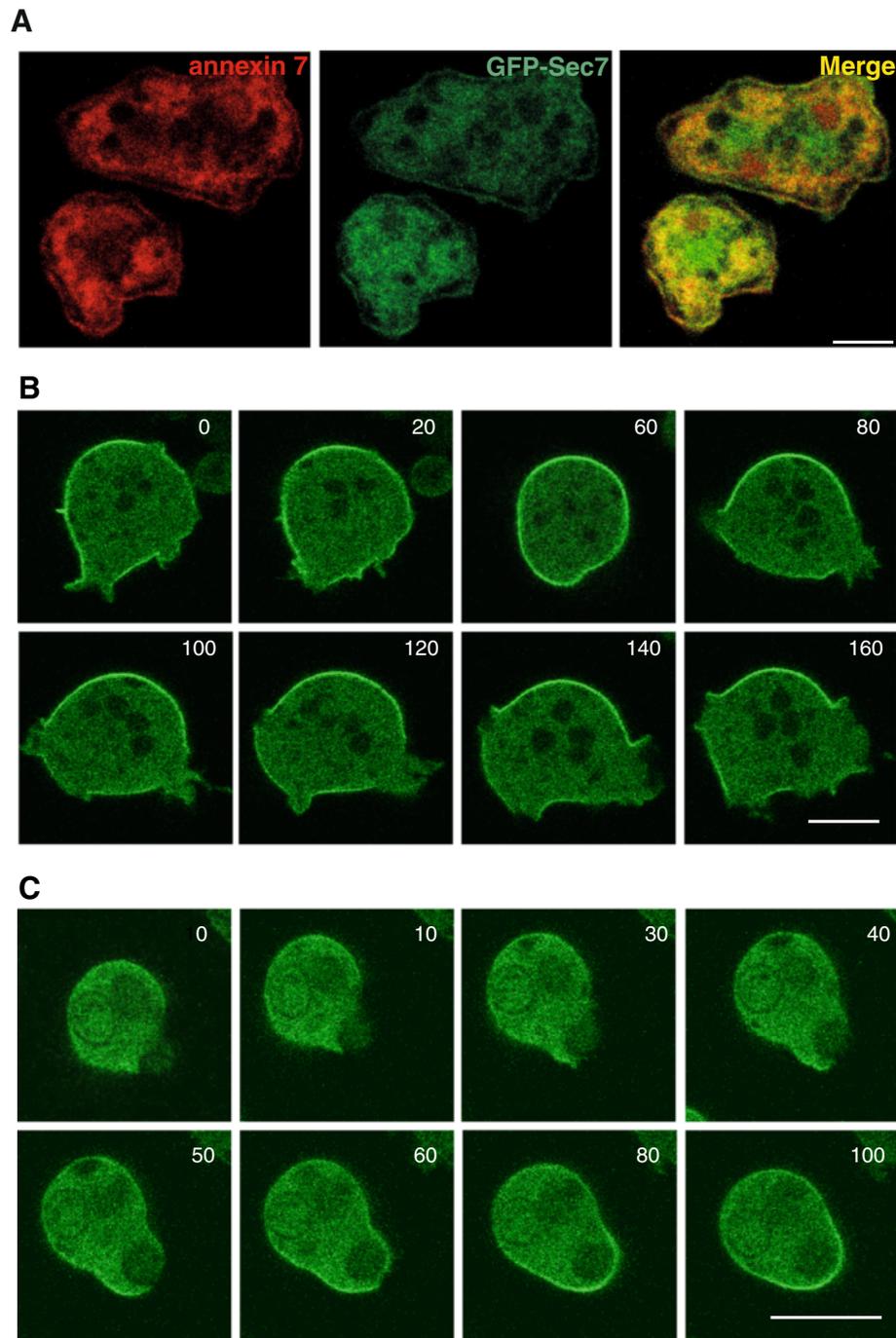


Figure 3 Subcellular localization of GFP-Sec7. To determine the localization of Sec7 we expressed a GFP-tagged Sec7 in Sec7 deficient cells. **(A)** GFP-Sec7 in methanol fixed cells. The cells were stained for annexin 7 with mAb 185-338-1 to reveal the plasma membrane. Annexin 7 is present in the cytosol, in the nucleus and at the plasma membrane. Bar, 5 μ m. **(B)** Live cell analysis of GFP-Sec7 expressing cells. Representative images of a series are shown. Bar, 10 μ m. **(C)** Live cell analysis of a GFP-Sec7 expressing cells during phagosome formation. Yeast particles were used to study phagocytosis. The time is given in seconds. Bar, 10 μ m.

cup formation, which allows cells to take up fluid [20], GFP-Sec7 was also excluded from the involved areas of the plasma membrane and reappeared upon completion of the process (Figure 3B, 80, 100 sec). Similarly, during

phagocytosis of yeast particles GFP-Sec7 was not enriched at the forming phagocytic cup (Figure 3C; 10, 30, 40 sec). Once phagosome formation was completed and the yeast particle engulfed GFP-Sec7 reappeared on the plasma

membrane (Figure 3C; 60, 80 sec). This behavior is unlike the one described for several PH-domains that recognize various phosphoinositides. It rather resembled the one of PTEN which is excluded from leading edges and whose distribution is complementary to the one of PI(3,4,5)P₃ [21]. The PH domain of PLC δ , which binds to PI(4,5)P₂ and labels the plasmamembrane was similarly depleted from the active plasmamembrane [22]. Plasmamembrane regions of the cell involved in pseudopod extension, macropinosome and phagosome formation are highly dynamic and require membrane insertion as well as a dynamic underlying actin cytoskeleton which are processes triggered by transient PI(3,4,5)P₃ formation. This analysis supports the proposed preference of Sec7 for PI(4,5)P₂.

In cell fractionation experiments the protein was predominantly found in the cytosolic fraction which is in agreement with data obtained for cytohesins. A signal in the 100,000 \times g pellet was only obtained when we loaded the material from 2 \times 10⁷ cells/ml whereas for whole cell lysate and 10,000 \times g pellet and supernatant, respectively, the proteins represent the material from 2 \times 10⁵ cells/ml (Additional file 2: Figure S2A). We also expressed GFP-tagged Sec7 domain (residues 256-443) and found that it was present throughout the cytosol and did not relocate to the plasma membrane (data not shown).

Identification of potential partners of Sec7

ARF GTPases are prominent interaction partners of the mammalian cytohesins. Further partners are components of cell adhesion complexes as cytohesin-1 for example interacts with the C-terminus of β 2-integrin. Another component of focal adhesions, paxillin, was also found in a complex with cytohesins [23,24]. We carried out a preliminary analysis and used GST fusions encompassing the N-terminus (aa 1-382), the Sec7 domain (aa 256-443) and the C-terminus comprising the PH domain (aa 381-931) bound to glutathione sepharose beads, and antibodies against GFP for pull downs of potential binding partners in *sec7* expressing GFP-Sec7, AX2 and *sec7* expressing LimD-GFP for control. LimD is an unrelated protein which is present in the cytosol and near the plasma membrane and is well expressed as GFP fusion [25]. It did not show overlapping binding partners with Sec7. The proteins obtained were identified by mass spectrometry and classified as enzymes (11), cytoskeletal proteins (6), proteins involved in vesicle trafficking (14), in particular components of the endocytic machinery like NSF, the β -subunit of the AP1 complex which plays a role in clathrin-dependent protein sorting, delta adaptin belonging to the AP3 adaptor-like complex which participates in the generation of a diverse group of secretory organelles, and the gamma subunit of the coatamer complex essential for the secretory pathway, signaling proteins (4) and others (Table 1). Among

the signaling proteins were zizA and zizB, two DOCK family proteins. Interestingly, for mammalian cytohesin-2 an association with Dock180 in a complex promoted ARF-to-Rac signaling [3]. When we used GO terms for classification, the majority of proteins from the pull downs fell into the category "transport" which was closely followed by "vesicle mediated transport". Fewer proteins belonged to the category "biosynthetic process" and "small molecule metabolic process" (Additional file 3: Table S1). We confirmed the interaction for the cytoplasmic protein coronin and the 20S proteasomal subunit α -4, a further putative interaction partner, with antibodies in independent experiments (Additional file 2: Figure S2B, C).

Characterization of a *D. discoideum* Sec7 deficient mutant

The *sec7* gene of *D. discoideum* (DDB_G0272486) is located on chromosome 2. It contains one short intron of 86 nucleotides located in the first third of the gene (starting from position 947 of the cDNA sequence). *D. discoideum* cells enter a developmental program upon starvation. 8 to 12 hours after the start of starvation on phosphate agar plates cells form multicellular aggregates which transform into motile slugs at around 18 hours. Slugs are phototactic and thermotactic and finally form fruiting bodies. We followed the accumulation of the Sec7 mRNA during the life cycle and found that it is present during growth and development. The levels reach a maximum during the aggregation stage, drop after aggregation and increase again during the slug stage (Figure 4A).

For the generation of Sec7 deficient cells (*sec7*) we used a gene replacement vector which contained nucleotides 1 to 441 and 1981 to 2469 of the cDNA. The intervening gene sequence was replaced by a blasticidin resistance cassette. Successful integration of the vector into the genome was confirmed by PCR and Southern blot analysis (Figure 4B,C). Several independent transformants were isolated, the characterization of one of them is shown. In the subsequent analysis we focused primarily on properties of the cells that are associated with processes involving regulation of the actin cytoskeleton and membrane dynamics.

Sec7 deficient cells were of the same size as the parent AX2 strain and were mainly mono- and binucleated. In immunofluorescence analysis we did not detect changes when we stained for F-actin, the actin-associated protein CAP and the endoplasmic reticulum located protein disulfide isomerase (PDI) as a marker for internal membranes (Figure 5A). Mutant cells grew well in shaking suspension with duplication times comparable to AX2 and reached similar final densities (\sim 1 \times 10⁷ cells/ml). This is indicative of an unaltered macropinocytosis. Also, when grown on *E. coli* B/r as food source in shaking suspension, both *sec7* and AX2 had doubling times

Table 1 Potential binding partners of Sec7 and its domains

Dictybase ID	Description	Identified in Sec7 domain pull down	Identified in PH domain pull down (C-terminus)	Identified in GFP-Sec7 immunoprecipitation	Identified in N-domain pull down
Membrane trafficking, membrane associated processes					
DDB0237869	cog3, oligomeric Golgi complex component			+	
DDB0237794	osbH, oxysterol binding				+
DDB0185207	vatB, v-ATPase		+		
DDB0234266	vatH, v-ATPase		+		
DDB0233782	Present in macropinocytic proteome; conserved; transporter?		+		
DDB0185052	nsfA, N-ethylmaleimide-sensitive fusion protein		+		
DDB0234240	p3d1, delta adaptin (endosomal membrane)		+		
DDB0234067	Ap1b1, adaptor-related protein complex 1, beta 1 subunit, beta adaptin, highly similar to AP-1 complex subunit beta-1 (AP1B1) and AP-2 complex subunit beta-1 (AP2B1), which play a role in clathrin-dependent protein sorting	+			
DDB0304807	tgrO4, immunoglobulin E-set domain-containing protein, tgr (tiger) = Transmembrane, IPT, IG, E-set, Repeat protein	+			
DDB0233801	copG, adaptin N-terminal domain-containing protein coatomer protein complex gamma subunit gamma-COP	+			
DDB0306480	Single C-terminal TM	+			
DDB0238597	Dnajc13, DnaJ (Hsp40) homolog, subfamily C, member 13, very similar to the mammalian DnaJ homolog subfamily C member 13, required in <i>D. melanogaster</i> (Rme-8) for receptor-mediated endocytosis 8	+			
DDB0185049	myoI, class VII unconventional myosin myosin VII, similar to the conserved MYO7A; unconventional myosin required in <i>Dictyostelium</i> for phagocytosis and substrate adhesion, interacts with talA	+			
DDB0234198	vps13A, vacuolar protein sorting-associated protein 13 family protein, putative ortholog of <i>S. cerevisiae</i> VPS13, involved in vacuolar protein sorting and protein-Golgi retention	+			
Related to signal transduction					
DDB0191258	pppA, protein phosphatase 2A subunit A		+		
DDB0235201	guanylate-binding protein GTP-binding protein, 92 kDa	+			
DDB0233622	zizB, DOCK family protein, putative guanine nucleotide exchange factor (GEF)	+			
DDB0233623	zizA, DOCK family protein, putative guanine nucleotide exchange factor (GEF)	+			
Cytoskeleton					
DDB0201554	abpC, filamin		+	+	
DDB0233781	Centrosomal, 80 kDa		+		
DDB0191169	tubB, tubulin		+		
DDB0191115	corA	+	+	+	
DDB0185096	cytoplasmic dynein heavy chain, dynein beta chain, flagellar outer arm	+			

Table 1 Potential binding partners of Sec7 and its domains (Continued)

DDB0185049	myosin class VII unconventional myosin	+
	myosin VII, similar to the conserved MYO7A; unconventional myosin required in Dictyostelium for phagocytosis and substrate adhesion, interacts with talA	
DDB0191103	cortaxillin I	+

of ~3 hours suggesting that the uptake of *E. coli* was normal.

Growth on a lawn of *Klebsiella* as determined by measuring the increase in diameter of the colony was significantly reduced (Figure 5B). Such a behavior could be due to reduced phagocytosis or to altered motility. Hence we assayed the phagocytic capability following yeast particle uptake and found that fewer *sec7* cells had ingested one or more yeast particles after 45 min than AX2 cells. A quantitative evaluation showed that significantly fewer *sec7* cells (~79%) had taken up yeast cells as compared to AX2. *sec7* cells expressing GFP-Sec7 reached almost wild type level with ~95% (Additional file 4: Figure S3). The failure to completely restore wild-type behavior may lie in different expression levels and partial loss of expression.

Cell substrate and cell cell adhesion are two characteristics which are potentially influenced by the cytohesins as they affect events at the plasma membrane such as integrin activation [4]. Consequently we wanted to assess the role of Sec7 for these two aspects. Cell substrate adhesion is assayed by subjecting cells that have attached to a plastic surface to a rotation on an orbital shaker at 100 rpm. The percentage of detached cells after one hour was similar for AX2 and mutant cells. Upon increase of rotation to 160 rpm the results for wild type and mutant showed no significant difference either (data not shown). Cell cell adhesion was tested in developing cells. During development *D. discoideum* cells express specific cell surface proteins like contact site A which mediate cell adhesion and allow aggregate formation [26]. When we assayed aggregate formation during development in shaking suspension by following the decrease in optical density of the suspension we did not detect differences between wild type and mutant (data not shown). This was also taken as an indication for correct expression of the cell adhesion molecules and for normal development of the mutant.

Later developmental stages were assayed by depositing cells on phosphate agar plates for starvation. The *sec7* strains formed aggregates, mounds, slugs and fruiting bodies in a timely fashion. Furthermore, the fruiting bodies had comparable morphologies (data not shown).

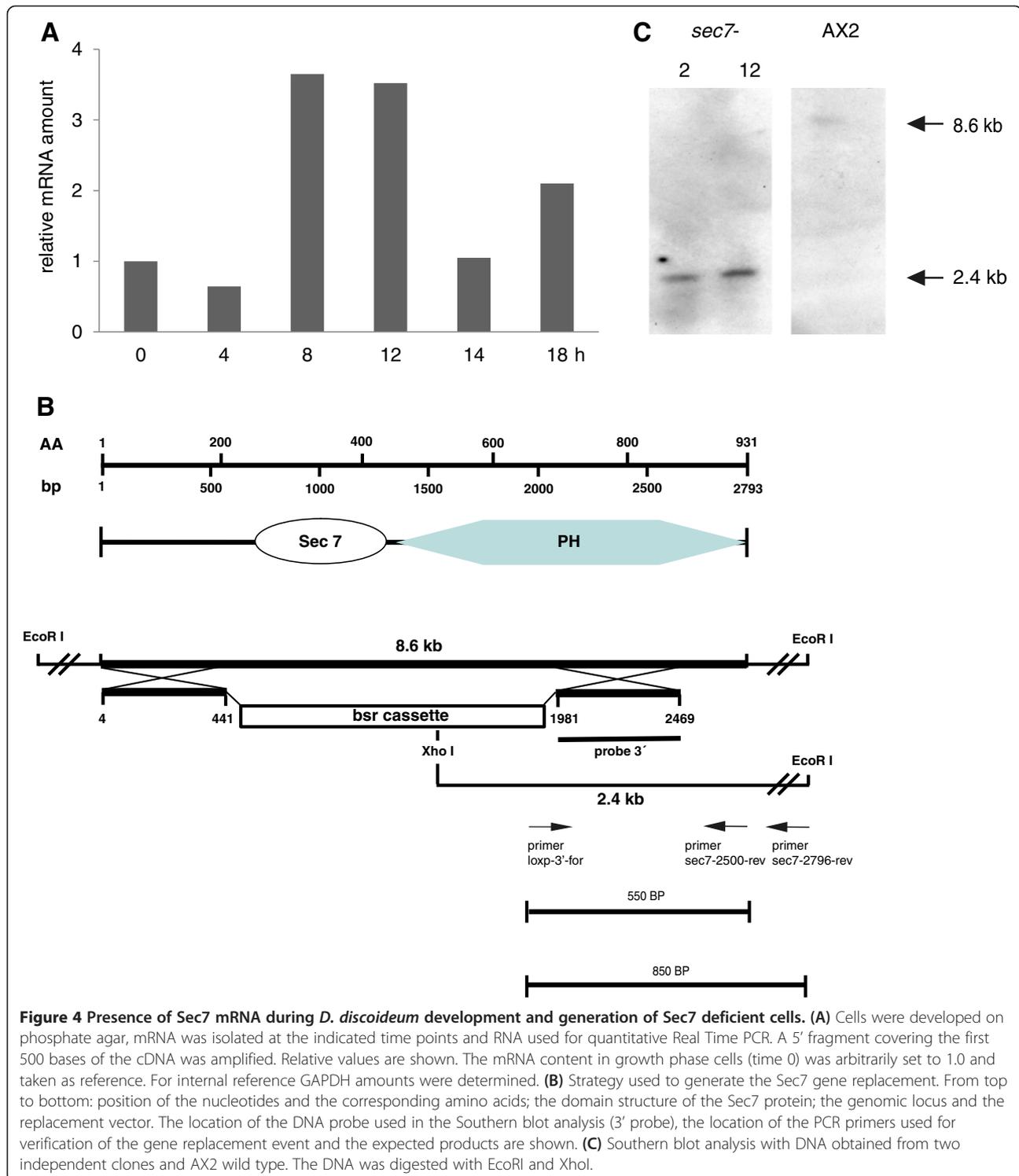
Slugs are phototactic and migrate towards light. Phototaxis is an essential feature in the wild where *Dictyostelium* has to reach the soil surface from where the spores can be dispersed. Directed migration of slugs

towards light was tested by keeping the plates in the dark and providing a lateral light source. After incubation for two days the trails of the slugs were stained with Amido Black and the migration pattern and the light sensing evaluated. AX2 slugs migrated over long distances and almost directly towards the light source. Many of the slugs had reached the edge of the plate. *sec7* slugs had a phototaxis defect. Their migration trails were shorter and the slugs never reached the edge of the plate. Moreover, their directionality was altered and the angle of deviation during slug phototaxis was increased. AX2 slugs migrate with an angle of ~13 degrees, *sec7* slugs with an angle of ~55 degrees and for *sec7* slugs expressing Sec7-GFP the angle was reduced to ~31 degrees (Figure 5C). We conclude that *sec7* slugs can sense light, but directionality is impaired.

As we had identified components associated with vesicle trafficking in our co-precipitation experiments with individual domains of Sec7 and the full length protein we tested the secretion of a lysosomal hydrolase and of cAMP phosphodiesterase. Lysosomal α -mannosidase is produced during growth and the first hours of development. The protein undergoes a variety of posttranslational modifications during its transit from the ER and Golgi to the late lysosomes before it is secreted [27]. We tested AX2 wild-type and *sec7* cells for the ability to secrete α -mannosidase during the first hours of development. ~60 and 64% of the total enzyme activity were found in the medium of AX2 and *sec7* cells, respectively, after 2 hours of development in shaking suspension. After 6 hours the values reached 75 and 77% of total enzyme activity. We concluded that secretion occurred normally in *sec7*, however, there was consistently less active enzyme present amounting to ~76% of wild-type activity (Tables 2 and 3). Secretion of cAMP phosphodiesterase was measured during early development by measuring phosphodiesterase activity in the medium. We observed a linear increase of activity in all strains, however, *sec7* had secreted less enzyme and the amounts were reduced to ~50% of wild-type level. From the analysis of the data it appears that the mutant produces less enzyme and that secretion is affected as well as. GFP-Sec7 showed an improvement reaching 60% of AX2 levels (Table 4).

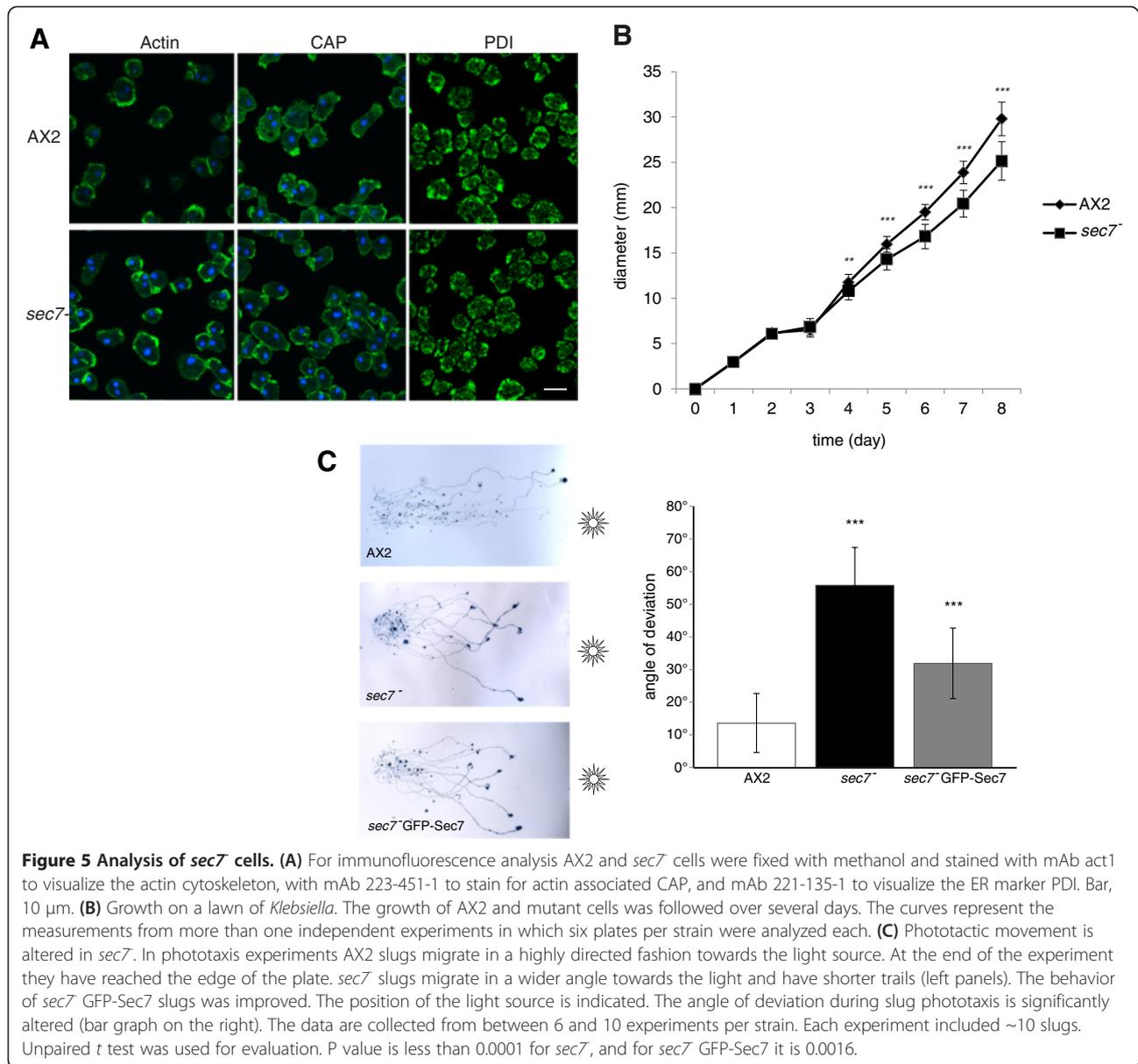
Analysis of signaling to the actin cytoskeleton, cell motility and chemotactic behaviour

On application of cAMP to cells, actin polymerization and depolymerization occurs in a characteristic pattern. After



an initial increase of the F-actin concentration, the filaments depolymerize quickly. This phase is followed by a long lasting phase of F-actin polymerization paralleling the formation of pseudopods. We tested whether Sec7 plays a role in the control of dynamic actin rearrangements and examined the actin polymerization response to

cAMP stimulation. We found that in *sec7* the relative F-actin content as revealed by TRITC-phalloidin staining of samples taken at different time points exhibited the typical biphasic pattern with peaks at 5 and 30 seconds as in AX2 suggesting that Sec7 is not required for cAMP-induced actin polymerization (Figure 6).



Dictyostelium cells exhibit an amoeboid type of cell motility. To assess the role of Sec7 for motility we analyzed the motility of single cells during the aggregation phase after six hours of starvation. Both AX2 and *sec7* cells were highly polarized and migrated towards a cAMP source, but the speed of the *sec7* cells was

significantly reduced as compared to AX2. AX2 cells migrated with a speed of $14.22 \pm 2.00 \mu\text{m}/\text{min}$, *sec7* cells with $8.66 \pm 2.25 \mu\text{m}/\text{min}$. The direction change reflecting the frequency of turning was similar, however, persistence which indicates the probability to continue movement in the same direction was significantly lower (Table 5).

Table 2 Ability of *sec7* cells to secrete enzymes during early development: secretion of α -Mannosidase

Strain	% secreted enzyme after		% total enzyme activity after 6 hours
	2 hours	6 hours	
AX2	60	75	100
<i>sec7</i>	64	77.4	75.80

Cells were starved in Soerensen phosphate buffer, pH 6.0, and at the indicated time points the α -mannosidase activity was determined. α -mannosidase secretion in percent and total α -mannosidase activity in percent are shown. The α -mannosidase activity was determined in the medium and in the cell pellet at the indicated time points after the beginning of starvation. Values are from one representative experiment.

Table 3 Ability of *sec7* cells to secrete enzymes during early development: total mannosidase activity in wild type and mutant cells at various stages of development

Strain	AX2			<i>sec7</i>		
	t0	t4	t6	t0	t4	t6
	3.72	4.42	4.26	2.3	2.58	2.67
	2.58	3.33	3.65	1.84	2.22	2.64
	1.5	2.38	2.2	1.29	2.19	2.24
	3.25	3.6	3.8	1.393	2.03	2.37
	3.73	4.12	4.44	0.74	1.05	1.25
SD	0.93948	3.5700	0.8831	1.51260	0.5752	0.5792
SEM	0.42015	0.3535	0.3949	0.58906	0.2572	0.2590
Mean	2.95600	3.5700	3.6700	0.26343	2.0140	2.2340
P value				0.0196	0.0074	0.0160

Time points (t) are given in hours. Five experiments were analyzed. For statistical analysis an unpaired t test was used. The differences were statistically significant.

Discussion

The cytohesin family of GEFs are activators for ARF GTPases. They have a broad range of roles ranging from signal transduction to cytoskeletal organization which can occur in an ARF-dependent and an ARF-independent function. Our analysis of Sec7 deficient *D. discoideum* cells revealed that loss of the protein affected several cellular processes including altered secretion, growth under certain conditions, phagocytosis, cell motility and phototaxis (Table 6). These processes are complex and several of them involve membrane flow and the actin cytoskeleton and the individual contributions are not easily deciphered. One example is phagocytosis of large particles like yeast cells which was significantly impaired whereas uptake of *E. coli* B/r was as efficient as in AX2 wild type cells. Such a differential behavior might be explained by earlier findings showing that *D. discoideum* can differentiate particles according to their surface properties and geometry. Furthermore, analysis of the actin organization around the phagosome revealed a distinct and dynamic spatiotemporal pattern [22,28]. A role of Sec7 in phagocytosis is also

supported by the potential binding partners as several of them have been implicated in this process and found associated with the phagosome [29]. We also noted changes when we tested the secretion of α -mannosidase and cAMP phosphodiesterase. Both enzymes are secreted during early development. In case of α -mannosidase we observed a similar rate of secretion as in AX2, for cAMP phosphodiesterase it was reduced. The total enzyme levels were significantly reduced for both. The second major defect was in cell migration where the *sec7* cells were slower and less persistent. For directed cell motility and chemotaxis phospholipid signaling and lipid metabolism is essential. In particular, PI(3,4,5)P₃ controls cell speed and pseudopod formation [30]. The reduced cell speed and less persistent migration of the *sec7* cells parallels the behavior of PTEN deficient *D. discoideum* cells with which Sec7 shares the constitutive localization at the plasma membrane and an absence from newly forming pseudopods. This might suggest that Sec7 acts downstream of PTEN [31].

Members of the cytohesin family of GEFs are recruited to the plasma membrane based on the affinity of their PH domain for specific phosphatidylinositol phosphates. A further mechanism is through interaction with the GTP bound form of ARF6 and ARL4 which leads to cytohesin recruitment and activation of ARF6 and ARF1 [32-34]. The activation of ARF proteins stimulates signaling pathways that regulate membrane trafficking and cell motility. Mammals have six ARFs forming three classes and playing roles in different cellular pathways [35]. *D. discoideum* harbors a single ARF homolog encoded by the *arfA* gene on chromosome 5. This is similar to *Giardia lamblia*, a flagellated protozoan parasite, which also has one ARF only. ArfA (DDB0191101) is most closely related to ARF1 with 85% identical and 90% homologous residues when compared to human ARF1 (NP_001019397.1) (E value, 3e-86) and less related to human ARF6 (CAG46762.1) sharing 67% identical and 82% homologous residues (E value 1e-69). In addition to ArfA *D. discoideum* has ten ARF related proteins, ArrA–K. ArfA is present in a macropinocytic proteome [36] whereas this has not been reported for

Table 4 Ability of *sec7* cells to secrete enzymes during early development: phosphodiesterase production

	PDE activity (U/ml) determined at the indicated time points after begin of starvation					
	AX2 (t1)	<i>sec7</i> (t1)	AX2 (t2)	<i>sec7</i> (t2)	AX2 (t3)	<i>sec7</i> (t3)
Mean	2.3767	1.1600	13.3567	6.0700	26.8433	12.6467
SD	0.6486	0.6161	2.9151	2.3226	2.0385	1.8360
SEM	0.3744	0.3557	1.6831	1.3410	1.1769	1.0600
N	3	3	3	3	3	3
P value		0.0780		0.0276		0.0034

Mutant and wild type cells were starved in Soerensen phosphate buffer, pH 6.0. At the indicated time points in hours after begin of starvation the activity of extracellular phosphodiesterase (in units/ml) was determined. The results from three experiments were evaluated. N, number of experiments. The difference in secretion obtained for *sec7* at 2 and 3 hours was statistically significant.

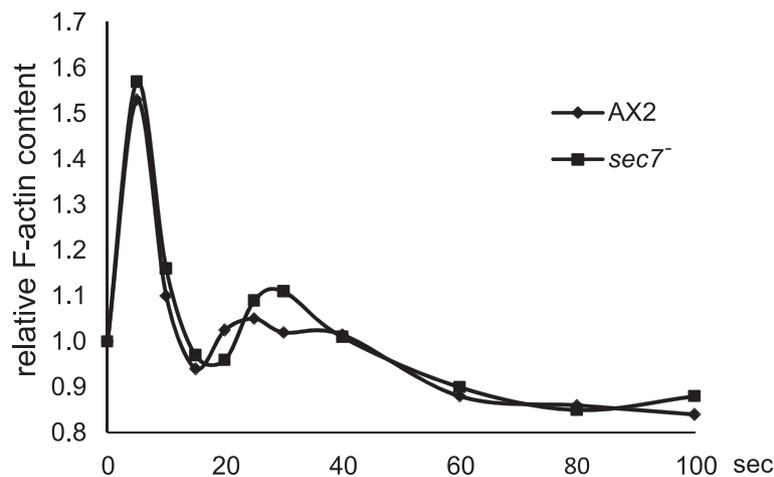


Figure 6 F-actin accumulation after a cAMP stimulus. cAMP-induced F-actin formation as measured with a phalloidin binding assay. Aggregation competent AX2 wild-type and *sec7⁻* cells were stimulated with cAMP and the F-actin content was determined at the indicated time points. Results from a representative experiment are shown.

Sec7. ArfA has also been studied by Chen et al. [37] who showed that two ArfGAPs belonging to the ASAP/ACAP type of ArfGTPases, which are distinguished by BAR, PH and Ank domains, act on ArfA. The *D. discoideum* ArfGTPases have roles in actin cytoskeleton organization and spore production. The studies on ACAP were more recently extended by Dias et al. [38] who revealed additional roles in cytokinesis, cell migration and cytoskeleton dynamics.

Another ArfA binding protein in *D. discoideum* is AdcA. This arrestin related protein bound ArfA in its GDP-bound conformation [39]. AdcA was associated with the endocytic pathway and early endosomes, and faintly labeled the plasma membrane. GFP-tagged ArfA localized in the cytosol, at the plasma membrane and mainly in the perinuclear region at the Golgi apparatus. From in vivo studies the authors concluded that ArfA-GFP was present on vesicles and tubules moving towards and away from the Golgi apparatus. They proposed a role for ArfA in trafficking events that link the Golgi to other organelles such as endosomes. Although ArfA had been found in proteomic studies on phagosomes and macropinosomes [36,40], ArfA-GFP was not convincingly located at these structures in vivo which was thought to be due to

transient interactions. The absence of GFP-Sec7 from these locations in our studies is compatible with these results.

In our attempt to identify potential binding partners of Sec7 we carried out pull downs using domains of Sec7 expressed as GST fusion proteins in *E. coli* and full length protein expressed as GFP fusion in *D. discoideum*. The majority of the putative interaction partners from all pull downs belonged to the processes of trafficking and vesicle trafficking although the overlap between the individual pull downs was limited. This could be due to the conformations of the proteins and the different accessibilities of the sequences. For some of the putative interaction partners mutants are available and their phenotypes can be compared to the one of the *sec7⁻* mutant. We did see overlaps of the phenotypes in several cases which might indicate that the proteins act in the same pathway. This applies particularly to phagocytosis and chemotactic motility (Additional file 3: Table S2).

Phosphoinositides are tightly regulated during chemotaxis in *D. discoideum*, in particular, PI(3,4,5)P₃ gradients are formed within the plasma membrane. They are thought to be of differing importance for sensing of shallow and steep cAMP gradients [30,41]. The PH domain

Table 5 Cell motility of aggregation stage cells

	Speed (µm/min)	Persistence (µm/min-deg)	Direction change (deg)	n
AX2	14.22±1.99	5.47±1.58	14.69±4.88	29
<i>sec7⁻</i>	8.66±2.25	3.3±1.33	16.28±6.84	31
P value	0.0001	0.0001	NS	

Persistence is a measure for the probability to continue movement in the same direction. The direction change (deg/min) reflects the frequency of turning, n, number of cells analyzed. The difference in speed and persistence was statistically significant. Cells from four independent experiments were evaluated. NS, not significant.

Table 6 Summary of phenotypes observed in the *sec7* strain

Phenotype	<i>sec7</i>
Cell size	wt
Nuclei number	wt
Growth in axenic medium (measure of macropinocytosis)	wt
Growth on <i>E. coli</i> in shaken suspension	wt
Growth on a lawn of <i>Klebsiella</i>	reduced
Phagocytosis of yeast cells	uptake reduced to 60% of AX2 level
Development	wt
Cell substrate adhesion	wt
Cell cell adhesion	wt
Phototaxis	directionality impaired, slug trail length reduced
Localization of F-actin and actin associated proteins (actin, CAP)	wt
Integrity of ER membrane system (PDI staining)	wt
Secretion of α -mannosidase	secretion occurs normally, however enzyme levels are reduced (~76% of AX2 level)
Secretion of phosphodiesterase	secretion appears to be impaired, enzyme levels are reduced (~50% of AX2 level)
F-actin assembly after cAMP stimulation	wt
Cell motility	speed reduced
Persistence during migration	lower

"wt" describes wild type behavior.

of the cytohesin family of ARF-GEFs can act as PI(3,4,5)P₃ sensor. We found that *D. discoideum* Sec7 had highest affinity for this phosphoinositide in lipid overlay assays followed by PI(4,5)P₂, whereas in liposome binding assays it preferred PI(4,5)P₂ and PI(3,4)P₂ over PI(3,4,5)P₃. When we analyzed GFP-tagged Sec7 in vivo the protein decorated the plasma membrane and it did not visibly associate with those membrane regions where PI(3,4,5)P₃ formation is thought to occur. The group of C. Weijer [21] had used a panel of PH domains with specificities for PI(3,4,5)P₃ and PI(3,4)P₂, among them the PI(3,4,5)P₃ specific PH domain of GRP1 (cytohesin-3), with which they analyzed the formation of these signaling molecules during phagocytosis and chemotaxis. They concluded that PI(3,4,5)P₃ levels transiently increased during phagocytosis and macropinosome formation at sites of engulfment as revealed by the recruitment of GRP1-PH to these regions. During chemotaxis towards cAMP PI(3,4,5)P₃ was formed and degraded to PI(4,5)P₂ in the plasma membrane. GRP1-PH and CRAC-PH, a *D. discoideum* protein specific for PI(3,4,5)P₃, translocated to the plasma membrane following cAMP stimulation [42]. Interestingly, the Weijer group found that different PI(3,4,5)P₃ binding PH domains behaved differently as GRP1-PH exhibited maximum binding several seconds later than other PH domains and remained at the membrane much longer which might be indicative of further determinants.

GFP-Sec7 did not exhibit a comparable pattern of plasma membrane association. Instead, it was constitutively associated with the plasma membrane and consistently disappeared from regions forming a new pseudopod, undergoing phagocytosis or macropinocytosis which require the insertion of new membrane. Sec7-GFP showed a behavior complementary to the one of PH domains sensing PI(3,4,5)P₃ and rather resembled the pattern reported for the phosphatase PTEN and the PH domain of PLC δ [43]. Like Sec7 deficient cells the PTEN null cells showed an impairment in phagocytosis of yeast cells and a normal uptake of bacteria [20]. They also had a cell migration defect [43,44].

Conclusion

The analysis of a *Dictyostelium* Sec7 mutant implicates the protein in processes that are related to membrane flow and actin dynamics and reveals a conserved function for this class of proteins. Its PH domain has the potential to recognize PI(4,5)P₂ and to a lesser extent PI(3,4,5)P₃ in vitro. The protein associates with the plasma membrane, however, during macropinocytosis, phagocytosis and pseudopod extension it is lost. It therefore does not act as a sensor for 3-phosphoinositide dynamics. This is supported by the absence of the sentry glutamate in Sec7 which in the cytohesins is essential for specific plasma membrane targeting. Instead, the glutamate is replaced by glutamine. The Sec7 plasma membrane

association is presumably mediated by PI(4,5)P₂ binding, however, other determinants such as binding to interacting proteins might also be important factors.

Materials and methods

Growth and development of *D. discoideum* strains and mutant generation

D. discoideum strains used were AX2, a *Sec7* deficient strain derived from AX2 (*sec7*), and *sec7* expressing GFP-*Sec7*. They were grown in shaking suspension (160 rpm) in axenic medium at 22°C or on a lawn of *Klebsiella* on SM agar plates [44]. For growth on *E. coli* in shaking suspension (160 rpm) *E. coli* B/r was harvested and resuspended in Soerensen phosphate buffer (17 mM sodium-potassium-phosphate, pH 6.0) at a density of 10¹⁰ cells/ml. Inoculation was with 5×10⁵ *D. discoideum* cells/ml. Growth was determined by following the increase in cell number over time. Development was done with cells starved in Soerensen phosphate buffer at a density of 1×10⁷ cells/ml in shaking suspension. Under these conditions, development proceeded until the tight aggregate stage. Upon development on a solid substratum fruiting body formation occurred. For this 5×10⁷ cells were spread onto phosphate agar plates (10 cm in diameter) and incubated at 22°C until fruiting bodies had formed. For evaluation of slug migration and phototaxis 5×10⁵ cells in 5 μl Soerensen phosphate buffer were spotted in the center of a water agar plate. Incubation was in the dark with a lateral light source. After 48 hours cells had formed slugs which migrated towards the light. They were transferred to nitrocellulose filters and detected by staining with Amido Black (0.1% in 25% isopropanol and 10% acidic acid).

The *Sec7* cDNA was amplified from cDNA that had been prepared from strain AX2, cloned into pGEM-T Easy (Promega). The sequence was verified and used for all further cloning steps. For inactivation of the *Sec7* gene, a gene replacement vector was generated. Residues 4 to 441 and 1981 to 2469 of the cDNA were cloned into pGEM-T Easy carrying a Blastidicin resistance gene under the control of the actin 15 promoter [45]. The plasmid was transformed into AX2 and transformants were selected using Blastidicin S (MP Biomedicals, Eschwege, Germany) at 1.5 μg/ml. Single colonies were selected on a *Klebsiella* lawn, DNA was isolated from nuclei using phenol/chloroform extraction [44] and PCR analysis was carried out with primers that revealed the gene replacement event. The gene replacement was further confirmed by Southern blot analysis. Several independent clones were identified, two of them were further analysed. As the results did not differ, the characterization of one of the clones is presented in the Results section.

Expression of recombinant protein

For expression of recombinant *Sec7* polypeptides as glutathione S transferase (GST) fusion proteins in *E. coli*, cDNA fragments encoding the N-terminal domain (amino acid residues 1-382), the *Sec7* domain (amino acid residues 256-443) and the C-terminus encompassing the PH domain (amino acid residues 381-931) and a polypeptide containing only the PH domain (amino acid residues 454-577) were cloned into pGEX vectors (GE Healthcare Life Sciences). *E. coli* strains XL1 Blue and, in case of the *Sec7* domain, Arctic Express Ril (Agilent Technologies) were used for expression of the GST fusion proteins. Full length *Sec7* cDNA was cloned into pDex79 and expressed as GFP fusion (GFP-*Sec7*) under control of the actin 15 promoter [46]. GFP was fused to the N-terminus of *Sec7*. The plasmid was transformed into *sec7* cells. Selection was with G418 (Life Technologies Corporation) at 4 μg/ml. For expression of the *Sec7* domain, cDNA sequences corresponding to amino acid residues 252-443 were cloned into pDex79.

Pull down assays and immunoprecipitation

To identify interaction partners of *Sec7*, GST-*Sec7* N-terminus, GST-*Sec7* domain, GST-*Sec7* C-terminus and GST for control were bound to glutathione sepharose 4B beads (GE Healthcare) and used for pull down assays. Incubation with cell lysates varied from 2 hours to overnight and was performed at 4°C. Lysates were prepared from AX2 cells, *sec7* expressing *Sec7*-GFP and *sec7* cells expressing LimD-GFP [25] using the following buffer: 10 mM Tris-HCl, pH 7.5, 150 mM NaCl, 1% NP 40, 0.5 mM EDTA, 1 mM PMSF and protease inhibitors (Sigma). Lysis was controlled microscopically. For immunoprecipitation of GFP-*Sec7*, monoclonal antibodies (mAb K3-184-2) as well as polyclonal antibodies against GFP bound to protein A sepharose beads were used [47]. Protein A sepharose beads were used as control to exclude proteins that bind to the sepharose matrix. The proteins from pull downs, immunoprecipitations and controls were separated by SDS-PAGE (10% to 12% acrylamide), stained with Coomassie Blue, bands from control and experiment were cut out and proteins processed for LC-MS at the Bioanalytics Facility of the CMMC. The Mascot search engine was used for identification of the proteins. For verification of the interaction immunoprecipitation of GFP-*Sec7* was repeated and probed for the presence of coronin and subunit α-4 (psmA4, (DDB0214953)) of the 20S proteasome using mAb 176-3-6 and 159-83-10, respectively [48,49].

Mutant analysis

Growth analysis, uptake of yeast particles and adhesion assays were done as described [5], analysis of F-actin assembly after cAMP stimulation was done as described [47]. Mannosidase activity in cell pellets and in the

supernatant was determined according to Loomis [50]. In brief, cells were starved at a density of 1×10^7 cells/ml. At the beginning of the experiment (t_0) and after 2, 4 and 6 hours 500 μ l were taken to measure mannosidase activity. For determination of secreted enzyme 100 μ l of the supernatant were mixed with 100 μ l Na-citrate buffer (pH 5.0) and 200 μ l substrate solution (2 μ l p-nitrophenyl- α -D mannopyranoside (150 mM). The substrate was dissolved in DMF. The reaction was stopped after 30 min incubation at 37°C by addition of 600 μ l sodium borate (0.2 M, pH 9.8) and the product extracted into butanol. Nitrophenol formation was estimated by measuring the absorbance at 405 nm. For determination of total enzyme activity cells were lysed by addition of Triton X-100 (0.5%). cAMP phosphodiesterase (PDE) activity was determined using a coupled enzymatic assay with cAMP as substrate [51]. AMP, the product of the hydrolysis, was further converted to IMP by adenosine deaminase. IMP was then converted to inosine by alkaline phosphatase. The decrease of the absorption at 265 nm was a measure of inosine formation which in contrast to adenosine does not absorb light at 265 nm. Reagents and enzymes were from Sigma. For immunofluorescence analysis methanol fixed cells were stained for actin (mAb act1 [52]), CAP (mAb 223–445-1 [53]), protein disulfide isomerase (PDI, mAb 221-135-1 [54]) and annexin 7 (mAb 185-338-1 [19]). For detection goat anti mouse antibodies coupled to Alexa Fluor 488 (Life technologies) were used. Analysis of fixed and living cells was done by laser scanning confocal microscopy using a Leica TCS SP5 microscope.

For cell motility analysis cells were plated after ~ 6 hours of starvation in a chamber (ibidi GmbH-Martinsried, Germany) and migration towards aggregation centers or towards a micropipette filled with 10 μ m cAMP was followed. Analysis was carried out by using the DIAS system as described [5]. For phototaxis 5×10^5 cells are placed in the center of a water agar plate. Slugs were allowed to form and migrate towards light. After 48 h, slugs and slime trails were transferred to nitrocellulose filters and stained with Amido Black.

Liposome binding assay

Phosphatidylserine (PS), phosphatidylcholine (PC), phosphatidylethanolamine (PE), PI(3)P, PI(4)P, PI(5)P, PI(3,4)P2, PI(3,5)P2, PI(4,5)P2, and PI(3,4,5)P3 were obtained from Sigma and diluted in chloroform. Liposome binding experiments were performed with a modified published liposome binding assay protocol [55]. Lipid mixtures containing 65% PC, 20% PE, 5% PS and 10% individual phosphoinositides were produced by mixing appropriate lipid solutions in chloroform/methanol. Slow flow nitrogen gas was used for the production of a film on the glass and vacuum desiccation for 30 min for solvent

removal. Sterile-filtered sucrose binding buffer (20 mM HEPES, pH 7.4, 100 mM KCl, 1 mM EDTA and 0.1 M sucrose) was added to a final lipid concentration of 1mg/ml and incubated at 37°C for 2 hrs. Lipids were then sonicated in a waterbath-sonicator for 10 sec.

To test liposome binding, a 100 μ l reaction mixture of freshly prepared liposomes and 5 μ g of purified protein were incubated for 15 min at room temperature and centrifuged at $100,000 \times g$ (42,000 rpm) at 4°C for 25 min in a Beckman table top ultracentrifuge Optima TLX (TLA 45 rotor). The supernatant was saved, and the pellet was resuspended in 100 μ l of sucrose binding buffer. Both fractions were then analyzed by SDS-PAGE followed by Coomassie blue staining. ImageJ was used for quantification.

Miscellaneous methods

Phosphoinositide-binding assays using lipid strips supplied by Echelon Biosciences, Inc. (Salt Lake City, Utah, USA) were performed as described [56]. For statistical analysis the Student's *t* test was used. For cell fractionation cells were lysed using Nuclepore filters (Whatman) in 20 mM Tris-HCl, pH 8.0, 50 mM NaCl and protease inhibitors (Sigma). Sequential centrifugation steps were done at $400 \times g$ (2 min) to remove unlysed cells, $10,000 \times g$ (10 min) to pellet nuclei and $100,000 \times g$ (60 min) to separate membrane and cytosolic fractions. Proteins were separated on SDS-PA gels (10% acrylamide), blotted onto nitrocellulose membranes and GFP-tagged protein detected with mAb K3-184-2 and for a cytosolic control protein using enhanced chemiluminescence [47]. Total RNA was isolated using phenol extraction. Quantitative Real Time PCR was done as described [57].

Additional files

Additional file 1: Figure S1. Evolutionary tree of the Sec7-PH domains. CLUSTALX alignments of the Sec7-PH domains from *Homo sapiens* and *Dictyostelium discoideum* and other selected organisms were used to create dendograms with TreeView. Boot strap values are provided at the node of each branch. The scale bar indicates amino acid substitutions per site. Organisms used: hs: *Homo sapiens*, ce: *Caenorhabditis elegans*, dd: *Dictyostelium discoideum*, dm: *Drosophila melanogaster*, sp: *Schizosaccharomyces pombe*, mo: *Magnaporthe oryzae*, as: *Arabidopsis thaliana*, Os: *Oryza sativa*.

Additional file 2: Figure S2. (A) Cell fractionation reveals the presence of GFP-Sec7 in the cytosol and in membrane fractions. The cells were opened using Nuclepore filters and aliquots separated by SDS PAGE (10% acrylamide). Proteins were detected with mAb K3-184-2 [47]. L, whole cell lysate, P0 (400 $\times g$); SN1, P1 (10,000 $\times g$); SN2, P2 (100,000 $\times g$). SN, supernatant; P, pellet. The signal obtained for whole cell lysate, 10,000 $\times g$ pellet and supernatant and 100,000 $\times g$ supernatant represents the material from 2×10^5 cells/ml, the signal in the 100,000 $\times g$ pellet corresponds to 2×10^7 cells/ml. **(B)** Coronin and proteasomal subunit interact with GFP-Sec7. GFP-Sec7 was immunoprecipitated using GFP antibodies and the pull down probed for the presence of coronin using mAb 176-3-6 and mAb 159-83-10 to detect the 28 kDa proteasomal subunit psmA4 (DDB0214953). psmA4 was observed in one pull down

experiment only and was therefore not included in Table 1. Here it proved to be a binding partner. For control, *sec7* expressing GFP-LimD was used. **(C)** Coimmunoprecipitation of GFP-Sec7 with coronin. Protein A sepharose beads carrying mAb 176-3-6 were used to precipitate coronin from cell lysates of *sec7* cells expressing GFP-Sec7 or GFP-LimD and the immunoprecipitates probed for the presence of GFP-Sec7 using GFP-specific antibodies.

Additional file 3: Table S1. GO classification of the Sec7 interaction partners and **Table S2** Mutant phenotypes of potential Sec7 interaction partners. The GO analysis was carried out at <http://go.princeton.edu/cgi-bin/GOTermMapper>.

Additional file 4: Figure S3. *sec7* cells have a phagocytosis defect. AX2, *sec7* and *sec7* expressing GFP-Sec7 cells were incubated with yeast and fixed after 45 minutes. The number of cells containing yeast particles was determined. The results shown are from three independent experiments. P values are given. The difference between *sec7* and AX2 is significant.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

AAN, TJR, WK and MS conceived the project, RM, CH, SKS, NNO, MP, TJR, MS, KS, MT, KY, RB-W, and CG designed and performed the experiments and analyzed the data. AAN, TJR, WK and MS analyzed the data and wrote the manuscript. All authors read and approved the final manuscript.

Acknowledgments

Work in the authors' labs (AAN and WK) is supported by the DFG, SFB 670 and 832, respectively, MS by SFB 863 and 914. TR is supported by SFB 670, SKS is a member of the CECAD graduate school. We thank Berthold Gaßen for antibody generation, Debora Hofmann and Philipp Niehues for help with immunofluorescence analysis, and Dr. S. Müller (CMMC) for performing mass spectrometry analysis.

Author details

¹Institute of Biochemistry I, Medical Faculty, Center for Molecular Medicine Cologne (CMMC) and Cologne Excellence Cluster on Cellular Stress Responses in Aging-Associated Diseases (CECAD), University of Cologne, 50931 Köln, Germany. ²Institute of Biochemistry II, Medical Faculty, University of Cologne, 50931 Köln, Germany. ³Institute of Anatomy and Cell Biology, Ludwig-Maximilians-University, 80336 München, Germany. ⁴Laboratory of Molecular Immunology, LIMES Institute of the University of Bonn, Bonn, Germany.

Received: 21 January 2013 Accepted: 29 July 2013

Published: 1 August 2013

References

1. Casanova JE: Regulation of Arf activation: the Sec7 family of guanine nucleotide exchange factors. *Traffic* 2007, **8**:1476–1485.
2. Nagel W, Schilcher P, Zeitlmann L, Kolanus W: The PH domain and the polybasic c domain of cytohesin-1 cooperate specifically in plasma membrane association and cellular function. *Mol Biol Cell* 1998, **9**:1981–1994.
3. White DT, McShea KM, Attar MA, Santy LC: GRASP and IPCEF promote ARF-to-Rac signaling and cell migration by coordinating the association of ARNO/cytohesin 2 with Dock180. *Mol Biol Cell* 2010, **21**:562–571.
4. Quast T, Tappertzhofen B, Schild C, Grell J, Czeloth N, Förster R, Alon R, Fraemohs L, Dreck K, Weber C, Lämmermann T, Sixt M, Kolanus W: Cytohesin-1 controls the activation of RhoA and modulates integrin-dependent adhesion and migration of dendritic cells. *Blood* 2009, **113**:5801–5810.
5. Shina MC, Müller R, Blau-Wasser R, Glöckner G, Schleicher M, Eichinger L, Noegel AA, Kolanus W: A cytohesin homolog in *dictyostelium* amoebae. *PLoS One* 2010, **5**:e9378. doi:10.1371/journal.pone.0009378.
6. Gillingham AK, Munro S: The small G proteins of the Arf family and their regulators. *Annu Rev Cell Dev Biol* 2007, **23**:579–611.
7. Eichinger L, Pachebat JA, Glöckner G, Rajandream MA, Suggang R, et al: The genome of the social amoeba *dictyostelium discoideum*. *Nature* 2005, **435**:43–57.
8. Heidel AJ, Lawal HM, Felder M, Schilde C, Helps NR, Tunggal B, Rivero F, John U, Schleicher M, Eichinger L, Platzer M, Noegel AA, Schaap P, Glöckner G: Phylogeny-wide analysis of social amoeba genomes highlights ancient origins for complex intercellular communication. *Genome Res* 2011, **21**:1882–1891.
9. Mossessova E, Gulbis JM, Goldberg J: Structure of the guanine nucleotide exchange factor Sec7 domain of human Arno and analysis of the interaction with ARF GTPase. *Cell* 1998, **92**:415–423.
10. Lowery J, Szul T, Seetharaman J, Jian X, Su M, Forouhar F, Xiao R, Acton TB, Montelione GT, Lin H, Wright JW, Lee E, Holloway ZG, Randazzo PA, Tong L, Sztul E: Novel C-terminal motif within Sec7 domain of guanine nucleotide exchange factors regulates ADP-ribosylation factor (ARF) binding and activation. *J Biol Chem* 2011, **286**:36898–36906.
11. Lietzke SE, Bose S, Cronin T, Klarlund J, Chawla A, Czech MP, Lambright DG: Structural basis of 3-phosphoinositide recognition by pleckstrin homology domains. *Mol Cell* 2000, **6**:385–394.
12. Ferguson KM, Kavran JM, Sankaran VG, Fournier E, Isakoff SJ, Skolnik EY, Lemmon MA: Structural basis for discrimination of 3-phosphoinositides by pleckstrin homology domains. *Mol Cell* 2000, **6**:373–384.
13. Klarlund JK, Guilherme A, Holik JJ, Virbasius JV, Chawla A, Czech MP: Signaling by phosphoinositide-3,4,5-trisphosphate through proteins containing pleckstrin and Sec7 homology domains. *Science* 1997, **275**:1927–1930.
14. Klarlund JK, Tsiaras W, Holik JJ, Chawla A, Czech MP: Distinct polyphosphoinositide binding selectivities for pleckstrin homology domains of GRP1-like proteins based on diglycine versus triglycine motifs. *J Biol Chem* 2000, **275**:32816–32821.
15. Rebecchi MJ, Scarlata S: Pleckstrin homology domains: a common fold with diverse functions. *Annu Rev Biophys Biomol Struct* 1998, **27**:503–528.
16. Lemmon MA: Pleckstrin homology domains: not just for phosphoinositides. *Biochem Soc Trans* 2004, **32**:707–711.
17. Pilling C, Landgraf KE, Falke JJ: The GRP1 PH domain, like the AKT1 PH domain, possesses a sentry glutamate residue essential for specific targeting to plasma membrane PI(3,4,5)P(3). *Biochemistry* 2011, **50**:9845–9856.
18. Narayan K, Lemmon MA: Determining selectivity of phosphoinositide-binding domains. *Methods* 2006, **39**:122–133.
19. Döring V, Veretout F, Albrecht R, Mühlbauer B, Schlatterer C, Schleicher M, Noegel AA: The in vivo role of annexin VII (synexin): characterization of an annexin VII-deficient *dictyostelium* mutant indicates an involvement in Ca(2+)-regulated processes. *J Cell Sci* 1995, **108**:2065–2076.
20. Hacker U, Albrecht R, Maniak M: Fluid-phase uptake by macropinocytosis in *dictyostelium*. *J Cell Sci* 1997, **110**:105–112.
21. Dormann D, Weijer G, Dowler S, Weijer CJ: In vivo analysis of 3-phosphoinositide dynamics during *dictyostelium* phagocytosis and chemotaxis. *J Cell Sci* 2004, **117**:6497–6509.
22. Clarke M, Engel U, Giorgione J, Müller-Taubenberger A, Prassler J, Veltman D, Gerisch G: Curvature recognition and force generation in phagocytosis. *BMC Biol* 2010, **8**:154.
23. Geiger C, Nagel W, Boehm T, van Kooyk Y, Figdor CG, Kremmer E, Hogg N, Zeitlmann L, Dierks H, Weber KSC, Kolanus W: Cytohesin-1 regulates β -2 integrin-mediated adhesion through both ARF-GEF function and interaction with LFA-1. *EMBO J* 2000, **19**:2525–2536.
24. Torii T, Miyamoto Y, Sanbe A, Nishimura K, Yamauchi J, Tanoue A: Cytohesin-2/ARNO, through its interaction with focal adhesion adaptor protein paxillin, regulates preadipocyte migration via the downstream activation of Arf6. *J Biol Chem* 2010, **285**:24270–24281.
25. Khurana B, Khurana T, Khaire N, Noegel AA: Functions of LIM proteins in cell polarity and chemotactic motility. *EMBO J* 2002, **21**:5331–5342.
26. Bozzaro S, Ponte E: Cell adhesion in the life cycle of *dictyostelium*. *Experientia* 1995, **51**:175–188.
27. Mierendorf RC Jr, Cardelli JA, Dimond RL: Pathways involved in targeting and secretion of a lysosomal enzyme in *dictyostelium discoideum*. *J Cell Biol* 1985, **100**:1777–1787.
28. Vogel G, Thilo L, Schwarz H, Steinhart R: Mechanism of phagocytosis in *dictyostelium discoideum*: phagocytosis is mediated by different recognition sites as disclosed by mutants with altered phagocytotic properties. *J Cell Biol* 1980, **86**:456–465.
29. Lefkir Y, Malbouyres M, Gotthardt D, Ozinsky A, Cornillon S, Bruckert F, Aderem AA, Soldati T, Cosson P, Letourneur F: Involvement of the AP-1 adaptor complex in early steps of phagocytosis and macropinocytosis. *Mol Biol Cell* 2004, **15**:861–869.

30. Kölsch V, Charest PG, Firtel RA: **The regulation of cell motility and chemotaxis by phospholipid signaling.** *J Cell Sci* 2008, **121**:551–559.
31. Wessels D, Lusche DF, Kuhl S, Heid P, Soll DR: **PTEN plays a role in the suppression of lateral pseudopod formation during *dictyostelium* motility and chemotaxis.** *J Cell Sci* 2007, **120**:2517–2531.
32. Cohen LA, Honda A, Varnai P, Brown FD, Balla T, Donaldson JG: **Active Arf6 recruits ARNO/cytohesin GEFs to the PM by binding their PH domains.** *Mol Biol Cell* 2007, **18**:2244–2253.
33. Hofmann I, Thompson A, Sanderson CM, Munro S: **The Arl4 family of small G proteins can recruit the cytohesin Arf6 exchange factors to the plasma membrane.** *Curr Biol* 2007, **17**:711–716.
34. Li CC, Chiang TC, Wu TS, Pacheco-Rodriguez G, Moss J, Lee FJ: **ARL4D Recruits cytohesin-2/ARNO to modulate actin remodeling.** *Mol Biol Cell* 2007, **18**:4420–4437.
35. Donaldson JG, Jackson CL: **ARF family G proteins and their regulators: roles in membrane transport, development and disease.** *Nat Rev Mol Cell Biol* 2011, **12**:362–375.
36. Journet A, Klein G, Brugière S, Vandenbrouck Y, Chapel A, Kieffer S, Bruley C, Masselon C, Aubry L: **Investigating the macropinocytic proteome of *dictyostelium* amoebae by high-resolution mass spectrometry.** *Proteomics* 2012, **12**:241–245.
37. Chen PW, Randazzo PA, Parent CA: **ACAP-a/B are ArfGAP homologs in *dictyostelium* involved in sporulation but not in chemotaxis.** *PLoS One* 2010, **5**:e8624. doi:10.1371/journal.pone.0008624.
38. Dias M, Blanc C, Thazar-Poulot N, Ben Larbi S, Cosson P, Letourneur F: ***Dictyostelium* ACAP-a is an ArfGAP involved in cytokinesis, cell migration and actin cytoskeleton dynamics.** *J Cell Sci* 2013, **126**:756–766.
39. Guetta D, Langou K, Grunwald D, Klein G, Aubry L: **FYVE-dependent endosomal targeting of an arrestin-related protein in amoeba.** *PLoS One* 2010, **5**:e13249.
40. Gotthardt D, Blancheteau V, Bosserhoff A, Ruppert T, Delorenzi M, Soldati T: **Proteomics fingerprinting of phagosome maturation and evidence for the role of a galpha during uptake.** *Mol Cell Proteomics* 2006, **5**:2228–2243.
41. Hoeller O, Kay RR: **Chemotaxis in the absence of PIP3 gradients.** *Curr Biol* 2007, **17**:813–817.
42. Parent CA, Blacklock BJ, Froehlich WM, Murphy DB, Devreotes PN: **G protein signaling events are activated at the leading edge of chemotactic cells.** *Cell* 1998, **95**:81–91.
43. Iijima M, Devreotes P: **Tumor suppressor PTEN mediates sensing of chemoattractant gradients.** *Cell* 2002, **109**:599–610.
44. Funamoto S, Meili R, Lee S, Parry L, Firtel RA: **Spatial and temporal regulation of 3-phosphoinositides by PI 3-kinase and PTEN mediates chemotaxis.** *Cell* 2002, **109**:611–623.
45. Adachi H, Hasebe T, Yoshinaga K, Ohta T, Sutoh K: **Isolation of *dictyostelium discoideum* cytokinesis mutants by restriction enzyme-mediated integration of the blasticidin S resistance marker.** *Biochem Biophys Res Commun* 1994, **205**:1808–1814.
46. Westphal M, Jungbluth A, Heidecker M, Mühlbauer B, Heizer C, Schwartz JM, Marriott G, Gerisch G: **Microfilament dynamics during cell movement and chemotaxis monitored using a GFP-actin fusion protein.** *Curr Biol* 1997, **7**:176–183.
47. Noegel AA, Blau-Wasser R, Sultana H, Müller R, Israel L, Schleicher M, Patel H, Weijer CJ: **The cyclase-associated protein CAP as regulator of cell polarity and cAMP signaling in *dictyostelium*.** *Mol Biol Cell* 2004, **15**:934–945.
48. de Hostos EL, Bradtke B, Lottspeich F, Guggenheim R, Gerisch G: **Coronin, an actin binding protein of *dictyostelium discoideum* localized to cell surface projections, has sequence similarities to G protein beta subunits.** *EMBO J* 1991, **13**:4097–4104.
49. Schauer TM, Nesper M, Kehl M, Lottspeich F, Müller-Taubenberger A, Gerisch G, Baumeister W: **Proteasomes from *dictyostelium discoideum*: characterization of structure and function.** *J Struct Biol* 1993, **111**:135–147.
50. Loomis WF Jr: **Developmental regulation of alpha-mannosidase in *dictyostelium discoideum*.** *J Bacteriol* 1970, **103**:375–381.
51. Riedel V, Gerisch G: **Regulation of extracellular cyclic-AMP-phosphodiesterase activity during development of *dictyostelium discoideum*.** *Biochem Biophys Res Commun* 1971, **42**:119–124.
52. Simpson PA, Spudis JA, Parham P: **Monoclonal antibodies prepared against *dictyostelium* actin: characterization and interactions with actin.** *J Cell Biol* 1984, **99**:287–295.
53. Gottwald U, Brokamp R, Karakesisoglou I, Schleicher M, Noegel AA: **Identification of a cyclase-associated protein (CAP) homologue in *dictyostelium discoideum* and characterization of its interaction with actin.** *Mol Biol Cell* 1996, **7**:261–272.
54. Monnat J, Hacker U, Geissler H, Rauchenberger R, Neuhaus EM, Maniak M, Soldati T: ***Dictyostelium discoideum* protein disulfide isomerase, an endoplasmic reticulum resident enzyme lacking a KDEL-type retrieval signal.** *FEBS Lett* 1997, **418**:357–362.
55. Blume JJ, Halbach A, Behrendt D, Paulsson M, Plomann M: **EHD proteins are associated with tubular and vesicular compartments and interact with specific phospholipids.** *Exp Cell Res* 2007, **313**:219–231.
56. Riyahi TY, Frese F, Steinert M, Omosigbo NN, Glöckner G, Eichinger L, Orabi B, Williams RS, Noegel AA: **RpkA, a highly conserved GPCR with a lipid kinase domain, has a role in phagocytosis and anti-bacterial defense.** *PLoS One* 2011, **6**:e27311.
57. Rashmi RN, Eckes B, Glöckner G, Groth M, Neumann S, Gloy J, Sellin L, Walz G, Schneider M, Karakesisoglou I, Eichinger L, Noegel AA: **The nuclear envelope protein Nesprin-2 has roles in cell proliferation and differentiation during wound healing.** *Nucleus* 2012, **3**:172–186.

doi:10.1186/1478-811X-11-54

Cite this article as: Müller et al.: The cytohesin paralog Sec7 of *Dictyostelium discoideum* is required for phagocytosis and cell motility. *Cell Communication and Signaling* 2013 **11**:54.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

