Caspase-11 Controls Interleukin-1β Release through Degradation of TRPC1

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SUMMARY

Caspase-11 is a highly inducible caspase that controls both inflammatory responses and cell death. Caspase-11 controls interleukin 1β (IL-1β) secretion by potentiating caspase-1 activation and induces caspase-1-independent pyroptosis downstream of noncanonical NLRP3 inflammasome activators such as lipopolysaccharide (LPS) and Gram-negative bacteria. However, we still know very little about the downstream mechanism of caspase-11 in regulating inflammation because the known substrates of caspase-11 are only other caspases. Here, we identify the cationic channel subunit transient receptor potential channel 1 (TRPC1) as a substrate of caspase-11. TRPC1 deficiency increases the secretion of IL-1β without modulating caspase-1 cleavage or cell death in cultured macrophages. Consistently, trpc1−/− mice show higher IL-1β secretion in the sepsis model of intraperitoneal LPS injection. Altogether, our data suggest that caspase-11 modulates the cationic channel composition of the cell and thus regulates the unconventional secretion pathway in a manner independent of caspase-1.

INTRODUCTION

Inflammatory responses activated by the innate immunity system constitute a highly efficient barrier in defending against diverse organismal insults. During an innate immune response, early local detection of the insult is mediated by the activation of pattern recognition receptors (PRRs) by pathogen- and damage-associated molecular patterns, which then triggers inflammation by stimulating the production and release of proinflammatory cytokines such as interleukin 1β (IL-1β), as well as pyroptosis, a proinflammatory form of cell death (Keller et al., 2008; Kepp et al., 2010; Martinon et al., 2005). Inflammation, however, must be tightly regulated, as excessive or chronic inflammation may lead to detrimental outcomes, including tissue damage or death as a result of septic shock.

Caspase-11, a member of the caspase family of cysteine proteases, is a critical mediator of inflammation and cell death (Kang et al., 2000; Wang et al., 1999). Caspase-11-deficient (Casp11−/−) mice are highly resistant to lipopolysaccharide (LPS)-induced septic shock, but are more sensitive to infection with Burkholderia, a cytosolic bacterium that escapes the vacuole, demonstrating the inflammatory response as a “double-edged sword” (Aachoui et al., 2013; Wang et al., 1998). Caspase-11 is highly inducible by activators of innate immunity such as LPS, interferon-γ (IFN-γ), and IFN-β (Kang et al., 2000, 2002; Lee et al., 2001; Rathinam et al., 2012; Wang et al., 1996). The expression of caspase-11 is required for caspase-1 activation and subsequent IL-1β secretion downstream of a subgroup of stimuli (Wang et al., 1998). Caspase-1 is directly involved in the processing of pro-IL-1β and the secretion of IL-1β. Caspase-11 potentiates caspase-1 activation downstream of noncanonical NLRP3 inflammasome activators, including diverse Gram-negative bacteria such as Escherichia coli (Kayagaki et al., 2011; Wang et al., 1998), but is dispensable for the activation of caspase-1 downstream of canonical NLRP3 inflammasome activators such as extracellular ATP, the lysosomotropic agent HLOMe, and silica crystals, as well as other canonical inflammasome complexes such as the NLRC4 or AIM2 inflammasomes. In addition, caspase-11 triggers caspase-1-independent pyroptosis and subsequent release of proinflammatory factors such as IL-1α and HMGB1 downstream of noncanonical stimuli, including several vacuolar and cytosolic Gram-negative bacteria (Aachoui et al., 2013; Broz et al., 2012; Case et al., 2013; Kayagaki et al., 2011). Finally, caspase-11 modulates actin cytoskeleton dynamics and restricts Legionella pneumophila growth by promoting bacterial vacuole fusion with lysosomes, as well as by regulating cell migration (Akhter et al., 2012; Li et al., 2007). Caspase-11, which was first characterized in the late 1990s (Wang et al., 1996, 1998), has recently received much attention (Broz et al., 2012; Rathinam et al., 2012). However, the downstream molecular effectors of caspase-11 remain to be identified. To the best of
our knowledge, caspase-3 and caspase-11 itself are the only reported substrates for caspase-11 (Kang et al., 2000).

Transient receptor potential channel 1 (TRPC1) belongs to the canonical TRP subfamily of proteins with six transmembrane domains that assemble to form cation-permeable pores as homo- or heterotetramers (Clapham, 2003). TRPC1 associates with other TRP channels, such as TRPC4 and TRPC5, as well as scaffolding proteins (Strübing et al., 2001). The composition of these complexes determines the distribution of TRPC1 between the endoplasmic reticulum (ER) and the plasma membrane, as well as the various characteristics and gating mechanisms of the channels in which TRPC1 takes part. Although TRPC1 is widely expressed in a variety of tissues and is known to contribute to various physiological functions, it was not known to be involved in innate immunity.

Here, we identify TRPC1 as a substrate for caspase-11 that regulates inflammatory responses. Upon LPS treatment, caspase-11 induction leads to the degradation of TRPC1 in macrophages. TRPC1-deficient macrophages secrete increased amounts of mature IL-1β in response to NLRP3 activators without affecting caspase-1 cleavage or pyroptosis. In addition, trpc1−/− mice show higher IL-1β secretion in their sera following LPS injection. Our data provide a mechanism whereby caspase-11 promotes IL-1β release by degrading TRPC1 independently of caspase-1, and suggest that TRPC1 plays a role in regulating innate immunity by modulating channel complexes during inflammatory responses.

**RESULTS**

**TRPC1 Is a Substrate for Caspase-11**

A yeast two-hybrid screen identified TRPC1 as an interacting protein of inflammatory caspases (data not shown). When coexpressed in human embryonic kidney 293T (HEK293T) cells, the larger isoform of pro-caspase-11 (p43) coimmunoprecipitated with hemagglutinin-TRPC1 (HA-TRPC1; Figure 1A). This interaction of TRPC1 with caspase-11 is selective because, among all TRPC channels, only TRPC1 and (with a lower affinity) TRPC2 could be immunoprecipitated with caspase-11 (Figure S1). Subunits p10 and p20 of caspase-11, but not caspase recruitment domain (CARD) alone, could be coimmunoprecipitated with TRPC1 (Figure 1B). TRPC1 contains six transmembrane domains in its central part and two large N- and C-terminal cytosolic domains (Beech, 2005). FLAG-caspase-11 was able to coimmunoprecipitate independently with HA-TRPC1 N-terminal (HA-TRPC1-N) alone, the N-terminal domain with the transmembrane domain (HA-TRPC1-DN), and the transmembrane domain with the C-terminal domain (HA-TRPC1-TM), but not with the central transmembrane domain alone (HA-TRPC1-TM; Figure 1C).

Since the catalytic domain (p20+p10) of caspase-11 interacts with the TRPC1 channel, we hypothesized that TRPC1 may be a substrate for caspase-11. Indeed, coexpression of caspase-11 and TRPC1 in HEK293T cells greatly decreased the levels of TRPC1 protein, which were restored upon addition of the caspase inhibitor z-VAD.fmk (Figure 2A). Notably coexpression of the proteolytically inactive mutant caspase-11C255G did not have any impact on the levels of TRPC1. Furthermore, incubation...
of purified recombinant caspase-11 p30 with S35-labeled TRPC1 in vitro led to the appearance of multiple TRPC1 fragments and a reduction of full-length TRPC1, which was inhibited by z-VAD.fmk (Figure 2B). On the other hand, recombinant caspase-1 did not cleave TRPC1 in vitro, although it effectively cleaved pro-IL-1β (Figures 2C and S2A). Coexpression of TRPC1 with caspase-1 did not lead to a reduction of TRPC1, contrary to that observed with caspase-11 (Figure S2B), although overexpressed caspase-1 could be coimmunoprecipitated with overexpressed TRPC1 (Figure S2C). Thus, TRPC1 is a substrate for caspase-11, but not caspase-1.

Given that the expression of caspase-11 is highly inducible by LPS (Wang et al., 1996), we next assessed the cleavage and levels of TRPC1 in LPS-treated cells. Ectopic HA-TRPC1 protein levels were substantially reduced 8 hr following LPS treatment, but not in the presence of the pan caspase inhibitor z-VAD.fmk or IDUN-6556, suggesting that TRPC1 is degraded by inflammatory caspases (Figure S2D). Consistently, the levels of endogenous TRPC1 protein decreased in primary macrophages upon LPS treatment, which induced the expression of caspase-11 (Figure 2D). Strikingly, TRPC1 remained stable in casp11−/−, casp11+/−, and casp11−/− macrophages treated with LPS (Figure 2E). On the other hand, the amounts of trpc1 mRNA remained unchanged following LPS treatment of both wild-type (WT) and casp11−/− macrophages, excluding the possibility that TRPC1 is regulated at the transcriptional level (Figure S2E).

Furthermore, we found that stimulation of primary macrophages with E. coli (J53 strain), which induced the expression of caspase-11, also led to a reduction in the levels of endogenous TRPC1 protein (Figure 2F). Notably, TRPC1 levels remained stable in WT macrophages treated with low-dose LPS (1 ng/ml) for 3.5 hr followed by ATP, which was sufficient to lead to the secretion of caspase-11, but not sufficient to induce the expression of caspase-11 (Figure S2F). On the other hand, TRPC1 was degraded in WT, but not casp11−/−, macrophages treated with high-dose LPS (100 ng/ml) for 16 hr followed by ATP, which was sufficient to lead to expression of caspase-11 (Figure S2F). Thus, caspase-1 activation in the absence of caspase-11 expression does not lead to TRPC1 degradation. Taken together, these results suggest that TRPC1 is a physiologically relevant substrate of caspase-11 and is degraded in a caspase-11-dependent manner in macrophages upon induction of caspase-11 after the activation of TLR signaling.

**Caspase-11 Controls the TRPC1-Dependent Decrease in Cytosolic Ca2+ following LPS Treatment**

Since TRPC1 is a membrane protein that can form channels permeable to Ca2+, we next assessed the effect of TRPC1 and caspase-11 on macrophages’ cytosolic Ca2+ concentration in response to LPS stimulation. We found that during continuous LPS treatment, cytosolic Ca2+ concentration demonstrated a transient increase at 2 hr followed by a sustained decrease for up to 10 hr (Figure 3A). Interestingly, this sustained decrease of Ca2+ concentration is dependent on caspase-11 and TRPC1, as no reduction of cytosolic Ca2+ concentration was observed in either trpc1−/− or casp11−/− mice (Figure 3B). Thus, we conclude that TRPC1 cleavage by caspase-11 has a functional impact on Ca2+ flux after LPS stimulation.

**TRPC1 Deficiency Promotes IL-1β Release but Has No Effect on Caspase-1 Activation or Pyroptosis**

Next, we investigated the implication of TRPC1 in the inflammatory response regulated by caspase-11. Caspase-11 is a critical mediator of LPS-induced septic shock (Wang et al., 1996, 1998); therefore, we first assessed the response of trpc1−/− mice to intraperitoneally injected LPS. Indeed, trpc1−/− mice showed significantly higher levels of IL-1β in serum following LPS injection as compared with WT mice, indicating that TRPC1 modulates inflammation in vivo by inhibiting IL-1β secretion (Figure 4A). On the other hand, trpc1−/− mice did not exhibit higher constitutive levels of circulating IL-1β in basal conditions, suggesting that the absence of TRPC1 alone is not sufficient to elicit an inflammatory response (Figure S3A). At the cellular level, caspase-11 is required for caspase-1 activation, IL-1β and IL-18 secretion, and pyroptosis in macrophages infected with E. coli (Kayagaki et al., 2011). Consistently, trpc1−/− macrophages showed higher secretion of mature...
IL-1β and IL-18 in response to *E. coli* (Figures 4B–4D). Since TRPC1 deficiency did not have any effect on caspase-1 cleavage, as indicated by the normal appearance of the caspase-1 subunit p20, we conclude that the cleavage of TRPC1 likely impacts events downstream of caspase-1 activation, including the unconventional secretion pathway that is responsible for mature IL-1β release. Interestingly, *trpc1*−/− macrophages also showed increased mature IL-1β release triggered by canonical NLRP3 inflammasome activators such as ATP, HLOMe, and silica, which is independent of caspase-11 (Kayagaki et al., 2011), but showed no difference in caspase-1 cleavage (Figures 4E and 4F). We thus conclude that LPS priming modulates the TRPC1 complexes by both caspase-11-dependent and -independent, redundant mechanisms. Notably, TRPC1 deficiency did not lead to any increase in the levels of pro-IL-1β, or secretion of TNF-α following LPS priming, ruling out a role of TRPC1 in TLR4 signaling (Figures 4C, 4F, and S3B). In addition, TRPC1-deficiency did not sensitize cells to caspase-11-dependent pyroptosis following infection by *E. coli*, or to caspase-1-dependent pyroptosis induced by canonical stimuli, including ATP and HLOMe (Figures S3C–S3F), supporting the notion that pyroptosis occurs independently of TRPC1 degradation.

**DISCUSSION**

In this study, we describe the involvement of the cationic channel subunit TRPC1 in promoting mature IL-1β release upon inactivation by caspase-11-mediated cleavage in response to Gram-negative bacteria *E. coli* and LPS. Our identification of TRPC1 as a substrate for caspase-11, and evidence for its degradation upon LPS-induced caspase-11 expression, suggest that the TRPC1 complex is remodeled at the onset of inflammation and plays a role in regulating innate immunity. Consistently, *trpc1*−/− macrophages show increased secretion of mature IL-1β upon infection by *E. coli* or stimulation with LPS together with extracellular ATP or lysosome disruption, but show no defect in caspase-1 cleavage or pyroptosis, suggesting that remodeling of the TRPC1 complex regulates the unconventional secretion pathway independently of caspase-1 activation. Thus, our results demonstrate a role for caspase-11 in regulating IL-1β secretion through the degradation of TRPC1 and the remodeling of associated channel complexes independently of caspase-1.

As we reported previously (Kang et al., 2000), casp1−/− mice, which were independently generated by two separate groups, do not express caspase-11 and accordingly should be referred to as casp1−/− casp11−/− mice. Casp1−/− Casp11−/− mice that are deficient for caspase-1 and express caspase-11 were generated recently and helped to clarify the distinct functional roles of caspase-1 and caspase-11 in innate immunity (Kayagaki et al., 2011). Importantly, caspase-11, but not caspase-1, is critical for sensitivity to LPS-induced septic shock and confers resistance against two pathogenic species of Gram-negative *Burkholderia*; therefore, caspase-11 is an important regulator of inflammation (Kayagaki et al., 2011; Rathinam et al., 2012; Wang et al., 1996, 1998). Caspase-11 is unique among caspases in that it is highly regulated at levels of transcription upon the activation of inflammatory responses (Wang et al., 1996, 1998). Pro-caspase-11 is inducible upon stimulation of multiple ligands involved in innate immunity, including LPS, IFNγ, and type I IFN (Broz et al., 2012; Gurung et al., 2012; Hur et al., 2001; Lee et al., 2001; Lin et al., 2000; Rathinam et al., 2012; Schauvliege et al., 2002; Wang et al., 1998; Yen and Ganea, 2009). Whether expression of caspase-11 is sufficient for its autoactivation is the subject of debate (Broz et al., 2012; Kang et al., 2000, 2004; Rathinam et al., 2012; Wang et al., 1998). Indeed, although pro-caspase-11 induction by extracellular LPS or IFNγ is sufficient for its autoprocessing in the p30 form, an additional signal that was recently identified to be cytosolic LPS is required to trigger caspase-11-dependent pyroptosis and caspase-1 activity (Broz et al., 2012; Hagar et al., 2013; Kayagaki et al., 2013; Rathinam et al., 2012). However, due to the lack of a specific caspase-11 activity assay and identification of a caspase-11-specific substrate, caspase-11 activity per se was not assessed in previous studies. Therefore, cytosolic LPS may also control pyroptosis and caspase-1 activation by acting downstream of caspase-11 activation. Detection of autoprocessed p30 demonstrates that caspase-11 is active in macrophages treated with extracellular LPS (Rathinam et al., 2012). In the present study, we show that TRPC1 is degraded by caspase-11 in macrophages treated with extracellular LPS, demonstrating that caspase-11 can be activated without triggering pyroptosis.

Despite the high interest in caspase-11, very few of its substrates have been identified. Caspase-11 can control cell mobility and intracellular vesicular trafficking by modulating actin cytoskeleton independently of its protease activity (Akhter et al., 2012; Li et al., 2007). On the other hand, caspase-11 is capable of autoprocessing (Kang et al., 2000). In addition, direct cleavage of caspase-3 by caspase-11 was shown to contribute to deleterious cell death in several animal models of neurodegenerative disease (Hisahara et al., 2001; Kang et al., 2000; Shibata et al., 2000). However, the requirement for caspase-3 in caspase-11-dependent pyroptosis has not yet been investigated (Kang et al., 2002;
Kayagaki et al., 2011). Lastly, the molecular mechanism of caspase-11-dependent caspase-1 activation downstream of noncanonical NLRP3 activators is unknown. Although caspase-11 associates with caspase-1, caspase-1 is not a direct substrate for caspase-11 (Wang et al., 1998). Thus, TRPC1 constitutes the only noncaspase substrate for caspase-11 identified so far.

TRPC1 assembles with other TRPCs to form heterotrameric cation-permeable channels with current characteristics different from those of homotetramers (Strübing et al., 2001). In particular, TRPC1 reduces the Ca\(^{2+}\) permeability of the respective channels in homodimers while participating in the channel pore of at least TRPC3, TRPC4, and TRPC5. TRPC1 also inhibits TRPV6 expression at the plasma membrane (Schindl et al., 2012; Strübing et al., 2001). In neurons, TRPC1 forms a complex with TRPC5 that localizes at the plasma membrane (Strübing et al., 2001). However, TRPC1 cannot be detected at the plasma membrane in macrophages where TRPC5 is not expressed (Kunert-Keil et al., 2006). Given the complexity of these functional interactions, TRPC1 degradation during an inflammatory response may alter the gating properties of TRPC1-containing channels in the plasma or intracellular membranes to promote unconventional protein secretion.

Taken together, our data point to a role of cationic channels in the downstream process of unconventional secretion of IL-1\(\beta\) and IL-18, and suggest that in addition to its already described regulatory functions in the cytoskeleton, cell death, and caspase-1 activation, caspase-11 controls IL-1\(\beta\) and IL-18 release by modulating the cationic channel composition of the cell. Our results also suggest a possible role for the TRPC1-associated channelosome in regulating unconventional secretion in general, which needs to be investigated in future studies.

**EXPERIMENTAL PROCEDURES**

**Mice**

C57Bl6, casp11\(^{-/-}\) (Wang et al., 1998), casp1\(^{-/-}\) mice deficient for both caspase-1 and caspase-11 (casp1\(^{-/-}\) casp11\(^{-/-}\) mice) (Kang et al., 2000; Kuida et al., 2002).
Effectively, for 4 hr at 4°C, cells were lysed in lysis buffer (Tris HCl, pH 7.6 50 mM, NaCl 400 mM, NP-40 1%, PMSF 1 mM, and protease inhibitors). HA-TRPC1 and FLAG-caspase-11 were immunoprecipitated, 5 x 10^6 cells/ml in DMEM (GIBCO) supplemented with 1x PS and 10% FBS. For all E. coli infection experiments, cells were grown in the absence of PS. For trichloroacetic acid (TCA) precipitation of the media, cells were rinsed and treated in DMEM+PS without serum.

**Cell-Growth Conditions**

HEK293T and LN-18 cells were maintained in Dulbecco’s modified Eagle’s medium (DMEM) and DMEM/F12 (GIBCO), respectively, supplemented with 1x penicillin/streptomycin (PS; GIBCO) and 10% fetal bovine serum (FBS; GIBCO). Immunofluorescence peritoneal macrophages were cultured at 0.5–1 x 10^6 cells/ml in DMEM (GIBCO) supplemented with 1x PS and 10% FBS. For all E. coli infection experiments, cells were grown in the absence of PS. For trichloroacetic acid (TCA) precipitation of the media, cells were rinsed and treated in DMEM+PS without serum.

**Activation of the Inflammasome**

E. coli (strain J35) were grown in lysogeny broth (LB) to log expansion phase, resuspended in PBS, and added to macrophages. For IL-1β secretion assay, gentamycin (50 μg/ml) was added 1.5 hr after infection and supernatant was collected after a total of 16 hr following infection.

** Constructs**

Constructs coding for FLAG-caspase-11, FLAG-caspase-11CARD98 (aa 1–98), FLAG-caspase-11CARD103 (aa 1–103), FLAG-caspase-11p30, FLAG-caspase-11p10, pRNA-IL-1β, and pRNA-TRPC1 were previously described (Li et al., 2009), FLAG-caspase-11p20 (aa 79–285) was cloned in pFLAG3X, HA-TRPC1, HA-TRPC1ΔC (deleted of the C-terminal domain, aa 1–649), HA-TRPC2, HA-TRPC3, HA-TRPC4, HA-TRPC5, and HA-TRPC6 were kind gifts from P.F. Worley and J. Yuan (Yuan et al., 2003), HA-TRPC1-N (N-terminal domain, aa 1–315), HA-TRPC1ΔN (deleted of the N-terminal domain, aa 306–760), HA-TRPC1-TM (transmembrane domain, aa 306–613) were cloned following PCR amplification.

**Reagents**

The following reagents were used: LPS from E. coli 0111:B4, ATP, and nigericin (Sigma-Aldrich); silica (MIN-U-SIL 5; U.S. Silica); HLLOMe (ChemImpex International); z-VAD-fmk (Alexis); and IDUN-6556 (Linton et al., 2005).

**Immunoprecipitation**

For coimmunoprecipitation experiments, HEK293T cells were lysed in lysis buffer (Tris HCl, pH 7.6 50 mM, NaCl 400 mM, NP-40 1%, PMSF 1 mM, and protease inhibitors). HA-TRPC1 and FLAG-caspase-11 were immunoprecipitated with anti-HA and anti-flag M2 agarose beads (Sigma-Aldrich), respectively, for 4 hr at 4°C. Immunoprecipitations were washed in lysis buffer and proteins were eluted in Laemmli sample buffer 2x. For anti-TRPC1 immunoprecipitation, 5 x 10^6 macrophages were lysed in RIPA buffer. TRPC1 was immunoprecipitated using protein A/G UltraLink resin (Thermo Scientific) and anti-TRPC1 antibody (Millipore), and eluted in Laemmli sample buffer.

**Transfection**

HEK293T cells were transfected using the CaCl2 procedure. LN-18 cells were transfected using transfectIT-LT1 transfection reagent (Mirus Bio).

**Western Blot Analysis**

Samples were analyzed by SDS-PAGE, transferred to PVDF membranes, blocked, and probed according to standard procedures. The following antibodies were used: anti-caspase-11 (Wang et al., 1998), anti-caspase-1 (Wang et al., 1998), anti-HA (Santa Cruz biotechnology), anti-FLAG M2 (Sigma-Aldrich), anti-TRPC1 (Millipore), anti-actin (Millipore), anti-IL-1β (R&D Systems), anti-IL-18 (MBL), and anti-laminB (Santa Cruz).

**In Vitro Caspase Cleavage Assay**

In vitro production of 32P-labeled TRPC1 and IL-1β was performed using TNT-coupled transcription/translation kits (Promega). Purification of recombinant Caspase-11 p30 and in vitro cleavage assay by recombinant Caspase-11 p30 or active human Caspase-1 (p30; Chemicon International) were performed as described previously (Kang et al., 2000).

**ELISA**

In vitro secretion of IL-1β and TNF-α in macrophage culture supernatants was measured using mouse IL-1β/IL-12 and TNF-α DuoSet kits (R&D Systems). In vivo secretion of IL-1β in the serum was assessed using the mouse IL-1β/IL-12 Quantikine kit (R&D).

**Intracellular Ca²⁺ Measurement**

Cells were loaded with the ratiometric Ca²⁺ indicator Fura-2/AM in Opti-MEM (at 37°C, 5% CO2) for 30 min, washed, and incubated for an additional 15 min prior to imaging. Ca²⁺ imaging was performed in Ringer’s solution containing 2 mM Ca²⁺. Excitation was carried out at 340 nm and 380 nm using a Lambda DG4 Ultra High Speed Wavelength Switcher (Sutter Instrument), and emission was collected at 510 ± 10 nm using a Hamamatsu ORCA R2 CCD camera and Slidebook data acquisition software.

**Cytotoxicity Assay**

Cell death was measured as the percentage of lactate dehydrogenase (LDH) release according to standard protocols. DMEM+10% FBS was used as the blank control, and supernatants of control cells treated with Triton X-100 0.01% were used to measure total LDH release. Cell death was measured by propidium iodide (PI) incorporation. PI (1 μg/ml) was added to the cell culture, and cells were observed by live imaging using a Nikon Ti inverted microscope equipped with a 10× Plan Apo NA 0.3 objective lens. Images were acquired with a Hamamatsu ORCA-R2 cooled CCD camera controlled with MetaMorph 7 software (Molecular Devices).

**SUPPLEMENTAL INFORMATION**

Supplemental Information includes three figures and can be found with this article online at http://dx.doi.org/10.1016/j.celrep.2014.02.015.

**AUTHOR CONTRIBUTIONS**


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