



Pathogenic mechanisms and clinical implications of congenital neutropenia syndromes

Fabian Hauck and Christoph Klein

Purpose of review

The purpose of this review is to summarize pathogenic mechanisms and clinical implications of the most illustrative genetic entities of congenital neutropenia syndromes.

Recent findings

Congenital neutropenia comprise monogenetic entities with or without additional immunologic and extrahaematopoietic syndromic features. Continuous careful explorations of known entities such as ELANE, GFI1, HAX1, G6PC3 deficiency and XLN help to define principles controlling differentiation and function of neutrophil granulocytes. Furthermore, the identification of novel genetic defects associated with congenital neutropenia, such as VPS45 deficiency, broadens our understanding of neutrophil biology. Pathogenic mechanisms imply protein and vesicle mistrafficking, endoplasmic reticulum stress, the unfolded protein response, destabilization of the mitochondrial membrane potential, disturbed energy metabolism, dysglycosylation and deregulated actin polymerization.

Summary

Advanced genetic and biochemical techniques have helped to expand our knowledge of congenital neutropenia syndromes. Known and novel genetic entities shed light on fundamental biological processes important for the homeostasis and functioning not only of the neutrophil granulocyte but as well of the entire haematopoietic system. Furthermore, treatment decisions become more tailored and might pave the road towards personalized molecular medicine.

Keywords

elastase neutrophil expressed, glucose 6 phosphatase catalytic subunit 3, growth factor independent 1 transcription repressor, haematopoietic cell specific LYN substrate 1 associated protein X-1, severe congenital neutropenia, vacuolar protein sorting 45 homolog, X-linked neutropenia

INTRODUCTION

Neutrophil granulocytes are the most abundant type of leukocytes and their inherited quantitative disorders are referred to as congenital neutropenia with or without additional immunologic and extrahaematopoietic manifestations (Table 1 [1–14,15^a, 16–41,42^{aa},43–53,54^a,55–61,62^{aa},63^a,64^{aa},65–76, 77^a,78–95,96^{aa},97–102]). The absolute neutrophil count (ANC) in the peripheral blood is used for dividing congenital neutropenia into mild ($1.0\text{--}1.5 \times 10^9/l$), moderate ($0.5\text{--}1.0 \times 10^9/l$) and severe ($<0.5 \times 10^9/l$) subtypes. Neutropenia may be constant, intermittent or periodic such as in cyclic neutropenia (CyN). Frequently, in congenital neutropenia, compensatory monocytosis is seen [103^a,104–106].

Patient registries founded in the early 1990s have allowed calculating an estimated incidence of $10\text{--}15/1 \times 10^6$ birth and a prevalence of more than $10/1 \times 10^6$ inhabitants. Concerted international

collaborations have facilitated the discovery of genetic defects associated with severe congenital neutropenia (SCN). However, even in the era of next-generation sequencing, the genetic cause of congenital neutropenia remains unknown in 30–50% of patients [107^a].

Clinically, congenital neutropenia are characterized by acute and life-threatening invasive bacterial and fungal infections such as cellulitis, pneumonia and sepsis and chronic stomatologic infections such as recurrent aphthosis, paradontopathy and tooth

Dr von Hauner Children's Hospital, Ludwig-Maximilians-University Munich, Munich, Germany

Correspondence to Christoph Klein, Dr von Hauner University Children's Hospital, Lindwurmstrasse 4, 80337 Munich, Germany. Tel: +49 89 5160 7700; fax: +49 89 5160 7702; e-mail: christoph.klein@med.uni-muenchen.de

Curr Opin Allergy Clin Immunol 2013, 13:596–606

DOI:10.1097/ACI.000000000000014

KEY POINTS

- Congenital neutropenia comprises monogenetic entities with or without additional immunologic and extrahaematopoietic manifestations.
- Pathogenic mechanisms imply RNA processing, protein and vesicle mistrafficking, endoplasmic reticulum stress, the unfolded protein response, destabilization of the mitochondrial membrane potential, disturbed energy metabolism and dysglycosylation, as well as deregulated actin polymerization.
- Molecular diagnosis is important for risk stratification, treatment choice and prognosis.

loss [107[■]]. Moreover, congenital neutropenia is considered a premalignant state of haematopoietic stem or progenitor cells and may degenerate into myelodysplastic syndrome (MDS) and acute myeloblastic leukaemia (AML) [108,109].

Granulocyte colony stimulating factor (G-CSF), which has been commercially available since 1993, has emerged as an effective first-line treatment and importantly reduced the mortality of congenital neutropenia [110,111]. However, a cumulative G-CSF dose of more than 10 000 µg/kg and a mean G-CSF dose per injection of more than 10 µg/kg are significantly related to malignant transformation [108,109]. The individual phenotype and the risk of malignancy vary for each genetic subtype of congenital neutropenia [112].

MAJOR PROGRESS

Qualitative neutrophil disorders such as myeloperoxidase deficiency (MPD), chronic granulomatous disease (CGD) and leukocyte adhesion deficiencies (LADs) are reviewed in great depth elsewhere. This review focuses on recent major progress in science on diseases associated with reduced numbers of neutrophil granulocytes [113[■]–115[■]].

BENIGN NEUTROPENIA

Benign neutropenia is a mild to moderate ($0.5\text{--}1.5 \times 10^9/l$) congenital neutropenia without numerical or morphological bone marrow abnormalities and without increased risk of bacterial or fungal infections [14,15[■],116,117]. Benign neutropenia is prevalent in approximately 4.5% of black and 0.8% of white Americans and can be found in the Arabian Peninsula and in various European regions as well [118–120].

In individuals from geographic regions where *Plasmodium spp.* are endemic, benign neutropenia is

associated with an autosomal recessive promotor polymorphism in the Duffy blood group chemokine receptor gene (*DARC* c.T-46>C) that abrogates the binding of the transcription factor GATA binding protein 1 (GATA1) and leads to a loss of DARC expression on erythrocytes [1–3]. Even though the resulting Duffy null blood group has been found protective against *Plasmodium vivax* infection and is associated with an increased susceptibility towards human immunodeficiency virus (HIV) infection, the precise molecular mechanism of benign neutropenia remains elusive [4–6,121[■]]. Current attempts to explain the phenotype of benign neutropenia often invoke nonhaematopoietic cells such as the endothelium and consecutive aberrant response to chemokines and transendothelial migration of neutrophil granulocytes [122,123].

Genome-wide association studies (GWAS) [124,125[■]] for benign neutropenia in the remaining ethnicities have identified a locus on chromosome 17q and further genetic regions.

ELANE DEFICIENCY

Heterozygous mutations in the elastase neutrophil expressed gene (*ELANE*) coding for the neutrophil elastase cause CyN and SCN type 1 (SCN1) in an autosomal dominant way and are the most common cause of congenital neutropenia in the Caucasian population (41% in Europeans and 55.6% in North Americans) [7,8]. More than 100 defined genetic variants in *ELANE* have been associated with SCN1 [126[■]].

CyN is characterized by an ANC that oscillates with a periodicity of approximately 21 days between almost normal values and complete disappearance of neutrophil granulocyte and that exhibits a reverse monocyte cycling, that is monocytes increase when neutrophil granulocytes decrease [9]. SCN1, however, displays a permanent bone marrow maturation arrest at the promyelocyte stage [7].

Even though specific *ELANE* mutations may lead to CyN and SCN1, respectively, there is no established firm genotype–phenotype correlation and we face a continuum rather than a clear-cut separation [7,8,10]. This might be due to inappropriate ANC monitoring, G-CSF treatment related cycle modification, incomplete mutation spectrum data, phenotype diversification by modifier genes and additional somatic mutations, especially in the colony stimulating factor 3 receptor gene (*CSF3R*) [11–14,15[■],127,128].

Nevertheless, on the basis of recent findings, several general conclusions may be justified. First, all known mutations lead to protein expression, so mere haploinsufficiency is not causing *ELANE*

Table 1. Genetic defects causing congenital neutropenia syndromes.

Disorder	Gene mutated	Inheritance	Haematopoietic manifestations	Extrahaematopoietic manifestations	Pathophysiology	Ref.
Benign neutropenia						
DARC polymorphism	DARC	AR	Mild to moderate CN, no severe infections, protection against <i>P. vivax</i> , susceptibility for HIV	-	In erythrocytes, abrogated GATA1 binding to DARC leads to LOE (Duffy null blood group), cause of CN unknown	[1-6]
Severe congenital neutropenia						
ELANE deficiency (CyN and SCN1)	ELANE	AD	CyN with 21 day periodicity, CN with permanent bone marrow maturation arrest, predominantly SCN1: MDS, AML	-	ELANE mistrafficking and/or UPR leads to autophagy and enhanced apoptosis of myeloid cells	[7-14,15*,16-20]
GFI1 deficiency (SCN2)	GFI1	AD	SCN and lymphopenia	-	Disturbed myeloid cell differentiation	[21-24]
HAX1 deficiency (SCN3)	HAX1	AR	CN, MDS, AML	Epilepsy, neurological impairment in some patients	Destabilization of mitochondrial membrane potential, enhanced apoptosis of myeloid and neuronal cells	[25-34]
G6PC3 deficiency (SCN4)	G6PC3	AR	CN, thrombocytopenia	Congenital heart and urogenital defects, failure to thrive, endocrinopathy, facial dysmorphism, inner ear hearing loss, hyperelastic skin, decreased subcutaneous fat tissue livedo reticularis	Impaired intracellular glucose homeostasis, dysglycosylation and UPR lead to enhanced apoptosis of myeloid cells	[35,36]
X-linked neutropenia (XLN, SCN-XL)	WAS	XL	CN, lymphopenia, MDS, AML	-	Disturbed actin polymerization, defective mitosis and cytokinesis	[37-41]
VPS45 deficiency (SCN5)	VPS45	AR	CN and neutrophil dysfunction, myelofibrosis, hypergammaglobulinaemia	Hepatosplenomegaly, nephromegaly (with extramedullary haematopoiesis)	Defective endosomal trafficking leads to impaired differentiation and motility and increased apoptosis of myeloid and mesenchymal cells	[42**]
Hypopigmentation disorders comprising congenital neutropenia						
Chédiak-Higashi syndrome	LYST	AR	CN, defective cytotoxicity, HLH, mild bleeding tendency, enlarged granules	Oculocutaneous albinism, neurodegeneration	Defective biogenesis of lysosomes, cytotoxic granules and melanosomes	[43-49]
Hermansky-Pudlak syndrome type 2	AP3B1	AR	CN, defective cytotoxicity, HLH, mild bleeding tendency, enlarged granules	Oculocutaneous albinism	Defective lysosomal protein sorting, for example of elastase and tyrosinase	[50-53,54*]
GrisCELLI syndrome type 2	RAB27A	AR	CN, defective cytotoxicity, HLH	Oculocutaneous albinism	Defective priming of cytotoxic granules and melanosomes	[55-58]
LAMTOR2 deficiency	LAMTOR2	AR	CN, defective cytotoxicity, lymphoid immunodeficiency	Oculocutaneous albinism, growth failure	Aberrant distribution of late endosomes	[59]

Complex disorders comprising congenital neutropenia					
AK2 deficiency	AK2	AR	CN and SCID, that is reticular dysgenesis	Inner ear hearing loss	Defective mitochondrial adenine nucleotide homeostasis, enhanced apoptosis [60,61]
STK4 deficiency	STK4	AR	CN, lymphopenia, CMC, warts	Congenital heart defects	Disturbed mitochondrial membrane potential, enhanced apoptosis [62,63,64]
Shwachman-Bodian-Diamond syndrome	SBDS	AR	CN, bone marrow failure, MDS, AML	Exocrine pancreatic insufficiency, skeletal dysplasia, hepatic and cardiac disorders	Mitotic spindle destabilization, genomic instability, enhanced apoptosis [65-68]
WHIM syndrome	CXCR4	AD	CN, warts, hypogammaglobulinaemia, infections, myelokathexis	-	Constitutively activated CXCR4 impairs chemokinesis of NG, DC and B cells [69-73]
Poikiloderma with neutropenia (Clericuzio-type)	USB1	AR	CN, bone marrow abnormalities	Poikiloderma, photosensitivity	Disturbed biogenesis of U6 snRNA, impaired cell viability [74-76,77]
Cartilage-hair hypoplasia	RMRP	AR	CN, variable cellular and humoral immunodeficiency, bone marrow failure	Short-limbed dwarfism, sparse hair, dyspermatogenesis, neuronal intestinal dysplasia, cancer susceptibility	Defective ribosome assembly, aberrant cell cycle control and telomere function [78-84]
CD40LG deficiency	CD40LG	XL	Intermittent CN, class-switch recombination deficiency, combined immunodeficiency	-	Abrogated CD40LG:CD40-signalling [85,86]
Barth syndrome	TAZ	XL	CN (in most patients)	Cardioskeletal myopathy, growth impairment, carnitine deficiency	Mitochondrial dysfunction, increased apoptosis in myeloid cells [87,88]
Cohen syndrome	VPS13B	AR	Intermittent CN	Chorioretinal dystrophy, postnatal microcephaly, myopathy	Altered vesicle sorting and transport [89-91]
GATA2 deficiency	GATA2	AD	CN, MDS, AML, MonoMAC syndrome	Sensory deafness, lymphoedema, pulmonary alveolar proteinosis	Complex ontogenic dysregulation of haematopoiesis and vascularization [92-95,96]
Glycogen storage disease Ib	SLC37A4	AR	CN, MDS, AML	Glycogen overload of the liver, hypoglycaemia, fasting hyperlactacidaemia	Impaired intracellular glucose homeostasis, probably as in G6PC3 deficiency [97-99]
Pearson syndrome	Deletion of mtDNA	MI	CN, bone marrow failure, vacuoles in erythroid precursors	Exocrine pancreas insufficiency, hepatopathy, nephropathy, endocrinopathy, neuromuscular degeneration	Variably sized mtDNA deletion, variable heteroplasmy, and mosaicism lead to metabolic disorder/energy failure and apoptosis in affected tissues [100-102]

AD, autosomal dominant; AML, acute myeloblastic leukaemia; AR, autosomal recessive; CMC, chronic mucocutaneous candidiasis; CN, congenital neutropenia; CyN, cyclic neutropenia; DC, dendritic cell; HLH, haemophagocytic lymphohistiocytosis; LOE, loss of expression; MDS, myelodysplastic syndrome; MI, maternal inheritance; mtDNA, mitochondrial deoxyribonucleic acid; NG, neutrophil granulocyte; SCID, severe combined immunodeficiency; SCN, severe congenital neutropenia; snRNA, small nuclear ribonucleic acid; UPR, unfolded protein response; XL, X-linked recessive; XIN, X-linked neutropenia. Adapted with permission from [103].

deficiency. Second, truncating mutations in the terminal exon 5 lead to expression of neutrophil elastase that lacks a binding site for the adaptor protein 3 (AP3) and are predominantly associated with SCN1. Third, missense mutations of the consensus splice acceptor in intron 4 lead to the use of a cryptic splice site in exon 5 and to the deletion of the catalytic site ($\Delta 161-170$) and are predominantly associated with CyN. Fourth, the common p.G815R mutation leads to G-CSF-refractory SCN1 with frequent progression to MDS and AML [15[¶],16,17].

The monomeric serine chymotryptic-type protease neutrophil elastase is a major component of neutrophil granulocyte azurophilic granules and is named after its first known substrate elastin [129]. Neutrophil elastase is synthesized as an inactive zymogen of 267 amino acid (aa) residues and is activated by limited proteolysis at both its amino and carboxy-terminus. The cleavage of the N-terminal 27 aa pre and two aa pro-signalling sequences and a C-terminal 20 aa propeptide is fundamental to neutrophil elastase trafficking and enzymatic function [130,131]. Neutrophil elastase targets a variety of bacterial virulence factors and processes cytokines, chemokines, G-CSF and G-CSFR. Thus, it is involved in bacterial killing and in various signalling and homeostatic circuits [132,133]. Activated neutrophil elastase is kept in check by serine proteinase inhibitors, that is serpins, such as $\alpha 1$ -protease inhibitor [134].

How exactly *ELANE* mutations lead to CyN and SCN1 remains a matter of debate and the three principal hypotheses that have emerged are not mutually exclusive. The inhibitor hypothesis assumes that *ELANE* mutations produce a negative feedback loop that leads to periodic impairment of neutrophil production and differentiation and uses mathematical models to demonstrate the 21-day periodicity of CyN [135].

The mislocalization hypothesis posits that neutrophil elastase is an AP3-cargo and trafficking through the *trans*-Golgi network towards the azurophilic granules is hampered as a consequence of C-terminal (truncating) mutations. Consequently, neutrophil elastase devoid of correct posttranslational processing accumulates in erroneous subcellular compartments [16,18,19]. How this interferes with (cyclic) neutrophil maturation arrest is not resolved and the answer might, at least partially, come from the unfolded protein response (UPR) hypothesis.

The UPR hypothesis proposes that *ELANE* mutations lead to neutrophil elastase misfolding during protein biosynthesis and cause an endoplasmic reticulum stress response as deduced by the upregulation of the stress marker heat shock 70-kDa protein 5 (HSPA5) and the splicing of

X-box-binding protein 1 (XBP1) mRNA. Endoplasmic reticulum stress then initiates autophagy or apoptosis of neutrophil granulocyte and causes SCN1 as illustrated in cell culture systems and transgenic murine models [19,20,136].

Even though it has become clear the *ELANE* mutation induced cell death is pivotal for the phenotype of congenital neutropenia, it is still unclear which factors determine CyN versus SCN1, why G-CSF treatment can overcome the observed maturation arrest and why MDS and AML preferentially develop in SCN1, but are scarce in CyN. It is noteworthy that leukaemogenesis in SCN1 can proceed as a consequence of somatic *CSF3R* mutations. The very mechanisms that deviate *ELANE* mutations towards CyN versus SCN1 may also control the threshold for selective pressure and additional somatic hits required for clonal outgrowth of leukaemia cells [137[¶]].

GFI1 DEFICIENCY (SCN2)

Very rarely, congenital neutropenia has been found to be associated with mutations in the transcription factor growth factor independent 1 transcription repressor (*GFI1*). A murine knockout model for the growth factor independent 1 gene (*Gfi1*) has been found to cause myeloid differentiation arrest and to lead to SCN as well [138,139]. In addition, the differentiation and function of further haematopoietic lineages such as dendritic cells, B cells and T helper cells are disturbed in *Gfi1*-deficient mice [140–143]. *Gfi1* is a predominantly repressive zinc-finger transcription factor that governs haematopoietic stem cell homeostasis and especially myeloid differentiation by regulating a plethora of biological processes such as transcriptional networks, microRNA processing and degradative ubiquitination-mediated apoptosis [144–147].

Almost paralleling these findings, heterozygous human mutations in *GFI1* leading to the dominant negative p.N382S and p.K403R substitutions and reducing the expression level of neutrophil elastase have been described to cause SCN2 with an additional lymphocyte proliferation defect [21,22]. Mechanistically, NEDD4-binding protein 2-like 2 (*N4BP2L2*) has been shown to interact with neutrophil elastase and *GFI1* and might link *GFI1* mutations to reduced *ELANE* expression in SCN2, even though there has not yet been any *N4BP2L2* mutation found in SCN [23].

Interestingly, the very same mutations have been found as monoallelic de-novo mutations in one patient presenting with CyN. The authors found cycling cytotoxic T cell responses against HLA-A2 presented neutrophil elastase epitopes as causing

this autoimmune CyN and their study [24] expands the pathogenetic mechanisms probably leading to CyN.

HAX1 DEFICIENCY (SCN 3)

Autosomal recessive mutations in haematopoietic cell specific LYN substrate 1 (HCLS1)-associated protein X-1 (*HAX1*) cause SCN3, classically known as Kostmann disease, and contribute to approximately 15% of SCN with a preponderance in the Turkish and Middle Eastern population [25,26,148].

HAX1 deficiency is characterized by SCN (often $<0.2 \times 10^9/l$) resulting from a maturation arrest at the promyelocyte stage and presenting during the first weeks of life with severe and life-threatening bacterial infections [27]. Some patients additionally display neurological impairment such as mental retardation and epilepsy and this is thought to result from *HAX1* mutations that do not only affect isoform A but also isoform B [28,29]. However, there might be a more complex genotype–phenotype relation, because compound heterozygous *HAX1* mutations targeting both transcripts have been described in SCN3 patients not affected by neuropathology [149,150^{***}].

HAX1 was initially identified as a ubiquitously expressed binding partner of HCLS1, which is involved in SRC-kinase signalling downstream of the B cell antigen receptor (BCR) [30]. HAX1 is an intracellular 35-kDa protein that predominantly localizes to the mitochondria but that can be detected in the endoplasmic reticulum and nuclear envelopes as well [30]. Biochemical analyses established a role of HAX1 in stabilizing the mitochondrial membrane potential ($\Delta\psi_m$) and were congruent with increased apoptosis observed in HAX1-deficient myeloid and extrahaematopoietic cells [26,31]. HAX1 interacts with presenilin-associated rhomboid-like (PARL) and HtrA serine peptidase 2 (HTRA2) that activate the pro-apoptotic BCL2-associated X protein (BAX) pathway [31,32]. In addition, HAX has been shown to protect the antiapoptotic X-linked inhibitor of apoptosis (XIAP) from ubiquitination-mediated proteasomal degradation [33]. Further evidence for an antiapoptotic role of HAX1 stems from the observation that it interacts with and down-regulates the protein level of ATPase Ca^{2+} transporting cardiac muscle slow twitch 2 (ATP2A2) that regulates the endoplasmic reticulum Ca^{2+} -concentration [34].

In line with an important function for HAX1 in cellular homeostasis is the observation that a number of viruses such as Epstein-Barr virus (EBV), human herpesvirus 8 (HHV-8), HIV and hepatitis C virus (HCV), which partially establish latency,

code for HAX-1 interacting proteins, even though the mechanistic consequences of these interactions remain elusive [151–156].

Additional HAX1–protein interactions that are coupled to particular biological processes have been elucidated and it seems that HAX1 is involved in Ras homolog family member A (RHOA) mediated neutrophil cell adhesion and migration as well as in posttranscriptional mRNA processing, stability and localization [157,158].

Taken as a whole, HAX1 seems to balance proapoptotic and antiapoptotic signalling networks and to modulate further biological processes, but the precise mechanisms of increased apoptosis in developing myeloid cells have still to be determined.

G6PC3 DEFICIENCY (SCN4)

Autosomal recessive mutations in glucose 6 phosphatase catalytic subunit 3 (*G6PC3*) cause SCN4, a predominantly quantitative disorder of neutrophil granulocytes with variable thrombocytopenia and associated syndromic features [159,160^{***}]. The most common extrahaematopoietic manifestations of G6PC3 deficiency comprise congenital heart and urogenital defects, failure to thrive, endocrine disorders, facial dysmorphism, inner ear hearing loss, hyperelastic skin, decreased subcutaneous fat tissue and livedo reticularis [160^{***}]. Mutations in *G6PC3* cluster in exons 1 and 6, which are coding for the C and N-terminus of G6PC3, respectively. However, the current mutational spectrum in a limited number of patients has not yet allowed a clear genotype–phenotype correlation [160^{***}].

G6PC3 is a ubiquitously expressed metabolic enzyme of the glucose 6 phosphatase (G6P) family, that is G6PC1–3, which localizes to the endoplasmic reticulum membrane where it is supposed to form a functional complex with glucose-6-phosphate transporter (G6PT), also known as solute carrier family 37 member 4 (SLC37A4) [161,162]. Neutrophil granulocyte G6PC3 deficiency deranges glucose recycling between the endoplasmic reticulum and the cytoplasm and leads to reduced levels of glucose uptake and its energy-carrying catabolites obtained by glycolysis [163]. In an experimental setting, this metabolic starving induces pronounced apoptosis of peripheral neutrophil granulocyte, whereas other haematopoietic lineages are less affected [159]. Mechanistically, a nutrient-sensitive network governed by the proapoptotic and antiapoptotic molecules glycogen synthase kinase 3 beta (GSK3B) and myeloid cell leukaemia sequence 1 (MCL1), respectively, in case of starvation targets the outer mitochondrial membrane and induces the intrinsic apoptotic pathway [164,165].

Importantly, defective endoplasmic reticulum glucose metabolism additionally interferes with proper glycosylation patterns of various proteins, such as components of the NADPH oxidase, which are important for neutrophil granulocyte functioning [35]. Furthermore, apoptosis in G6PC3-deficient myeloid cells might be linked to the UPR, but whether this is linked to dysglycosylation has not yet been examined [160^{***}].

In G6pc3-deficient murine neutrophil granulocytes, metabolic alterations and increased apoptosis can be counteracted by application of G-CSF [36]. It is tempting to speculate that the favourable clinical response towards G-CSF reported for G6PC3-deficient patients might follow similar correction mechanisms [160^{***}]. Currently, for G6PC3 deficiency, no malignant transformation has been reported, and in light of the particular pathogenesis and comorbidity, this might be an important feature of SCN4 and influence our therapeutic approach [160^{***}].

X-LINKED NEUTROPENIA

Mutations inactivating the Wiskott–Aldrich syndrome (*WAS*) gene product Wiskott–Aldrich syndrome protein (*WASP*) cause classical Wiskott–Aldrich syndrome (*WAS*) or X-linked thrombocytopenia (*XLT*). *WAS* and *XLT* determine a spectrum rather than two distinct nosological entities [166,167]. The former is characterized by lymphoid and myeloid immunodeficiency, autoimmunity, predisposition to malignant lymphoma, eczema and microthrombocytopenia, while the latter presents predominantly with microthrombocytopenia and bleeding diathesis [166,167].

WASP is central to actin dynamics and in the haematopoietic system couples the membrane-proximal signalling layer to the cytoskeletal layer [168]. Consequently, a variety of cellular processes such as immunological synapse and kinapse formation, TCR and BCR signalling, integrin inside-out signalling, vesicle trafficking and cellular morphogenesis are critically hampered [169]. An important means of *WASP* regulation is an autoinhibitory conformation that is based on an intramolecular interaction between the verprolin homology domain-cofilin homology domain-acidic region (*VCA*) and the GTPase-binding domain (*GBD*). This closed conformation interferes with binding of the actin-related protein 2/3 (*ARP2/3*) complex and with monomeric actin to the carboxyl terminus of *WASP* [169].

Contrasting to *WAS* and *XLT* wherein inactivating *WASP* mutations lead to absent or residual protein expression, activating *WASP* mutations that target the *GBD* and interfere with autoinhibition

cause X-linked neutropenia (*XLN*, *SCN-XL*) [37–39]. Constitutively active *WASP* results in intensified and misplaced actin polymerization and results in defective mitosis and cytokinesis as well as apoptosis-prone proliferation, especially in myeloid lineages [40]. Apart from neutropenia and monocytopenia, *XLN* shows reduced natural killer (*NK*) and T cell counts, impaired T cell proliferation and a genomic instability with a predisposition to *MDS* [38–41].

In summary, distinct *WAS* mutations lead to various disease entities and impressively highlight the multifaceted biological roles of *WASP* at the interface of the membrane-proximal signalling layer with the cytoskeletal layer in the haematopoietic system.

VPS45 DEFICIENCY (SCN-5)

Lately, autosomal recessive mutations in vacuolar protein sorting 45 homolog (*VPS45*) have been ascribed to a congenital neutrophil defect syndrome (*SCN5*) in seven patients from five consanguineous families [42^{***}].

VPS45 controls membrane trafficking through the endosomal system and defective endosomal trafficking and protein sorting/recycling appear to be the pathogenic mechanisms [42^{***},170,171].

Elegant phenotypic and functional analyses performed with *VPS45*-deficient primary neutrophil granulocyte and fibroblasts found quantitative and qualitative neutrophil impairments due to hampered development, decreased motility and increased apoptosis of myeloid and mesenchymal cells [42^{***}]. In addition, complementation assays reversed the phenotype in fibroblasts and a morpholino-injected zebrafish model recapitulated the *SCN* found in man [109].

Clinically, the patients presented *SCN*, thrombocytopenia, and anaemia, the latter probably due to an important myelofibrosis. In addition, extramedullary haematopoiesis leads to hepatosplenomegaly and nephromegaly [42^{***}]. All patients were refractory to G-CSF treatment, and in one of the affected kindred, additional neurodevelopmental sequelae were noted [42^{***}].

VPS45 deficiency highlights the importance of vesicle trafficking in the haematopoietic system, but especially the pathogenic mechanism of myelofibrosis awaits further clarification.

POIKILODERMA WITH NEUTROPENIA (CLERICUZIO-TYPE)

Mutations in the U6 snRNA biogenesis 1 (*USB1*) gene cause an autosomal recessive genodermatosis that

associates poikiloderma, that is erythematous rash, irregular skin pigmentation, teleangiectasia and hyperkeratosis, with neutropenia (PN), bone marrow and peripheral blood abnormalities [74,75]. The syndrome was first described by Clericuzio in Navajo Indians and consecutively found in Apache Indians and other kindred [74,76,172]. Infrequently, short stature, nail dystrophy and MDS have been reported as well [173].

USB1 is a phosphodiesterase and its yeast ortholog is involved as a RNA exonuclease in posttranslational processing of spliceosomal small nuclear RNA (snRNA) that is important for cell viability [77]. The clinical phenotype of PN overlaps with those of dyskeratosis congenita and Rothmund-Thomson syndrome (RTS), and even though in PN telomere shortage has not yet been described, this might be a common pathomechanism [173,174].

CONCLUSION

With the advent of joint databases, commonly available G-CSF treatment and advanced genetic and biochemical techniques, our knowledge of congenital neutropenia syndromes is constantly growing. Novel genetic entities pinpoint towards fundamental biological processes important for the homeostasis and functioning not only of the neutrophil granulocyte but also of the haematopoietic system and its niche. Even though not all aspects of molecular pathogenesis have been elucidated up to now, treatment decisions, that is G-CSF application versus haematopoietic stem cell transplantation, become more sophisticated and might open up into personalized molecular medicine.

Acknowledgements

We value the personal, clinical and scientific contributions of the patients, their families and our colleagues. We acknowledge the SCN registries in Germany and France and the international Care-for-Rare Alliance. This work was partially supported by the Erare NeutroNET grant.

Conflicts of interest

There are no conflicts of interest.

REFERENCES AND RECOMMENDED READING

Papers of particular interest, published within the annual period of review, have been highlighted as:

- of special interest
- of outstanding interest

1. Peiper SC, Wang ZX, Neote K, *et al.* The Duffy antigen/receptor for chemokines (DARC) is expressed in endothelial cells of Duffy negative individuals who lack the erythrocyte receptor. *J Exp Med* 1995; 181:1311–1317.

2. Reich D, Nalls MA, Kao WH, *et al.* Reduced neutrophil count in people of African descent is due to a regulatory variant in the Duffy antigen receptor for chemokines gene. *PLoS Genet* 2009; 5:e1000360.
 3. Tournamille C, Colin Y, Cartron JP, Le Van Kim C. Disruption of a GATA motif in the Duffy gene promoter abolishes erythroid gene expression in Duffy-negative individuals. *Nat Genet* 1995; 10:224–228.
 4. Horuk R, Chitnis CE, Darbonne WC, *et al.* A receptor for the malarial parasite *Plasmodium vivax*: the erythrocyte chemokine receptor. *Science* 1993; 261:1182–1184.
 5. Miller LH, Mason SJ, Clyde DF, McGinniss MH. The resistance factor to *Plasmodium vivax* in blacks: the Duffy-blood-group genotype, FyFy. *N Engl J Med* 1976; 295:302–304.
 6. Ramsuran V, Kulkarni H, He W, *et al.* Duffy-null-associated low neutrophil counts influence HIV-1 susceptibility in high-risk South African black women. *Clin Infect Dis* 2011; 52:1248–1256.
 7. Dale DC, Person RE, Bolyard AA, *et al.* Mutations in the gene encoding neutrophil elastase in congenital and cyclic neutropenia. *Blood* 2000; 96:2317–2322.
 8. Horwitz M, Benson KF, Person RE, *et al.* Mutations in ELA2, encoding neutrophil elastase, define a 21-day biological clock in cyclic haematopoiesis. *Nat Genet* 1999; 23:433–436.
 9. Lange RD. Cyclic hematopoiesis: human cyclic neutropenia. *Exp Hematol* 1983; 11:435–451.
 10. Thusberg J, Vihinen M. Bioinformatic analysis of protein structure-function relationships: case study of leukocyte elastase (ELA2) missense mutations. *Hum Mutat* 2006; 27:1230–1243.
 11. Germeshausen M, Zeidler C, Stuhmann M, *et al.* Digenic mutations in severe congenital neutropenia. *Haematologica* 2010; 95:1207–1210.
 12. Lanciotti M, Caridi G, Rosano C, *et al.* Severe congenital neutropenia: a negative synergistic effect of multiple mutations of ELANE (ELA2) gene. *Br J Haematol* 2009; 146:578–580.
 13. Newburger PE, Pindyck TN, Zhu Z, *et al.* Cyclic neutropenia and severe congenital neutropenia in patients with a shared ELANE mutation and paternal haplotype: evidence for phenotype determination by modifying genes. *Pediatr Blood Cancer* 2010; 55:314–317.
 14. Salpante SJ, Benson KF, Luty J, *et al.* Double de novo mutations of ELA2 in cyclic and severe congenital neutropenia. *Hum Mutat* 2007; 28:874–881.
 15. Horwitz MS, Corey SJ, Grimes HL, Tidwell T. ELANE mutations in cyclic and severe congenital neutropenia: genetics and pathophysiology. *Hematol Oncol Clin North Am* 2013; 27:19–41; vii.
- This study comprehensively reviews SCN due to ELANE deficiency.
16. Horwitz MS, Duan Z, Korkmaz B, *et al.* Neutrophil elastase in cyclic and severe congenital neutropenia. *Blood* 2007; 109:1817–1824.
 17. Massullo P, Druhan LJ, Bunnell BA, *et al.* Aberrant subcellular targeting of the G185R neutrophil elastase mutant associated with severe congenital neutropenia induces premature apoptosis of differentiating promyelocytes. *Blood* 2005; 105:3397–3404.
 18. Li W, Feng Y, Hao C, *et al.* The BLOC interactomes form a network in endosomal transport. *J Genet Genomics* 2007; 34:669–682.
 19. Kollner I, Sodeik B, Schreek S, *et al.* Mutations in neutrophil elastase causing congenital neutropenia lead to cytoplasmic protein accumulation and induction of the unfolded protein response. *Blood* 2006; 108:493–500.
 20. Grenda DS, Murakami M, Ghatak J, *et al.* Mutations of the ELA2 gene found in patients with severe congenital neutropenia induce the unfolded protein response and cellular apoptosis. *Blood* 2007; 110:4179–4187.
 21. Person RE, Li FQ, Duan Z, *et al.* Mutations in proto-oncogene GFI1 cause human neutropenia and target ELA2. *Nat Genet* 2003; 34:308–312.
 22. Zarebski A, Velu CS, Baktula AM, *et al.* Mutations in growth factor independent-1 associated with human neutropenia block murine granulopoiesis through colony stimulating factor-1. *Immunity* 2008; 28:370–380.
 23. Salpante SJ, Rojas ME, Korkmaz B, *et al.* Contributions to neutropenia from PFAAP5 (N4BP2L2), a novel protein mediating transcriptional repressor cooperation between Gfi1 and neutrophil elastase. *Mol Cell Biol* 2009; 29:4394–4405.
 24. Armistead PM, Wiedner E, Akande O, *et al.* Cyclic neutropenia associated with T cell immunity to granulocyte proteases and a double de novo mutation in GFI1, a transcriptional regulator of ELANE. *Br J Haematol* 2010; 150:716–719.
 25. Kostmann R. Infantile genetic agranulocytosis; agranulocytosis infantilis hereditaria. *Acta Paediatr Suppl* 1956; 45:1–78.
 26. Klein C, Grudzien M, Appaswamy G, *et al.* HAX1 deficiency causes autosomal recessive severe congenital neutropenia (Kostmann disease). *Nat Genet* 2007; 39:86–92.
 27. Carlsson G, Fasth A. Infantile genetic agranulocytosis, morbus Kostmann: presentation of six cases from the original 'Kostmann family' and a review. *Acta Paediatr* 2001; 90:757–764.
 28. Germeshausen M, Grudzien M, Zeidler C, *et al.* Novel HAX1 mutations in patients with severe congenital neutropenia reveal isoform-dependent genotype-phenotype associations. *Blood* 2008; 111:4954–4957.
 29. Carlsson G, van't Hooft I, Melin M, *et al.* Central nervous system involvement in severe congenital neutropenia: neurological and neuropsychological abnormalities associated with specific HAX1 mutations. *J Intern Med* 2008; 264:388–400.

30. Suzuki Y, Demoliere C, Kitamura D, *et al*. HAX-1, a novel intracellular protein, localized on mitochondria, directly associates with HS1, a substrate of Src family tyrosine kinases. *J Immunol* 1997; 158:2736–2744.
31. Chao JR, Parganas E, Boyd K, *et al*. Hax1-mediated processing of HtrA2 by Parl allows survival of lymphocytes and neurons. *Nature* 2008; 452:98–102.
32. Cilenti L, Soundarapandian MM, Kyriazis GA, *et al*. Regulation of HAX-1 antiapoptotic protein by Omi/HtrA2 protease during cell death. *J Biol Chem* 2004; 279:50295–50301.
33. Kang YJ, Jang M, Park YK, *et al*. Molecular interaction between HAX-1 and XIAP inhibits apoptosis. *Biochem Biophys Res Commun* 2010; 393:794–799.
34. Vafiadaki E, Arvanitis DA, Pagakis SN, *et al*. The antiapoptotic protein HAX-1 interacts with SERCA2 and regulates its protein levels to promote cell survival. *Mol Biol Cell* 2009; 20:306–318.
35. Hayee B, Antonopoulos A, Murphy EJ, *et al*. G6PC3 mutations are associated with a major defect of glycosylation: a novel mechanism for neutrophil dysfunction. *Glycobiology* 2011; 21:914–924.
36. Jun HS, Lee YM, Song KD, *et al*. G-CSF improves murine G6PC3-deficient neutrophil function by modulating apoptosis and energy homeostasis. *Blood* 2011; 117:3881–3892.
37. Devriendt K, Kim AS, Mathijs G, *et al*. Constitutively activating mutation in WASP causes X-linked severe congenital neutropenia. *Nat Genet* 2001; 27:313–317.
38. Ancliff PJ, Blundell MP, Cory GO, *et al*. Two novel activating mutations in the Wiskott-Aldrich syndrome protein result in congenital neutropenia. *Blood* 2006; 108:2182–2189.
39. Beel K, Cotter MM, Blatny J, *et al*. A large kindred with X-linked neutropenia with an I294T mutation of the Wiskott-Aldrich syndrome gene. *Br J Haematol* 2009; 144:120–126.
40. Moulding DA, Blundell MP, Spiller DG, *et al*. Unregulated actin polymerization by WASp causes defects of mitosis and cytokinesis in X-linked neutropenia. *J Exp Med* 2007; 204:2213–2224.
41. Westerberg LS, Meelu P, Baptista M, *et al*. Activating WASP mutations associated with X-linked neutropenia result in enhanced actin polymerization, altered cytoskeletal responses, and genomic instability in lymphocytes. *J Exp Med* 2010; 207:1145–1152.
42. Vilboux T, Lev A, Malicdan MC, *et al*. A congenital neutrophil defect syndrome associated with mutations in VPS45. *N Engl J Med* 2013; 369:54–65. This study describes VPS45 deficiency and defective endosomal trafficking as a novel pathogenic mechanism of SCN.
43. Chediak MM. New leukocyte anomaly of constitutional and familial character. *Rev Hematol* 1952; 7:362–367.
44. Higashi O. Congenital gigantism of peroxidase granules; the first case ever reported of qualitative abnormality of peroxidase. *Tohoku J Exp Med* 1954; 59:315–332.
45. Barbosa MD, Nguyen QA, Tchernev VT, *et al*. Identification of the homologous beige and Chediak-Higashi syndrome genes. *Nature* 1996; 382:262–265.
46. Nagle DL, Karim MA, Woolf EA, *et al*. Identification and mutation analysis of the complete gene for Chediak-Higashi syndrome. *Nat Genet* 1996; 14:307–311.
47. Bossi G, Griffiths GM. CTL secretory lysosomes: biogenesis and secretion of a harmful organelle. *Semin Immunol* 2005; 17:87–94.
48. Williams RL, Urbe S. The emerging shape of the ESCRT machinery. *Nat Rev Mol Cell Biol* 2007; 8:355–368.
49. Karim MA, Suzuki K, Fukai K, *et al*. Apparent genotype-phenotype correlation in childhood, adolescent, and adult Chediak-Higashi syndrome. *Am J Med Genet* 2002; 108:16–22.
50. Hermansky F, Pudlak P. Albinism associated with hemorrhagic diathesis and unusual pigmented reticular cells in the bone marrow: report of two cases with histochemical studies. *Blood* 1959; 14:162–169.
51. Wei ML. Hermansky-Pudlak syndrome: a disease of protein trafficking and organelle function. *Pigment Cell Res* 2006; 19:19–42.
52. Clark RH, Stinchcombe JC, Day A, *et al*. Adaptor protein 3-dependent microtubule-mediated movement of lytic granules to the immunological synapse. *Nat Immunol* 2003; 4:1111–1120.
53. Enders A, Zieger B, Schwarz K, *et al*. Lethal hemophagocytic lymphohistiocytosis in Hermansky-Pudlak syndrome type II. *Blood* 2006; 108:81–87.
54. Jessen B, Bode SF, Ammann S, *et al*. The risk of hemophagocytic lymphohistiocytosis in Hermansky-Pudlak syndrome type 2. *Blood* 2013; 121:2943–2951. This study establishes the relative risk of HLH in HPS2 and proposes that preemptive HSCTx is not justified for this entity.
55. Klein C, Philippe N, Le Deist F, *et al*. Partial albinism with immunodeficiency (Griscelli syndrome). *J Pediatr* 1994; 125:886–895.
56. Meeths M, Bryceson YT, Rudd E, *et al*. Clinical presentation of Griscelli syndrome type 2 and spectrum of RAB27A mutations. *Pediatr Blood Cancer* 2010; 54:563–572.
57. Van Gele M, Dynoodt P, Lambert J. Griscelli syndrome: a model system to study vesicular trafficking. *Pigment Cell Melanoma Res* 2009; 22:268–282.
58. Ohbayashi N, Mamishi S, Ishibashi K, *et al*. Functional characterization of two RAB27A missense mutations found in Griscelli syndrome type 2. *Pigment Cell Melanoma Res* 2010; 23:365–374.
59. Bohn G, Allroth A, Brandes G, *et al*. A novel human primary immunodeficiency syndrome caused by deficiency of the endosomal adaptor protein p14. *Nat Med* 2007; 13:38–45.
60. Lagresle-Peyrou C, Six EM, Picard C, *et al*. Human adenylate kinase 2 deficiency causes a profound hematopoietic defect associated with sensorineural deafness. *Nat Genet* 2009; 41:106–111.
61. Pannicke U, Honig M, Hess I, *et al*. Reticular dysgenesis (aleukocytosis) is caused by mutations in the gene encoding mitochondrial adenylate kinase 2. *Nat Genet* 2009; 41:101–105.
62. Abdollahpour H, Appaswamy G, Kotlarz D, *et al*. The phenotype of human STK4 deficiency. *Blood* 2012; 119:3450–3457. This study together with ref. [140] describes STK4 deficiency.
63. Crequer A, Picard C, Patin E, *et al*. Inherited MST1 deficiency underlies susceptibility to EV-HPV infections. *PLoS One* 2012; 7:e44010. This study expands the phenotype of STK4 deficiency.
64. Nehme NT, Pachlopnik Schmid J, Debeurme F, *et al*. MST1 mutations in autosomal recessive primary immunodeficiency characterized by defective naive T-cell survival. *Blood* 2012; 119:3458–3468. This study together with ref. [138] describes STK4 deficiency.
65. Boocock GR, Morrison JA, Popovic M, *et al*. Mutations in SBDS are associated with Shwachman-Diamond syndrome. *Nat Genet* 2003; 33:97–101.
66. Austin KM, Gupta ML Jr, Coats SA, *et al*. Mitotic spindle destabilization and genomic instability in Shwachman-Diamond syndrome. *J Clin Invest* 2008; 118:1511–1518.
67. Raaijmakers MH, Mukherjee S, Guo S, *et al*. Bone progenitor dysfunction induces myelodysplasia and secondary leukaemia. *Nature* 2010; 464:852–857.
68. Dror Y, Donadieu J, Koglmeyer J, *et al*. Draft consensus guidelines for diagnosis and treatment of Shwachman-Diamond syndrome. *Ann N Y Acad Sci* 2011; 1242:40–55.
69. Zuelzer WW. Myelokathexis: a new form of chronic granulocytopenia. Report of a case. *N Engl J Med* 1964; 270:699–704.
70. Hernandez PA, Gorlin RJ, Lukens JN, *et al*. Mutations in the chemokine receptor gene CXCR4 are associated with WHIM syndrome, a combined immunodeficiency disease. *Nat Genet* 2003; 34:70–74.
71. Balabanian K, Lagane B, Pablos JL, *et al*. WHIM syndromes with different genetic anomalies are accounted for by impaired CXCR4 desensitization to CXCL12. *Blood* 2005; 105:2449–2457.
72. Lagane B, Chow KY, Balabanian K, *et al*. CXCR4 dimerization and beta-arrestin-mediated signaling account for the enhanced chemotaxis to CXCL12 in WHIM syndrome. *Blood* 2008; 112:34–44.
73. McDermott DH, Liu Q, Ulrick J, *et al*. The CXCR4 antagonist plerixafor corrects pancytopenia in patients with WHIM syndrome. *Blood* 2011; 118:4957–4962.
74. Clericuzio C, Hoyme H, Aase J. Immune deficient poikiloderma: a new genderatoma. *Am J Hum Genet* 1991; 86(1):A661.
75. Volpi L, Roversi G, Colombo EA, *et al*. Targeted next-generation sequencing appoints c16orf57 as clericuzio-type poikiloderma with neutropenia gene. *Am J Hum Genet* 2010; 86:72–76.
76. Arnold AW, Itin PH, Pigors M, *et al*. Poikiloderma with neutropenia: a novel C16orf57 mutation and clinical diagnostic criteria. *Br J Dermatol* 2010; 163:866–869.
77. Mroczek S, Krwawicz J, Kutner J, *et al*. C16orf57, a gene mutated in poikiloderma with neutropenia, encodes a putative phosphodiesterase responsible for the U6 snRNA 3' end modification. *Genes Dev* 2012; 26:1911–1925. This study describes the *USB1* gene product as a putative phosphodiesterase involved in snRNA stability and proposes telomere maintenance as a common pathogenic mechanism of PN, RTS and dyskeratosis congenita.
78. McKusick VA, Eldridge R, Hostetler JA, *et al*. Dwarfism in the Amish. II. Cartilage-hair hypoplasia. *Bull Johns Hopkins Hosp* 1965; 116:285–326.
79. Makitie O. Cartilage-hair hypoplasia in Finland: epidemiological and genetic aspects of 107 patients. *J Med Genet* 1992; 29:652–655.
80. Ridanpaa M, van Eenennaam H, Pelin K, *et al*. Mutations in the RNA component of RNase MRP cause a pleiotropic human disease, cartilage-hair hypoplasia. *Cell* 2001; 104:195–203.
81. Makitie O, Juvonen E, Dunkel L, *et al*. Anemia in children with cartilage-hair hypoplasia is related to body growth and to the insulin-like growth factor system. *J Clin Endocrinol Metab* 2000; 85:563–568.
82. Makitie O, Pukkala E, Teppo L, Kaitila I. Increased incidence of cancer in patients with cartilage-hair hypoplasia. *J Pediatr* 1999; 134:315–318.
83. Bordon V, Gennery AR, Slatter MA, *et al*. Clinical and immunologic outcome of patients with cartilage hair hypoplasia after hematopoietic stem cell transplantation. *Blood* 2010; 116:27–35.
84. de la Fuente MA, Recher M, Rider NL, *et al*. Reduced thymic output, cell cycle abnormalities, and increased apoptosis of T lymphocytes in patients with cartilage-hair hypoplasia. *J Allergy Clin Immunol* 2011; 128:139–146.
85. Winkelstein JA, Marino MC, Ochs H, *et al*. The X-linked hyper-IgM syndrome: clinical and immunologic features of 79 patients. *Medicine (Baltimore)* 2003; 82:373–384.
86. Kracker S, Gardes P, Mazerolles F, Durandy A. Immunoglobulin class switch recombination deficiencies. *Clin Immunol* 2010; 135:193–203.

87. Barth PG, Scholte HR, Berden JA, *et al.* An X-linked mitochondrial disease affecting cardiac muscle, skeletal muscle and neutrophil leucocytes. *J Neurol Sci* 1983; 62:327–355.
88. Barth PG, Valianpour F, Bowen VM, *et al.* X-linked cardioskeletal myopathy and neutropenia (Barth syndrome): an update. *Am J Med Genet A* 2004; 126A:349–354.
89. Kolehmainen J, Black GC, Saarinen A, *et al.* Cohen syndrome is caused by mutations in a novel gene, COH1, encoding a transmembrane protein with a presumed role in vesicle-mediated sorting and intracellular protein transport. *Am J Hum Genet* 2003; 72:1359–1369.
90. Chandler KE, Kidd A, Al-Gazali L, *et al.* Diagnostic criteria, clinical characteristics, and natural history of Cohen syndrome. *J Med Genet* 2003; 40:233–241.
91. El Chehadeh S, Aral B, Gigot N, *et al.* Search for the best indicators for the presence of a VPS13B gene mutation and confirmation of diagnostic criteria in a series of 34 patients genotyped for suspected Cohen syndrome. *J Med Genet* 2010; 47:549–553.
92. Hahn CN, Chong CE, Carmichael CL, *et al.* Heritable GATA2 mutations associated with familial myelodysplastic syndrome and acute myeloid leukemia. *Nat Genet* 2011; 43:1012–1017.
93. Ostergaard P, Simpson MA, Connell FC, *et al.* Mutations in GATA2 cause primary lymphedema associated with a predisposition to acute myeloid leukemia (Emberger syndrome). *Nat Genet* 2011; 43:929–931.
94. Hsu AP, Sampaio EP, Khan J, *et al.* Mutations in GATA2 are associated with the autosomal dominant and sporadic monocytopenia and mycobacterial infection (MonoMAC) syndrome. *Blood* 2011; 118:2653–2655.
95. Emberger JM, Navarro M, Dejean M, Izarn P. Deaf-mutism, lymphedema of the lower limbs and hematological abnormalities (acute leukemia, cytopenia) with autosomal dominant transmission. *J Genet Hum* 1979; 27:237–245.
96. Pasquet M, Bellanne-Chantelot C, Tavittian S, *et al.* High frequency of GATA2 mutations in patients with mild chronic neutropenia evolving to MonoMac syndrome, myelodysplasia, and acute myeloid leukemia. *Blood* 2013; 121:822–829.
- This study describes GATA2 deficiency as the cause of MonoMac syndrome and congenital MDS.
97. Beaudet AL, Anderson DC, Michels VV, *et al.* Neutropenia and impaired neutrophil migration in type IB glycogen storage disease. *J Pediatr* 1980; 97:906–910.
98. Gerin I, Veiga-da-Cunha M, Achouri Y, *et al.* Sequence of a putative glucose 6-phosphate translocase, mutated in glycogen storage disease type Ib. *FEBS Lett* 1997; 419:235–238.
99. Narisawa K, Tada K, Kuzuya T. Neutropenia in type IB glycogen storage disease. *J Pediatr* 1981; 99:334–335.
100. Pearson HA, Lobel JS, Kocoshis SA, *et al.* A new syndrome of refractory sideroblastic anemia with vacuolization of marrow precursors and exocrine pancreatic dysfunction. *J Pediatr* 1979; 95:976–984.
101. Seneca S, De Meirleir L, De Schepper J, *et al.* Pearson marrow pancreas syndrome: a molecular study and clinical management. *Clin Genet* 1997; 51:338–342.
102. Wong LJ. Molecular genetics of mitochondrial disorders. *Dev Disabil Res Rev* 2010; 16:154–162.
103. Boztug K, Klein C. Genetics and pathophysiology of severe congenital neutropenia syndromes unrelated to neutrophil elastase. *Hematol Oncol Clin North Am* 2013; 27:43–60; vii.
- This study comprehensively reviews SCN other than ELANE deficiency.
104. Skokowa J, Welte K. Dysregulation of myeloid-specific transcription factors in congenital neutropenia. *Ann N Y Acad Sci* 2009; 1176:94–100.
105. Manroe BL, Weinberg AG, Rosenfeld CR, Browne R. The neonatal blood count in health and disease. I. Reference values for neutrophilic cells. *J Pediatr* 1979; 95:89–98.
106. Sennels HP, Jorgensen HL, Hansen AL, *et al.* Diurnal variation of hematology parameters in healthy young males: the Bispebjerg study of diurnal variations. *Scand J Clin Lab Invest* 2011; 71:532–541.
107. Donadiou J, Beaupain B, Mahlaoui N, Bellanne-Chantelot C. Epidemiology of congenital neutropenia. *Hematol Oncol Clin North Am* 2013; 27:1–17; vii.
- This study comprehensively reviews epidemiologic aspects of congenital neutropenia.
108. Donadiou J, Leblanc T, Bader Meunier B, *et al.* Analysis of risk factors for myelodysplasias, leukemias and death from infection among patients with congenital neutropenia. Experience of the French Severe Chronic Neutropenia Study Group. *Haematologica* 2005; 90:45–53.
109. Rosenberg PS, Alter BP, Bolyard AA, *et al.* The incidence of leukemia and mortality from sepsis in patients with severe congenital neutropenia receiving long-term G-CSF therapy. *Blood* 2006; 107:4628–4635.
110. Bonilla MA, Gillio AP, Ruggiero M, *et al.* Effects of recombinant human granulocyte colony-stimulating factor on neutropenia in patients with congenital agranulocytosis. *N Engl J Med* 1989; 320:1574–1580.
111. Dale DC, Bonilla MA, Davis MW, *et al.* A randomized controlled phase III trial of recombinant human granulocyte colony-stimulating factor (filgrastim) for treatment of severe chronic neutropenia. *Blood* 1993; 81:2496–2502.
112. Klein C. Genetic defects in severe congenital neutropenia: emerging insights into life and death of human neutrophil granulocytes. *Annu Rev Immunol* 2011; 29:399–413.
113. Klebanoff SJ, Kettle AJ, Rosen H, *et al.* Myeloperoxidase: a front-line defender against phagocytosed microorganisms. *J Leukoc Biol* 2013; 93:185–198.
- This study comprehensively reviews MPD.
114. Holland SM. Chronic granulomatous disease. *Hematol Oncol Clin North Am* 2013; 27:89–99; viii.
- This study comprehensively reviews CGD.
115. van de Vijver E, van den Berg TK, Kuijpers TW. Leukocyte adhesion deficiencies. *Hematol Oncol Clin North Am* 2013; 27:101–116; viii.
- This study comprehensively reviews LAD.
116. Broun GO Jr, Herbig FK, Hamilton JR. Leukopenia in Negroes. *N Engl J Med* 1966; 275:1410–1413.
117. Haddy TB, Rana SR, Castro O. Benign ethnic neutropenia: what is a normal absolute neutrophil count? *J Lab Clin Med* 1999; 133:15–22.
118. Hsieh MM, Everhart JE, Byrd-Holt DD, *et al.* Prevalence of neutropenia in the U.S. population: age, sex, smoking status, and ethnic differences. *Ann Intern Med* 2007; 146:486–492.
119. Denic S, Showqj S, Klein C, *et al.* Prevalence, phenotype and inheritance of benign neutropenia in Arabs. *BMC Blood Disord* 2009; 9:3.
120. Papadaki HA, Xylouri I, Coulocheri S, *et al.* Prevalence of chronic idiopathic neutropenia of adults among an apparently healthy population living on the island of Crete. *Ann Hematol* 1999; 78:293–297.
121. Kourtit AP, Hudgens MG, Kayira D. Neutrophil count in African mothers and newborns and HIV transmission risk. *N Engl J Med* 2012; 367:2260–2262.
- This study establishes a relationship between the Duffy null trait and an increased susceptibility to perinatal HIV infection.
122. Lee JS, Frevert CW, Wurfel MM, *et al.* Duffy antigen facilitates movement of chemokine across the endothelium in vitro and promotes neutrophil transmigration in vitro and in vivo. *J Immunol* 2003; 170:5244–5251.
123. Lee JS, Wurfel MM, Matute-Bello G, *et al.* The Duffy antigen modifies systemic and local tissue chemokine responses following lipopolysaccharide stimulation. *J Immunol* 2006; 177:8086–8094.
124. Nalls MA, Couper DJ, Tanaka T, *et al.* Multiple loci are associated with white blood cell phenotypes. *PLoS Genet* 2011; 7:e1002113.
125. Crosslin DR, McDavid A, Weston N, *et al.* Genetic variants associated with the white blood cell count in 13,923 subjects in the eMERGE Network. *Hum Genet* 2012; 131:639–652.
- Together with ref. [28], this article establishes the association of particular genetic loci with total white blood cell counts and subtype counts.
126. Germeshausen M, Deerberg S, Peter Y, *et al.* The spectrum of ELANE mutations and their implications in severe congenital and cyclic neutropenia. *Hum Mutat* 2013; 34:905–914.
- This study shows that there is only a partial genotype–phenotype correlation for particular ELANE mutations with SCN or CyN and that the risk of leukemogenesis correlates more with disease severity than with a specific mutation.
127. Dong F, Brynes RK, Tidow N, *et al.* Mutations in the gene for the granulocyte colony-stimulating-factor receptor in patients with acute myeloid leukemia preceded by severe congenital neutropenia. *N Engl J Med* 1995; 333:487–493.
128. Carlsson G, Aprikyan AA, Ericson KG, *et al.* Neutrophil elastase and granulocyte colony-stimulating factor receptor mutation analyses and leukemia evolution in severe congenital neutropenia patients belonging to the original Kostmann family in northern Sweden. *Haematologica* 2006; 91:589–595.
129. Janoff A, Scherer J. Mediators of inflammation in leukocyte lysosomes. IX. Elastolytic activity in granules of human polymorphonuclear leukocytes. *J Exp Med* 1968; 128:1137–1155.
130. Gullberg U, Bengtsson N, Bulow E, *et al.* Processing and targeting of granule proteins in human neutrophils. *J Immunol Methods* 1999; 232:201–210.
131. Adkison AM, Raptis SZ, Kelley DG, Pham CT. Dipeptidyl peptidase I activates neutrophil-derived serine proteases and regulates the development of acute experimental arthritis. *J Clin Invest* 2002; 109:363–371.
132. Korkmaz B, Horwitz MS, Jenne DE, Gauthier F. Neutrophil elastase, proteinase 3, and cathepsin G as therapeutic targets in human diseases. *Pharmacol Rev* 2010; 62:726–759.
133. Hunter MG, Druhan LJ, Massullo PR, Avalos BR. Proteolytic cleavage of granulocyte colony-stimulating factor and its receptor by neutrophil elastase induces growth inhibition and decreased cell surface expression of the granulocyte colony-stimulating factor receptor. *Am J Hematol* 2003; 74:149–155.
134. Ye S, Goldsmith EJ. Serpins and other covalent protease inhibitors. *Curr Opin Struct Biol* 2001; 11:740–745.
135. Horwitz M, Benson KF, Duan Z, *et al.* Role of neutrophil elastase in bone marrow failure syndromes: molecular genetic revival of the chalone hypothesis. *Curr Opin Hematol* 2003; 10:49–54.
136. Nana S, Murakami M, Xia J, *et al.* Activation of the unfolded protein response is associated with impaired granulopoiesis in transgenic mice expressing mutant Elane. *Blood* 2011; 117:3539–3547.
137. Beekman R, Valkhof MG, Sanders MA, *et al.* Sequential gain of mutations in severe congenital neutropenia progressing to acute myeloid leukemia. *Blood* 2012; 119:5071–5077.
- This study shows that leukemogenesis in SCN is a multistep process with abnormal G-CSF signalling as an important driver of leukemic transformation.

138. Hock H, Hamblen MJ, Rooke HM, *et al.* Intrinsic requirement for zinc finger transcription factor Gfi-1 in neutrophil differentiation. *Immunity* 2003; 18:109–120.
139. Karsunky H, Zeng H, Schmidt T, *et al.* Inflammatory reactions and severe neutropenia in mice lacking the transcriptional repressor Gfi1. *Nat Genet* 2002; 30:295–300.
140. Rathinam C, Geffers R, Yucel R, *et al.* The transcriptional repressor Gfi1 controls STAT3-dependent dendritic cell development and function. *Immunity* 2005; 22:717–728.
141. Rathinam C, Klein C. Transcriptional repressor Gfi1 integrates cytokine-receptor signals controlling B-cell differentiation. *PLoS One* 2007; 2:e306.
142. Zhu J, Guo L, Min B, *et al.* Growth factor independent-1 induced by IL-4 regulates Th2 cell proliferation. *Immunity* 2002; 16:733–744.
143. Zhu J, Davidson TS, Wei G, *et al.* Down-regulation of Gfi-1 expression by TGF-beta is important for differentiation of Th17 and CD103+ inducible regulatory T cells. *J Exp Med* 2009; 206:329–341.
144. Horman SR, Velu CS, Chaubey A, *et al.* Gfi1 integrates progenitor versus granulocytic transcriptional programming. *Blood* 2009; 113:5466–5475.
145. Zhuang D, Qiu Y, Kogan SC, Dong F. Increased CCAAT enhancer-binding protein epsilon (C/EBPepsilon) expression and premature apoptosis in myeloid cells expressing Gfi-1 N382S mutant associated with severe congenital neutropenia. *J Biol Chem* 2006; 281:10745–10751.
146. Velu CS, Baktula AM, Grimes HL. Gfi1 regulates miR-21 and miR-196b to control myelopoiesis. *Blood* 2009; 113:4720–4728.
147. Martejn JA, van der Meer LT, van Emst L, *et al.* Gfi1 ubiquitination and proteasomal degradation is inhibited by the ubiquitin ligase Triad1. *Blood* 2007; 110:3128–3135.
148. Xia J, Bolyard AA, Rodger E, *et al.* Prevalence of mutations in ELANE, GFI1, HAX1, SBDS, WAS and G6PC3 in patients with severe congenital neutropenia. *Br J Haematol* 2009; 147:535–542.
149. Carlsson G, Elinder G, Malmgren H, *et al.* Compound heterozygous HAX1 mutations in a Swedish patient with severe congenital neutropenia and no neurodevelopmental abnormalities. *Pediatr Blood Cancer* 2009; 53:1143–1146.
150. Xue SL, Li JL, Zou JY, *et al.* A novel compound heterozygous HAX1 mutation in a Chinese patient with severe congenital neutropenia and chronic myelomonocytic leukemia transformation but without neurodevelopmental abnormalities. *Haematologica* 2012; 97:318–320.
- This study challenges the view that *HAX1* mutations affecting both isoforms A and B cause neuropathology.
151. Sharp TV, Wang HW, Koumi A, *et al.* K15 protein of Kaposi's sarcoma-associated herpesvirus is latently expressed and binds to HAX-1, a protein with antiapoptotic function. *J Virol* 2002; 76:802–816.
152. Dufva M, Olsson M, Rymo L. Epstein-Barr virus nuclear antigen 5 interacts with HAX-1, a possible component of the B-cell receptor signalling pathway. *J Gen Virol* 2001; 82:1581–1587.
153. Matsuda G, Nakajima K, Kawaguchi Y, *et al.* Epstein-Barr virus (EBV) nuclear antigen leader protein (EBNA-LP) forms complexes with a cellular antiapoptosis protein Bcl-2 or its EBV counterpart BHRF1 through HS1-associated protein X-1. *Microbiol Immunol* 2003; 47:91–99.
154. Yedavalli VS, Shih HM, Chiang YP, *et al.* Human immunodeficiency virus type 1 Vpr interacts with antiapoptotic mitochondrial protein HAX-1. *J Virol* 2005; 79:13735–13746.
155. Modem S, Reddy TR. An antiapoptotic protein, Hax-1, inhibits the HIV-1 rev function by altering its sub-cellular localization. *J Cell Physiol* 2008; 214:14–19.
156. Banerjee A, Saito K, Meyer K, *et al.* Hepatitis C virus core protein and cellular protein HAX-1 promote 5-fluorouracil-mediated hepatocyte growth inhibition. *J Virol* 2009; 83:9663–9671.
157. Cavnar PJ, Berthier E, Beebe DJ, Huttenlocher A. Hax1 regulates neutrophil adhesion and motility through RhoA. *J Cell Biol* 2011; 193:465–473.
158. Sarnowska E, Grzybowska EA, Sobczak K, *et al.* Hairpin structure within the 3'UTR of DNA polymerase beta mRNA acts as a posttranscriptional regulatory element and interacts with Hax-1. *Nucleic Acids Res* 2007; 35:5499–5510.
159. Boztug K, Appaswamy G, Ashikov A, *et al.* A syndrome with congenital neutropenia and mutations in G6PC3. *N Engl J Med* 2009; 360:32–43.
160. Boztug K, Rosenberg PS, Dorda M, *et al.* Extended spectrum of human glucose-6-phosphatase catalytic subunit 3 deficiency: novel genotypes and phenotypic variability in severe congenital neutropenia. *J Pediatr* 2012; 160:679–683; e2.
- This study extends the genotypic and phenotypic spectrum of G6PC3 deficiency and shows that this SCN entity might not have an increased risk of MDS/AML.
161. Hutton JC, O'Brien RM. Glucose-6-phosphatase catalytic subunit gene family. *J Biol Chem* 2009; 284:29241–29245.
162. Boztug K, Klein C. Novel genetic etiologies of severe congenital neutropenia. *Curr Opin Immunol* 2009; 21:472–480.
163. Jun HS, Lee YM, Cheung YY, *et al.* Lack of glucose recycling between endoplasmic reticulum and cytoplasm underlies cellular dysfunction in glucose-6-phosphatase-beta-deficient neutrophils in a congenital neutropenia syndrome. *Blood* 2010; 116:2783–2792.
164. Maurer U, Charvet C, Wagman AS, *et al.* Glycogen synthase kinase-3 regulates mitochondrial outer membrane permeabilization and apoptosis by destabilization of MCL-1. *Mol Cell* 2006; 21:749–760.
165. Zhao Y, Altman BJ, Coloff JL, *et al.* Glycogen synthase kinase 3alpha and 3beta mediate a glucose-sensitive antiapoptotic signaling pathway to stabilize Mcl-1. *Mol Cell Biol* 2007; 27:4328–4339.
166. Ochs HD, Filipovich AH, Veys P, *et al.* Wiskott-Aldrich syndrome: diagnosis, clinical and laboratory manifestations, and treatment. *Biol Blood Marrow Transplant* 2009; 15:84–90.
167. Albert MH, Bittner TC, Nonoyama S, *et al.* X-linked thrombocytopenia (XLT) due to WAS mutations: clinical characteristics, long-term outcome, and treatment options. *Blood* 2010; 115:3231–3238.
168. Dustin ML, Depoil D. New insights into the T cell synapse from single molecule techniques. *Nat Rev Immunol* 2011; 11:672–684.
169. Thrasher AJ, Burns SO. WASP: a key immunological multitasker. *Nat Rev Immunol* 2010; 10:182–192.
170. Kinchen JM, Ravichandran KS. Phagosome maturation: going through the acid test. *Nat Rev Mol Cell Biol* 2008; 9:781–795.
171. Rahajeng J, Caplan S, Naslavsky N. Common and distinct roles for the binding partners Rabenosyn-5 and Vps45 in the regulation of endocytic trafficking in mammalian cells. *Exp Cell Res* 2010; 316:859–874.
172. Erickson RP. Southwestern Athabaskan (Navajo and Apache) genetic diseases. *Genet Med* 1999; 1:151–157.
173. Walne AJ, Vulliamy T, Beswick R, *et al.* Mutations in C16orf57 and normal-length telomeres unify a subset of patients with dyskeratosis congenita, poikiloderma with neutropenia and Rothmund-Thomson syndrome. *Hum Mol Genet* 2010; 19:4453–4461.
174. Wang LL, Levy ML, Lewis RA, *et al.* Clinical manifestations in a cohort of 41 Rothmund-Thomson syndrome patients. *Am J Med Genet* 2001; 102:11–17.