



Detecting functional groups of *Arabidopsis* mutants by metabolic profiling and evaluation of pleiotropic responses

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Metabolic profiles and fingerprints of *Arabidopsis thaliana* plants with various defects in plastidic sugar metabolism or photosynthesis were analyzed to elucidate if the genetic mutations can be traced by comparing their metabolic status. Using a platform of chromatographic and spectrometric tools data from untargeted full MS scans as well as from selected metabolites including major carbohydrates, phosphorylated intermediates, carboxylates, free amino acids, major antioxidants, and plastidic pigments were evaluated. Our key observations are that by multivariate statistical analysis each mutant can be separated by a unique metabolic signature. Closely related mutants come close. Thus metabolic profiles of sugar mutants are different but more similar than those of photosynthesis mutants. All mutants show pleiotropic responses mirrored in their metabolic status. These pleiotropic responses are typical and can be used for separating and grouping of the mutants. Our findings show that metabolite fingerprints can be taken to classify mutants and hence may be used to sort genes into functional groups.

Keywords: metabolic profiling, fingerprinting, functional genomics, *Arabidopsis thaliana*, pleiotropic response, chloroplast

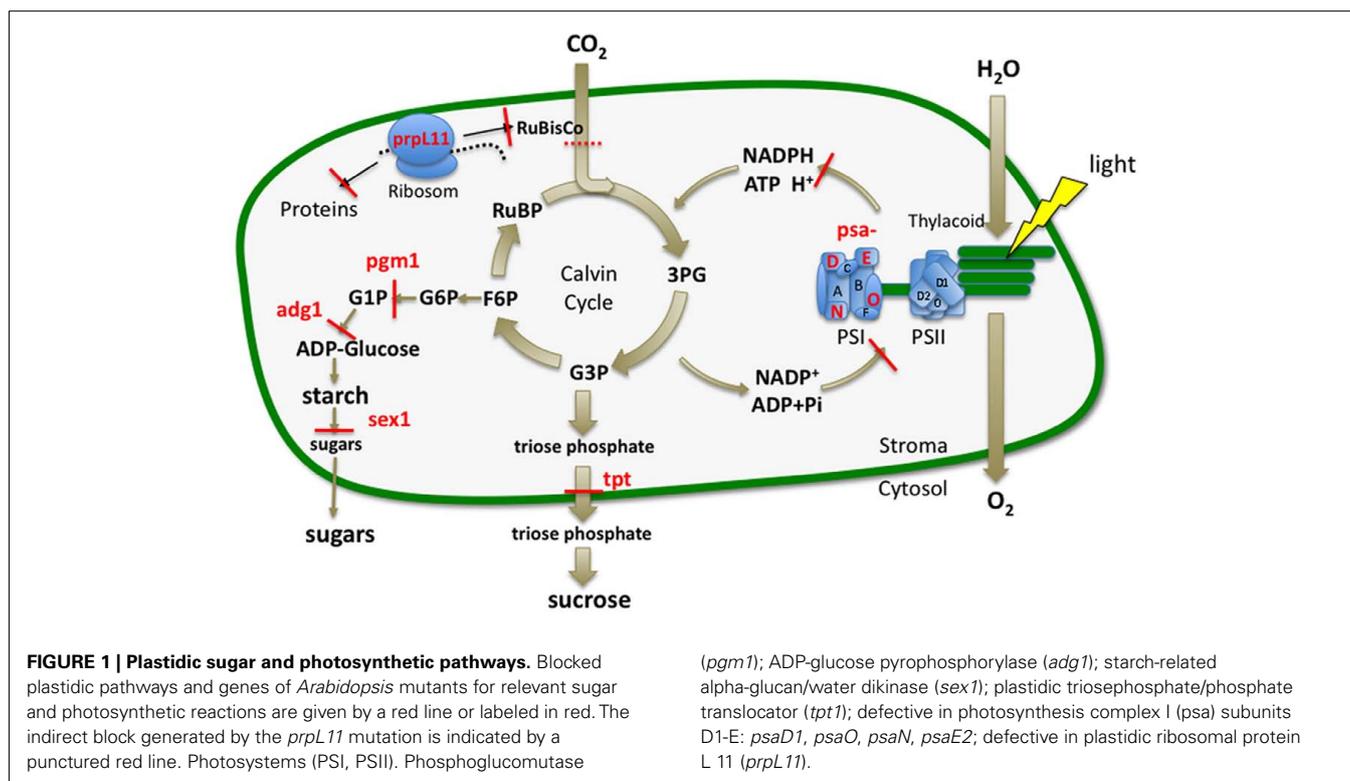
INTRODUCTION

Approximately 30% of the genes of the model organism *Arabidopsis thaliana* have unknown functions and only 22% are experimentally characterized according to the October 2011 TAIR statistics (Brown et al., 2005; Clare et al., 2006; Swarbreck et al., 2008). Even in the best-studied organism *E. coli* it is unclear what 40% of the genes are doing (Tohsato et al., 2010). Such deficiencies along with the perspective of the rapidly increasing number of sequenced genomes underline the need to assign functions to unknown genes. Strategies for their exploration range from structural biology providing crystal structures vital to explain how, e.g., the ubiquitin ligase (Pickart, 2001) works, to the field of functional bioinformatics exploiting accumulated database knowledge such as sequence motifs, similarities of genes, expression data, predicted secondary structures, or structural classifications of proteins (King et al., 2004). A major part however is studied by screening forward and reverse genetic mutants. In recent years metabolomics came into focus contributing information about small biochemical molecules to solve the puzzle of functional genomics. Driven by advances in mass spectrometry and computational biology in particular metabolite profiling and fingerprinting became powerful tools complementing insights derived from genome, transcriptome, and proteome with data of metabolite content (Fiehn, 2002; Hall et al., 2002; Bino et al., 2004). Applications are exemplified in a number of ground-breaking publications focused on aspects of method development and to a growing extend on assessing functional genomics (Fiehn et al., 2000; Roessner et al., 2001, 2006; Bolling and Fiehn, 2005; Sekiguchi et al., 2005; Bijlsma et al., 2006; Messerli et al., 2007; Winder et al., 2008; Tohge and Fernie, 2010). The forward strategy of profiling genetic mutants in the context

of biochemical pathways however harbors the risk that pleiotropic effects mask primary events. Using conditional mutants may circumvent these effects. However their construction is mostly tricky and may require time-consuming measurements. We therefore asked with this work if primary events from metabolic profiles can be defined revealing a metabolic signature in spite of secondary changes. This should be possible if these changes are recognized as such. A primary effect might be visible as an accumulation or drop of metabolites generated directly by a defective enzyme and in tight relation to its position in a pathway. Secondary effects instead involve changes of multiple phenotypic traits, e.g., as growth or stress response (Williams, 1957). Here one has to consider that mutations outside metabolic pathways might generate pleiotropic effects when analyzed on the basis of changes in metabolism.

As a model system we selected available *Arabidopsis* mutants with various plastidic defects known to effect sugar metabolism or photosynthesis (Figure 1). We generated metabolite profiles and fingerprints from leaf material applying the methodology recently developed in our laboratory (Kogel et al., 2010). Here extractions and measurements, e.g., by LC-diode array detection or with soft ionization by IC-ESI/MS/MS are adapted to individual metabolite groups to ensure optimal recovery and detection rates. We show that with statistical analysis similar metabolic patterns can be detected sorting the candidates into several functional groups. With the targeted analysis of 74 metabolites we found that their content reflected the phenotypes and could be related to the affected pathway. Accordingly mutations with minor effects on plant growth displayed less distinct metabolic changes.

For mutants strongly retarded in growth dramatic secondary effects such as the accumulation of stress indicative metabolites or



changes of leaf pigment composition along with remarkable primary effects were detected. An alignment of all changes revealed similarities and differences between the candidates and enabled the recognition of metabolic signatures for the functional groups.

These results suggest that our approach of metabolite profiling is suitable to detect similarities between different mutants that allows their grouping into functional categories.

MATERIALS AND METHODS

PLANT LINES

Mutants were established in the *Arabidopsis* (*A. thaliana*) ecotype *Columbia* (Col0) background whereas *psaE2* was in ecotype *Was-silewskija* (Ws). *Arabidopsis* lines: controls Col0, Ws, mutant lines defective in starch synthesis: *pgm1* (phosphoglucomutase1; Lin et al., 1988); *adg1* (ADP-glucose pyrophosphorylase; Caspar et al., 1985); defective in starch utilization: *sex1* (starch-related alpha-glucan/water dikinase; Yu et al., 2001), defective in triosephosphate export: *tpt1* (plastidic triosephosphate/phosphate translocator; Schneider et al., 2002); defective in photosynthesis complex I (PSI): *psaE2* (Ihnatowicz et al., 2007); *PsaD1* (Ihnatowicz et al., 2004); *psaO* (Leister, 2003); *psaN* (Leister, 2003); defective in plastidic ribosomal protein L 11: *prpL11* (Pesaresi et al., 2001).

CULTIVATION AND HARVEST

Seeds of *Arabidopsis* mutants were obtained from the Nottingham *Arabidopsis* Stock Centre (NASC, Nottingham, UK), and all of the mutant plants were identified by polymerase chain reaction (PCR) and confirmed by reverse transcription (RT). Plants were cultivated in growth chambers under short day conditions with a diurnal cycle of 8 h light/16 h darkness at 22°C under

120 $\mu\text{mol m}^{-2} \text{s}^{-1}$ of illumination. Leaves of four biological replicates were harvested 4 weeks after sowing and 4 h after onset of light and immediately frozen into liquid nitrogen.

METABOLOME PROFILING (TARGETED ANALYSIS) AND METABOLITE FINGERPRINTING (UNTARGETED)

Metabolome analysis was conducted from 50 to 100 mg samples of shock-frozen *Arabidopsis* leaf tissue for four biological replicates as described previously (Kogel et al., 2010).

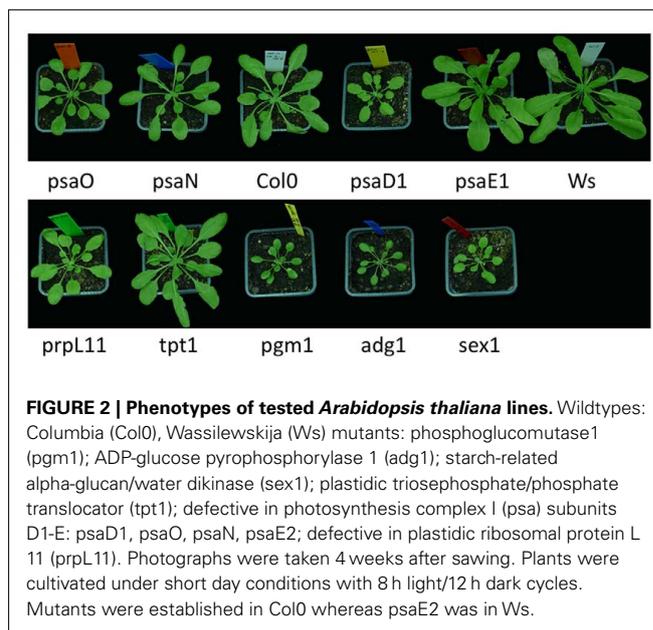
In brief, for the targeted analysis of *sugars* and *free amino acids* samples were extracted in 80% ethanol. Enzymatic assays for sugars were performed using a microtiter plate reader (BioTek, Bad Friedrichshall, Germany). Amino acid contents were determined after derivatization using the fluorophore 6-aminoquinolyl-*N*-hydroxysuccinimidyl carbamate (AQC; Cohen and Michaud, 1993) and a reversed phase HPLC-separation Dionex Summit HPLC-system and fluorescence detector. *Thiol-compounds* (glutathione, γ -glutamylcysteine, and cysteine) were extracted with 0.1 M hydrochloric acid (HCl). To assay total thiol-compounds aliquots of the extract were reduced with 10 mM DTT and then derivatized with monobromobimane. For the measurement of oxidized compounds, the thiol-groups of the reduced forms were blocked with NEM prior to the above procedure. Derivatized thiol-compounds were subsequently separated with the HPLC-system described above. *Tocopherols* (δ , γ , and α) were extracted with 100% methanol and separated isocratically with the HPLC-system described above. Total and reduced *ascorbate* contents were determined spectrophotometrically by measuring 2,2'-dipyridyl/ FeCl_2 -complexes generated after extraction of samples with 5% 5-sulfosalicylic acid and reduction of FeCl_3 by ascorbate. For total

ascorbate assays aliquots of the extracts were first reduced by dithiothreitol (DTT) and remaining sulfhydryl groups were then blocked with *N*-ethylmaleimide (NEM) prior to the above procedure. For the measurement of leaf pigments 50 mg frozen powder was extracted with 400 μ l methanol. During the following steps, samples were shielded from light and kept on ice. After incubation for 5 min at 4°C, 400 μ l 50 mM Tris/HCl pH 8.0/1 M NaCl were added, and the mixture was incubated for 5 min before addition of 800 μ l chloroform for extraction of the carotenoids from the methanol phase. Samples were inverted for 5 min, incubated for 10 min, and then centrifuged at 13,000 rpm for 5 min to achieve phase separation. Extraction with chloroform was repeated with 400 μ l chloroform, and the lower phases were pooled and vacuum dried in a speed-vac. For reverse-phase chromatography, extracts were dissolved in 200 μ l eluent B, and 20 μ l aliquots were resolved on a HPLC-System (Dionex, Idstein, GE) consisting of an Acclaim PA C16 column (id. 4.6 mm, length 150 mm, particle size 5 μ m) on an Ultimate 3000 HPLC connected to an ICS 2600 photodiode array detector. A trimeric gradient using eluent A (water), eluent B (methanol:acetonitrile:isopropanol:water, 73:20:5:2), and eluent C (methanol:ethylacetate, 68:32) was applied. The profile was 0–13 min 96%B, 13–17 min 100%C, 17–30 min 100%C, 30–32 min 96%B, and 32–40 min 96%B over 40 min with a flow rate of 0.5 ml/min and temperatures of 4°C for the autosampler and 24°C for the column oven. Pigments were detected and quantified mainly based on their absorption at 450 nm, or on standard curves with pure substances (Sigma-Aldrich, Munich, GE, CaroteNature, Lupsingen, CH). Identification was achieved according to the retention time and to specific 3D spectra between 300 and 700 nm. *Phosphorylated intermediates and carboxylates* were extracted with perchloric acid and were determined as described by Horst et al. (2010) applying ion chromatography with an ICS3000 HPLC-system (Dionex) and ESI/MS/MS detection using a QTrap 3200 Triple-Quadrupole mass spectrometer with turboV ion source (Applied Biosystems) operated in multiple reaction monitoring mode. *Untargeted metabolome profiling* (fingerprinting) was conducted from the perchloric acid extracts and with the HPLC- and mass spec-system described for the targeted analysis. Negative ions for the ESI/MS full scan were generated at –4.5 kV and a declustering potential of –20 V. The entrance potential (EP) was from –6 to –4 V, and gas pressures were 20 psi (curtain), 30 psi (nebulizer), and 20 psi (turbogas). A mass range of *m/z* 60–600 was recorded with 1 scan/s over 80 min. Five hundred peaks were aligned and evaluated after import into the program Marker View™ (Applied Biosystems) with a retention time tolerance of 0.75 min and a mass tolerance of 1.0 amu. Retention time corrections and normalization was done with the internal standard pipes. For PC analysis of fingerprinting data, quality control (QC) samples for PC analysis were generated as described (Bijlsma et al., 2006) by pooling equal-volume amounts from all analyzed samples. Artifact peaks were removed before the analysis was conducted with Marker View™.

RESULTS

PHENOTYPIC CHARACTERIZATION

The phenotypes obtained for the mutants in growth chambers (Figure 2) 4 weeks after sawing confirmed earlier findings (Caspar et al., 1985; Lin et al., 1988; Pesaresi et al., 2001; Yu et al.,



2001; Schneider et al., 2002; Leister, 2003; Ihnatowicz et al., 2007). Most retarded in growth were the starch synthesis and mobilization mutants *pgm1*, *adg1*, and *sex1*. The impact of insufficient starch supply generating a general shortage of carbon is consistent with the growth retardation observed. A similar phenotype developed the PSI mutant *psaD1*. In addition, we confirmed the observation of a light green phenotype for *psaD1*, which is consistent with a reduction of chlorophylls and carotenenes (Ihnatowicz et al., 2004). Two other KO-mutants of the PSI proteins *psaO*, *psaN*, and a mutant of the large subunit of the plastidic ribosome *prpL11* showed a milder phenotype still being clearly smaller than the control but larger than *psaD1*. The triosephosphate/phosphate translocator mutant *tpt1* and a fourth PSI mutant *psaE2* propagated in ecotype Ws displayed phenotypes looking hardly different from their wildtypes Col0 and *Wassilewskija*, respectively. This again is in agreement with earlier findings (Varotto et al., 2000).

METABOLOME ANALYSIS OF ARABIDOPSIS LEAVES

To rapidly obtain an unbiased overview of metabolite distributions of the *Arabidopsis* lines we performed IC/MS fingerprinting from four biological replicates of leaves extracted with perchloric acid. For this untargeted approach full MS scans from 60 to 600 amu were recorded and 431 masspeaks were finally evaluated statistically by principal component analysis. As depicted in Figure 3A the multivariate method sorted the three sugar mutants *sex1*, *adg1*, and *pgm1* into a metabolic group being more similar to the triosephosphate transporter mutant *tpt1*, the ribosomal mutant *prpL11* and surprisingly *psaD1*, one of the four mutants of PSI. The remaining three PSI mutants appeared to be more distant and comparable to the wildtypes Col0 or Ws. According to PC2 *tpt1* and *prpL11* and the three PSI mutants *psaO*, *psaN*, *psaD1* but not *psaE2* were more similar. Three pairs of lines namely *adg1/pgm1*, *Ws/psaE2*, and *psaO/psaN* clustered more closely.

To verify this sorting and to identify responsible metabolites subsequently a targeted profiling approach was conducted. Here

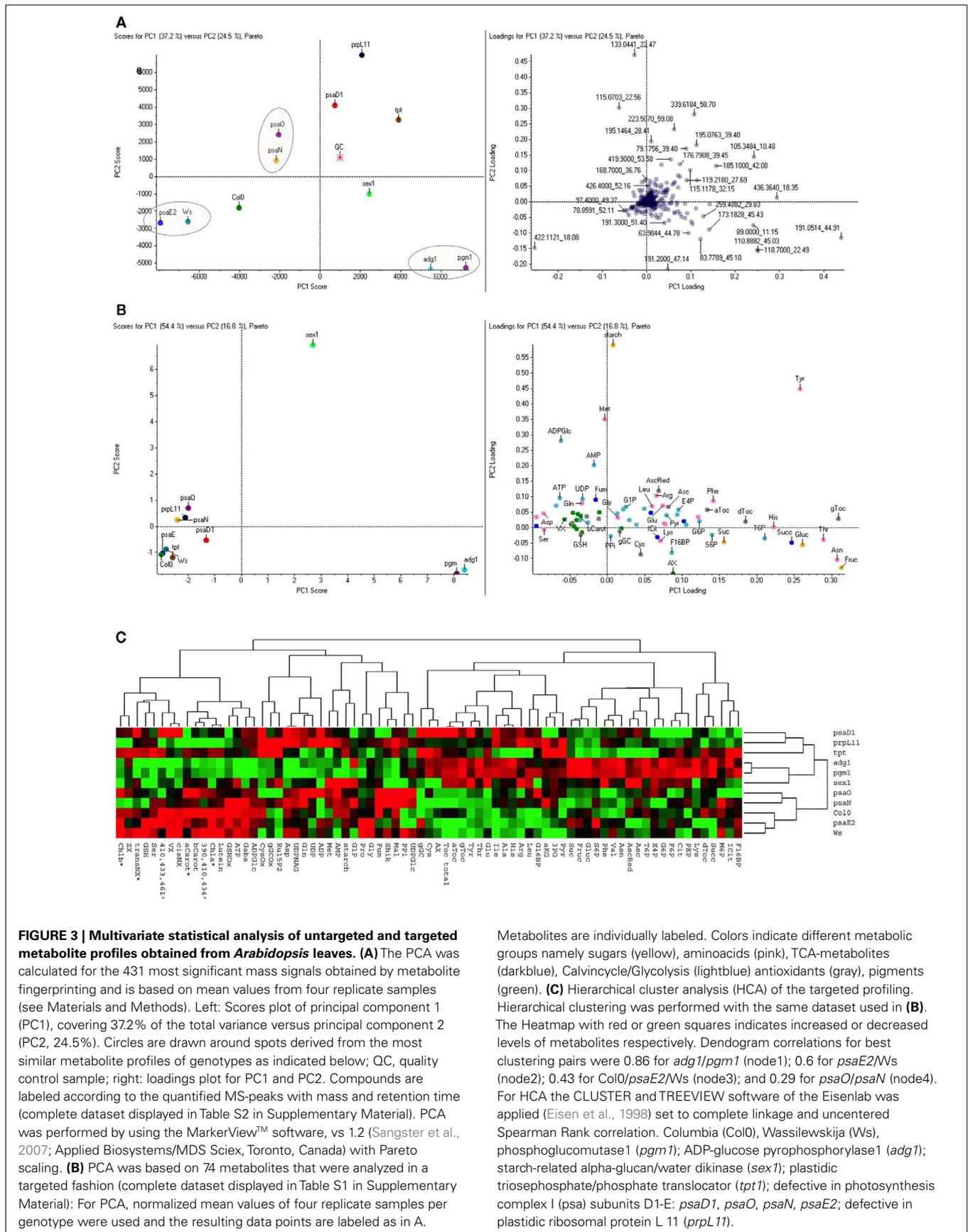


FIGURE 3 | Multivariate statistical analysis of untargeted and targeted metabolite profiles obtained from *Arabidopsis* leaves. (A) The PCA was calculated for the 431 most significant mass signals obtained by metabolite fingerprinting and is based on mean values from four replicate samples (see Materials and Methods). Left: Scores plot of principal component 1 (PC1), covering 37.2% of the total variance versus principal component 2 (PC2, 24.5%). Circles are drawn around spots derived from the most similar metabolite profiles of genotypes as indicated below; QC, quality control sample; right: loadings plot for PC1 and PC2. Compounds are labeled according to the quantified MS-peaks with mass and retention time (complete dataset displayed in Table S2 in Supplementary Material). PCA was performed by using the MarkerView™ software, vs 1.2 (Sangster et al., 2007; Applied Biosystems/MDS Sciex, Toronto, Canada) with Pareto scaling. **(B)** PCA was based on 74 metabolites that were analyzed in a targeted fashion (complete dataset displayed in Table S1 in Supplementary Material): For PCA, normalized mean values of four replicate samples per genotype were used and the resulting data points are labeled as in A.

Metabolites are individually labeled. Colors indicate different metabolic groups namely sugars (yellow), aminoacids (pink), TCA-metabolites (darkblue), Calvin cycle/Glycolysis (lightblue) antioxidants (gray), pigments (green). **(C)** Hierarchical cluster analysis (HCA) of the targeted profiling. Hierarchical clustering was performed with the same dataset used in **(B)**. The Heatmap with red or green squares indicates increased or decreased levels of metabolites respectively. Dendrogram correlations for best clustering pairs were 0.86 for *adg1/pgm1* (node1); 0.6 for *psaE2/Ws* (node2); 0.43 for *Col0/psaE2/Ws* (node3); and 0.29 for *psaO/psaN* (node4). For HCA the CLUSTER and TREEVIEW software of the Eisenlab was applied (Eisen et al., 1998) set to complete linkage and uncentered Spearman Rank correlation. Columbia (*Col0*), *Wassilewskija* (*Ws*), phosphoglucomutase1 (*pgm1*); ADP-glucose pyrophosphorylase 1 (*adg1*); starch-related alpha-glucan/water dikinase (*sex1*); plastidic triosephosphate/phosphate translocator (*tpt1*); defective in photosynthesis complex I (*psa*) subunits D1-E: *psaD1*, *psaO*, *psaN*, *psaE2*; defective in plastidic ribosomal protein L 11 (*prpL11*).

the content of 74 metabolites including major carbohydrates, phosphorylated intermediates, carboxylates, free aminoacids, major antioxidants such as tocopherol, ascorbate, and glutathione as well as plastidic pigments (for complete dataset see Table S1 in Supplementary Material) were measured. Evaluation of this biased approach with hierarchical cluster analysis confirmed the separation of the untargeted approach (Figure 3C). As for the untargeted analysis the same three clustering pairs of plant lines *adg1/pgm1*, *Ws/psaE2*, and *psaO/psaN* fell into three clusters with the highest correlations.

By the alternative evaluation of the targeted dataset with PCA now the main principal component with 54.4% of the total variance clearly separated the three sugar mutants *sex1*, and the closely clustering *adg1* and *pgm1* from all other lines tested (Figure 3B). Most important for this separation were the increased hexoses, γ -tocopherol, asparagine, threonine, and tyrosine while malate, aspartate, and most of the pigments were decreased (for complete dataset see of the loadings see Table S2 in Supplementary Material). A dominant difference of *sex1* to all other lines was detected by PC2. Next to the outstanding starch elevation also high methionine and tyrosine levels and strong increases of ADP-glucose and AMP were most responsible for this distribution. A better resolution for the candidates clustering at the negative side of the scoring plot in Figure 3C was obtained by applying the algorithm Autoscaling instead of alternative Pareto Scaling. However the unequal footing of Pareto Scaling allows a better interpretation of the Loadings plot (Figure A1A in Appendix). Elimination of the three dominating mutants led to a clustering of the remaining lines that was most comparable to the untargeted approach (Figure A1B in Appendix).

COMPARING METABOLIC SIGNATURES

To allow a more detailed interpretation of the observed global changes, metabolite contents of the targeted analysis were aligned and displayed relative to the wildtype Col0. As depicted in Figure 4, by comparison similarities detected statistically now became obvious and allowed to define distinct metabolic signatures for the plant lines. As expected *adg1* and *pgm1*, both hampered in starch synthesis contained very little starch, while for *sex1* a remarkable eightfold excess was measured. The soluble sugars for these lines accumulated up to ninefold in the starchless but only twofold in *sex1*. Most prominent but not surprising was the strong variance of the starch precursor ADP-glucose, which was almost absent in the starchless but twofold elevated in *sex1*. Here interestingly also AMP and methionine were elevated one- and fivefold, respectively. Outstanding for the amino acids of the starchless was the up to tenfold increase of the famine indicating asparagine (Usadel et al., 2008). Histidine and tyrosine levels were further major differences. Remarkable for the pigments of the starchless was the twofold-increased antheraxanthin (AX), a carotenoid of the xanthophyll cycle. In *Sex1* instead AX was at wildtype level. Less dramatic different sugar levels were detected for the other mutants tested. *PsaD1* however, a further PSI mutant, displayed a starch deficiency together with a hexose reduction. The metabolic pattern of *psaD1* is peculiar for a fivefold elevation of AX. The pattern of amino acids and antioxidants resembled to *sex1* or the *adg1/pgm1* pair with respect to high tyrosine, threonine, asparagine, histidine, and

the tocopherols α and γ but not δ . The pattern of phosphorylated intermediates and carboxylates was widely different from the other lines tested. Only here most of the metabolites were below the level of wildtype including the Calvin cycle metabolites 3PG, F16BP, F6P, E4P, and Ru15P2. The observed decrease of the hexoses is similar in *prpL11* and *tpt1* mutants. *Tpt1* as *psaD1* and the starchless *pgm1* and *adg1* lines showed a remarkable increase in AX while the remaining metabolic signatures were different. Furthermore in *tpt1* the levels of almost half of the amino acids were reduced, while they were most often elevated for the other lines. Major differences were the elevated serine and the reduced glycine and methionine levels. A further variant metabolic signature displayed the mutant *prpL11*. Besides AX here all measured pigments were up to 50% below wildtype including chlorophyll and the carotenes. Delta-tocopherol was reduced as in all PSI mutants and in *Ws*. Methionine, asparagine, and tyrosine were increased by one- to twofold. AMP, ADP, UDP, Pyr, and PPI were up as well. The clustering *psaO* and *psaN* showed a further widely similar metabolic profile. Here sugars, antioxidants and pigments were close to wildtype while the latter were slightly reduced for *psaN*. The amino acids were increased almost as in *prpL11*. From the phosphorylated metabolites ADP-glucose was increased to 150% for both lines. PPI and G16BP was about 100% elevated. Shikimate was increased between 10 and 20%. In *psaO* F16P2 was reduced by 50% other than in *psaN* (+10%). The other metabolites of the Calvin cycle were increased. *PsaE* and its control *Ws* showed the third pair of similar signatures. Remarkably, here in both lines proline was increased twofold. Major differences are the almost twofold-increased PPI, which reminded to the *psaO/psaN* pair and a twofold increase of S6P, which is consistent with an increased sucrose level. S6P was elevated whenever sucrose was elevated with the exception of *psaN*. A decrease of S6P was found only in the three lines *prpL11*, *tpt1*, and *psaD1*. In these lines additionally hexoses were reduced while sucrose was like in wildtype.

Interestingly 13 metabolites were found to be elevated always above the Col0 wildtype independently of the candidate. These were γ -GC for the antioxidants; Phe, Leu, Ile, Val, Tyr, Thr, and Glu for the aminoacids and G16P2, PPI, Pyr, ADP, and Ru15BP for the phosphorylated metabolites. Only one metabolite, an unidentified carotenoid (peak spectrum 390, 410, 434 nm) was found to be decreased below Col0 in all mutants.

DISCUSSION

GENETIC DEFECTS ARE REFLECTED BY METABOLIC PROFILES

The high similarity found for the metabolic profiles of the pathway neighboring starch biosynthesis mutants *adg1* and *pgm1* was not unexpected. Their signature confirmed earlier findings describing the accumulation of soluble sugars as more photoassimilates are exported from the chloroplast during the day (Caspar et al., 1985; Lin et al., 1988). The lack of starch means starvation for carbon and less investment into biomass resulting in serious growth retardation as observed. The deficiency is moreover responsible for mobilization of carbon from amino acids released by protein degradation (Usadel et al., 2008) and other carbon sources. Our findings of highly elevated asparagine and the general increase of most other amino acids are in agreement with this statement. It furthermore explains the measured accumulation of

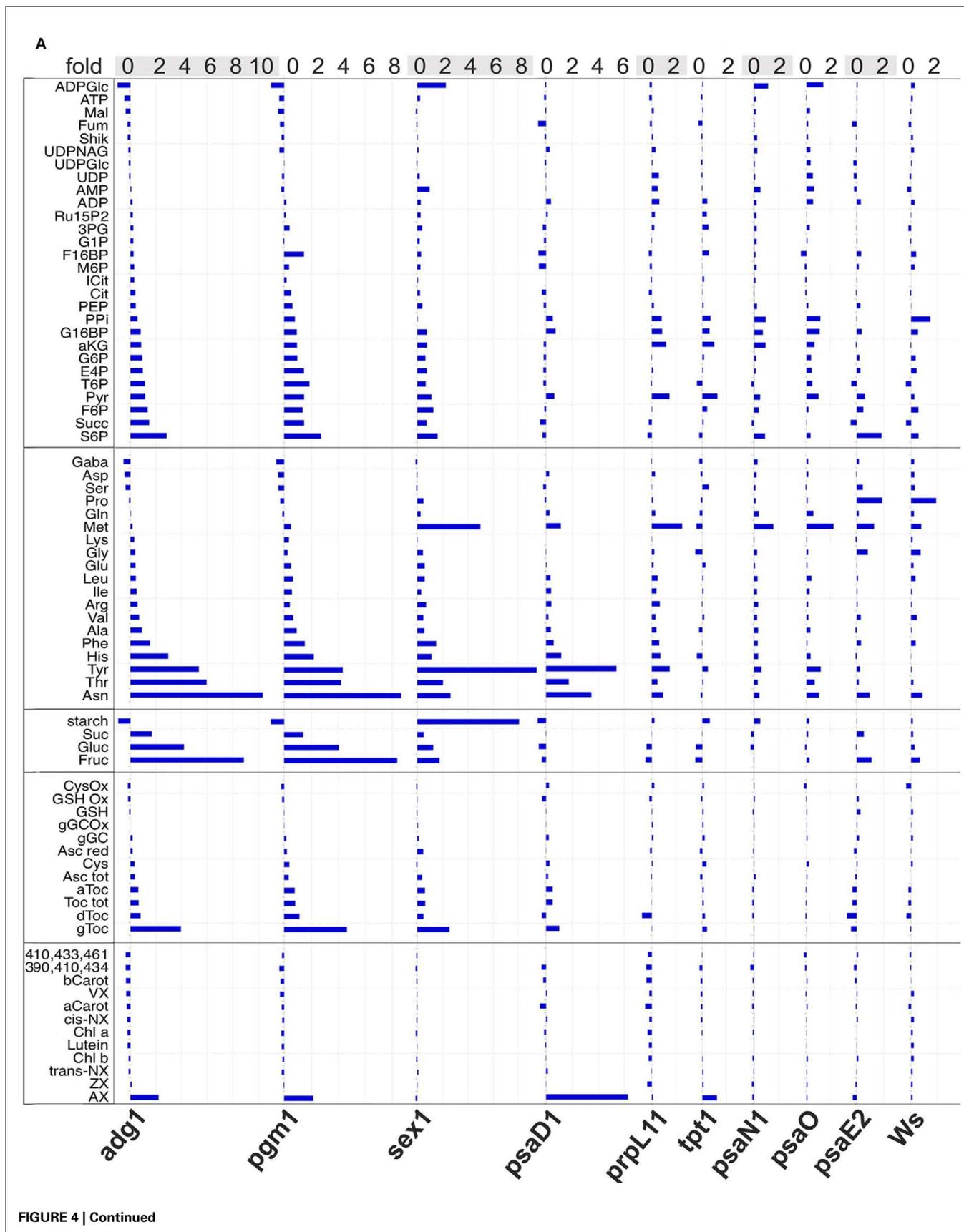
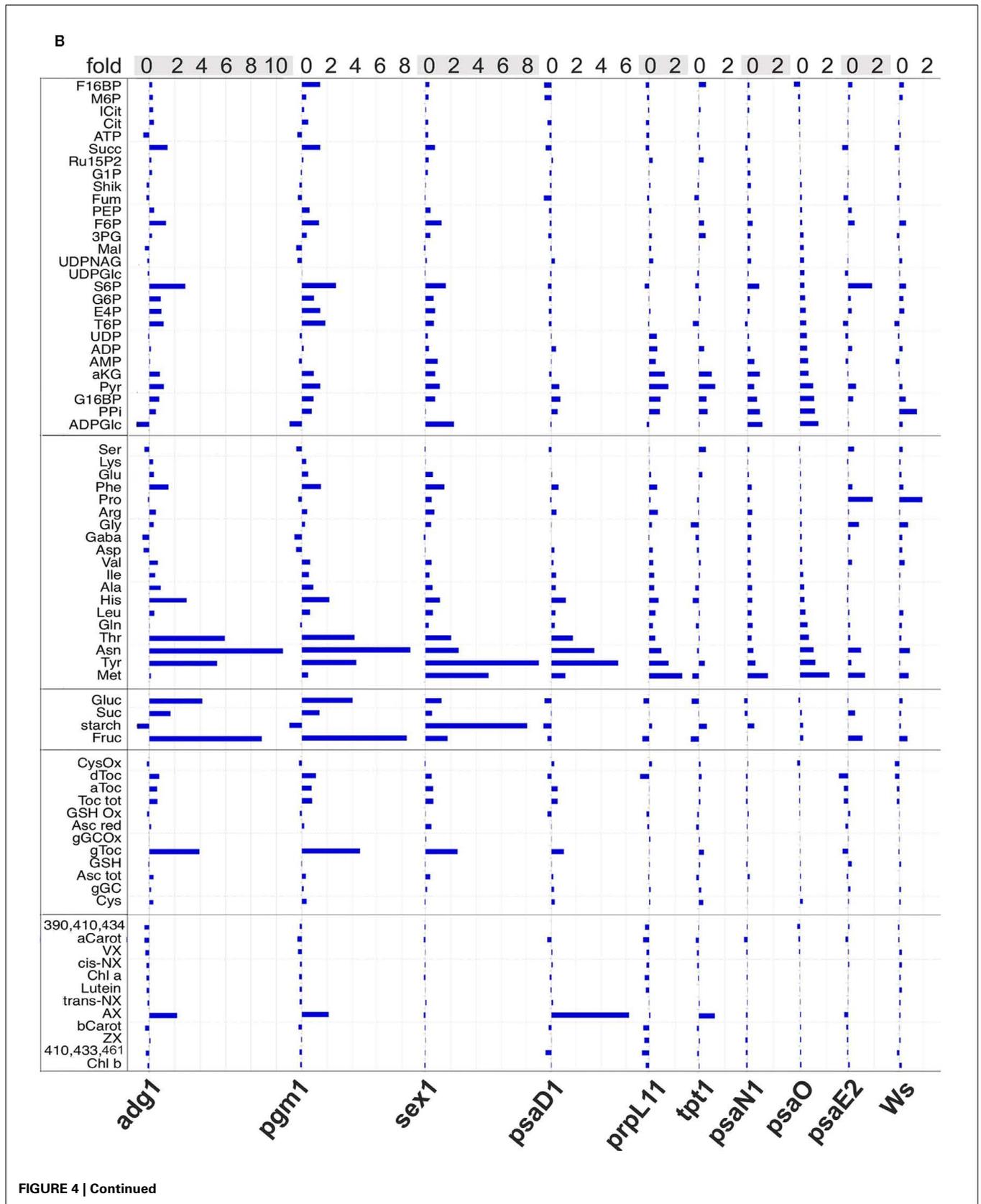


FIGURE 4 | Continued



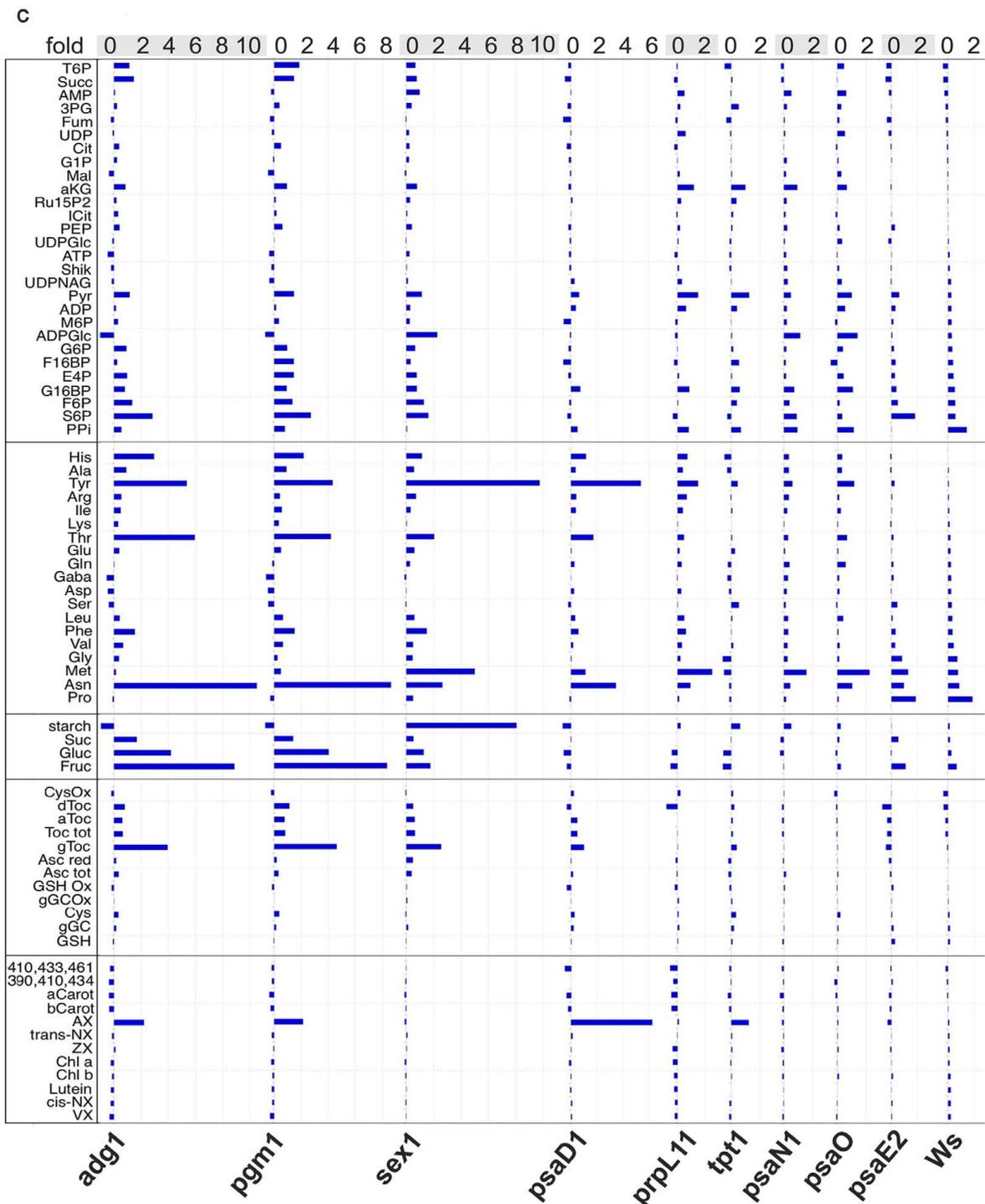


FIGURE 4 | Alignment of metabolic profiles measured for *Arabidopsis* mutants. Contents of metabolites are displayed as n-fold increase or decrease (top line) relative to the control Col0. The plant lines are named at the bottom. The five metabolic groups boxed at the left side contain from top to bottom phosphorylated metabolites (Pmet) and carboxylates, amino acids, sugars, antioxidants, and plant pigments. In A profiles are sorted from low to high amounts relative the mutant *adg1* (*adg1*-signature), enhancing the visualization of similarities. In (B) the

psaO- and in (C) the *Ws*-signature is displayed. The three most similar pairs of signatures detected were *adg1/pgm1* in (A), *psaO/psaN* in (B), and *Ws/psaE2* in (C). Columbia (Col0), Wassilewskija (*Ws*), phosphoglucomutase1 (*pgm1*); ADP-glucose pyrophosphorylase1 (*adg1*); starch-related alpha-glucan/water dikinase (*sex1*); plastidic triosephosphate/phosphate translocator (*tpt1*); defective in photosynthesis complex I (*psa*) subunits D1-E: *psaD1*, *psaO*, *psaN*, *psaE2*; defective in plastidic ribosomal protein L 11 (*prpL11*).

phosphorylated intermediates and carboxylates and the decrease of malate, which can be released from the depot of the vacuole (Gerhardt et al., 1987).

Unexpectedly, the carotenoid antheraxanthin (AX) was found to be in twofold excess, while besides zeaxanthin all other leaf pigments were decreased. Again this decrease might be a consequence from carbon mobilization. As processing and protection from light is essential and other pigments are missing, the deepoxidation reaction of the xanthophyll cycle toward AX, known as non-photochemical quenching (NPQ), might remain the only residual compensation of excess photosynthetic energy (Demmig-Adams and Adams III, 1996).

Starvation and light stress activates a protective antioxidant network (Golan et al., 2006). This could explain the observed increase of ascorbate, the tocopherols (Abbasi et al., 2007) and the stress indicative aromatic amino acids tyrosine (Schenke et al., 2011) and phenylalanine from which it is known, that they play an eminent role in abiotic stress and originate from plastids (Herrmann and Weaver, 1999; Ding et al., 2007; Rippert et al., 2009).

Sex1, the glucan/water dikinase mediates the phosphorylation of glucosyl residues at the surface of starch granules that is required for normal rates of starch breakdown. Therefore *sex1* mutants accumulate very high levels of starch (Yu et al., 2001; Kotting et al., 2005). We found the metabolic profile sorted next to the starchless mutants. In spite of the similar growth retardation we confirmed that certain metabolite contents were dramatically different as the increase of starch and ADP-glucose. The increase of soluble sugars and most amino acids again pointed to a massive mobilization of carbon. Although it is known, that blocking the metabolism of starch breakdown in *Arabidopsis* leaves triggers chloroplast degradation (Stettler et al., 2009), we could only measure little decreased leaf pigment levels and no AX accumulation. Thus the photosynthetic light reaction was not severely effected. This might explain a modified stress response compared to the starchless mutant in which tyrosine was even more (10-fold) increased and proline (Szabados and Savoure, 2009) and a fivefold-elevated methionine (Vogt, 1995) might serve as alternative protectors.

The *psaD1* protein is known to be essential for photosynthesis as it stabilizes the PSI complex and enables docking of ferredoxin (Jordan et al., 2001). The loss of PSI stops the photosynthetic electron flow and hence the energy supply of the whole plant. A second gene coding for the *psaD2* homolog rescues *psaD1* functions only partially generating 40% of the *psaD* protein. *psaD1* mutants consequently show a decrease in growth rate and down-regulation of most genes involved in the light phase of photosynthesis. A reduced synthesis of chlorophylls and carotenoids combined with an increased photosensitivity was observed (Haldrup et al., 2003; Ilnatowicz et al., 2004; Jensen et al., 2007). Accordingly we confirmed the stunted phenotype that resembled the three sugar mutants. The metabolic signature however appeared to be highly divers. The blocked loading of the Calvin cycle with NADPH and ATP explains the decrease of both, starch and hexoses, a status not detected for other candidates.

As seen for the *adg1/pgm1* pair, antheraxanthin again was elevated, while the chlorophylls and carotenoids were low. However here the elevation was the highest (fivefold) among all lines.

Similarly an increase of the xanthophyll cycle metabolites (VAZ) combined with decreased photosynthetic pigments in *psaD1* was observed before (Ilnatowicz et al., 2004). The additional elevation of most of the analyzed antioxidants and again tyrosine indicated that *psaD1* was stressed although the metabolic signature indicates a different less dramatic response.

For the remaining *Arabidopsis* lines, the metabolic differences found were less extreme. The two other PSI mutants, *psaO* and *psaN* showed further closely clustering metabolic profiles being more similar to *psaE2*, Col0, and Ws than to *psaD1*. This outcome correlates with the phenotypes, where *psaO* and *psaN* appeared a little smaller as the control but clearly larger than *psaD1*. The single copy gene *psaO* codes for a protein mainly involved in state transitions, forming a domain for LHCII binding and stabilization. It allows for responding to changes in illumination, balancing excitation pressure (Jensen et al., 2007). Therefore, under the light conditions applied it makes sense that this mutation was only of minor impact.

In *psaN* mutants the docking of the luminal electron donor plastocyanin to PSI is inefficient (Jensen et al., 2007). It was shown, that the lower activity of PSI is partially compensated by an increase in PSI complexes lacking *PsaN* (Haldrup et al., 1999).

In agreement with earlier findings (Varotto et al., 2000) in our hands the *psaE2* mutant in the Ws background displayed a wildtype-like growth and leaf coloration. Additionally we uncovered a wildtype-like metabolite profile indicating, that the loss of the E subunit of PSI under the applied growth conditions seems to be of minor importance for the plant. Accordingly earlier findings showed, that the E2 is not essential for PSI accumulation and thylakoid electron flow and although NADP⁺ photoreduction is impaired, mutants exhibit normal photoautotrophic growth (Zhao et al., 1993; Xu et al., 1994; Ilnatowicz et al., 2007). Most differences that we detected therefore must be regarded as ecotype specific, e.g., the remarkable fourfold elevation of proline versus Col0. The multifunctional proline is known to act as an osmolyte binding water conferring plants with a higher drought tolerance and tolerance for other abiotic stress (Verbruggen and Hermans, 2008; Szabados and Savoure, 2009). WS in particular is known to be more susceptible than other ecotypes to reactive oxygen species-induced damages (Kalbina and Strid, 2006; Li et al., 2006).

For the *tpt1* mutant it is known that one arm of the carbon dioxide partitioning via the Calvin cycle is blocked. As this reduces the export of photoassimilates and favors the rate of starch synthesis a starch excess phenotype as a secondary effect develops. The export block is fully compensated by accelerated starch turnover and export of neutral sugars. Instead of triosephosphate G6P, generated from exported Glucose, is used for cytosolic sucrose biosynthesis (Schneider et al., 2002). Besides this, *tpt1* is not the only abundant transporter of C3 molecules (Fischer et al., 1997). Accordingly the phenotype and metabolome data obtained confirmed that this knockout does not severely compromise *Arabidopsis*. Compared to the other starch mutants tested, the *tpt1* signature appeared to be weak. We could confirm the findings of Schneider et al. (2002) by measuring a nearly 100% reduction of hexoses and increase of starch, respectively. Furthermore, as seen for other candidates above, an increase in AX again seems

to be needed to outbalance excitation energy accumulated from reduced pigment levels.

Our *prpL11* cultivars confirmed earlier findings in which the phenotype developed pale green leaves together with a drastically reduced growth. In this mutant a component of the large 50s subunit of the plastidic ribosome is missing. As a consequence of impaired ribosomal activity the translation of the large subunit of Rubisco (RbcL) and of other plastidic proteins is significantly reduced resulting in severe defects of chloroplast function and a reduction of photosynthetic activity. (Pesaresi et al., 2001; Morita-Yamamuro et al., 2004). In accordance with this harsh intervention besides AX, we found all pigments to be reduced in this mutant. These reductions were higher than in all other candidates. The overall metabolic profile of *prpL11* is on one side more similar to the *tpt1* mutant and to *psaD1* as judged by HCA and untargeted PCA, but with respect to the amino acids it is like the *psaO/psaN* pair.

CERTAIN METABOLITES ARE INCREASED OR DECREASED IN ALL PLASTIDIC MUTANTS TESTED

Considering that each of the mutations assayed must have at least some impact on the plants metabolic status, the metabolites found to be increased or decreased in all mutants might be indicative for a common feature shared by the tested set of lines. What all mutants have in common is the compromised plastidic integrity. For the photosynthesis mutants normal light conditions might already be an overdose for the plastidic light harvesting function. The blocks of the plastidic carbon assimilation, utilization or export functions of the other mutants likewise results in organelles with adapted metabolism generating a shortage or reallocation of carbon. Energy- or carbon-shortage both as a consequence are generating more or less stress depending on the impact of the mutation. A common stress response, e.g., in combination with the protective antioxidant network (Golan et al., 2006) for compromised plastids might explain, why some stress indicative metabolites as γ -glutamylcysteine, phenylalanine, tyrosine, isoleucine, or threonine are increased in all lines.

Outbalancing of the hampered organelles might additionally trigger adaptations of carbon mobilization or cause accumulation at certain metabolic crossings in the mutant leaves. An example would be an increased desamination of amino acids or a reduced NO_3^- assimilation. Today there is little doubt that C-metabolites are important signal molecules acting at various regulatory steps of the NO_3^- assimilation pathway (Lam et al., 1996). The increased alpha-ketoglutarate (2 OG) together with the elevated glutamate might be indicative here. Pyruvate as well was found to be increased in all mutants. As a precursor for the likewise increased valine and leucine and for acetyl-CoA, this central metabolite

might not be utilized to a normal level in plastidic mutants. Here again the carbon-shortage might generate a feedback on the precursor levels from unneeded lipid- and isoprenoid biosynthesis producing less cell membranes, chlorophyll, or carotenoids. Another adaption of the mutants might be indicated by the accumulated ribulose-1,5-bisphosphate (R15BP). Again the reduced plastidic functions of carbon flow or photosynthetic yield could generate a down-regulation of the Calvin cycle.

Our data show that we can detect similar metabolic patterns for mutants with related defects.

The pleiotropic responses allow identifying such homologous signatures out of a set of mutants. It is now challenging to continue in generating further metabolic profiles for mutants with defects in different and more distant pathways as well as, e.g., for different ecotypes. It should furthermore be tested, if certain parameter of cultivation, e.g., the time-point of harvest or the intensity of light influences the separation of the mutants. As certain metabolite levels dramatically change during the day and under different external parameters, it might be useful to find optimized conditions for profiling. The separation of mutants might as well be largely independent of such parameters, as long as all candidates are treated in the same way. This would probably simplify future experimental designs. An established spectrum of metabolic signatures should finally enable the classification of unknown plant mutants into functional groups.

ACKNOWLEDGMENTS

The authors would like to thank Christine Hösl at the FAU Erlangen-Nuremberg for plant care and Urte Schlüter for useful discussions.

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at http://www.frontiersin.org/Plant_Physiology/10.3389/fpls.2011.00082/abstract

Table S1 | Compilation of metabolite data of the targeted metabolite analysis content. Measured data from 74 metabolites analyzed in targeted assays are given as mean values of four independent replicates \pm SD. Significantly different metabolite contents between genotypes and Col-0 with $P < 0.05$ according to a Welch-Satterthwaite test (Junker et al., 2006) are indicated by an asterisk. Columbia (Col-0), Wassilewskija (Ws), phosphoglucomutase 1 (*pgm1*); ADP glucose pyrophosphorylase 1 (*adg1*); starch-related alpha-glucan/water dikinase (*sex1*); plastidic triose phosphate/phosphate translocator (*tpt1*); defective in photosynthesis complex I (*psa*) subunits D1-E: *psaD1*, *psaO*, *psaN*, *psaE2*; defective in plastidic ribosomal protein L 11 (*prpL11*).

Table S2 | Loadings of the targeted PCA. Values for metabolites loading on the PCA for the targeted analysis of 11 *Arabidopsis* lines (Figure 3B) are sorted from high to low and given for the principal components PC1 (column 5) and PC2 (column 10).

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

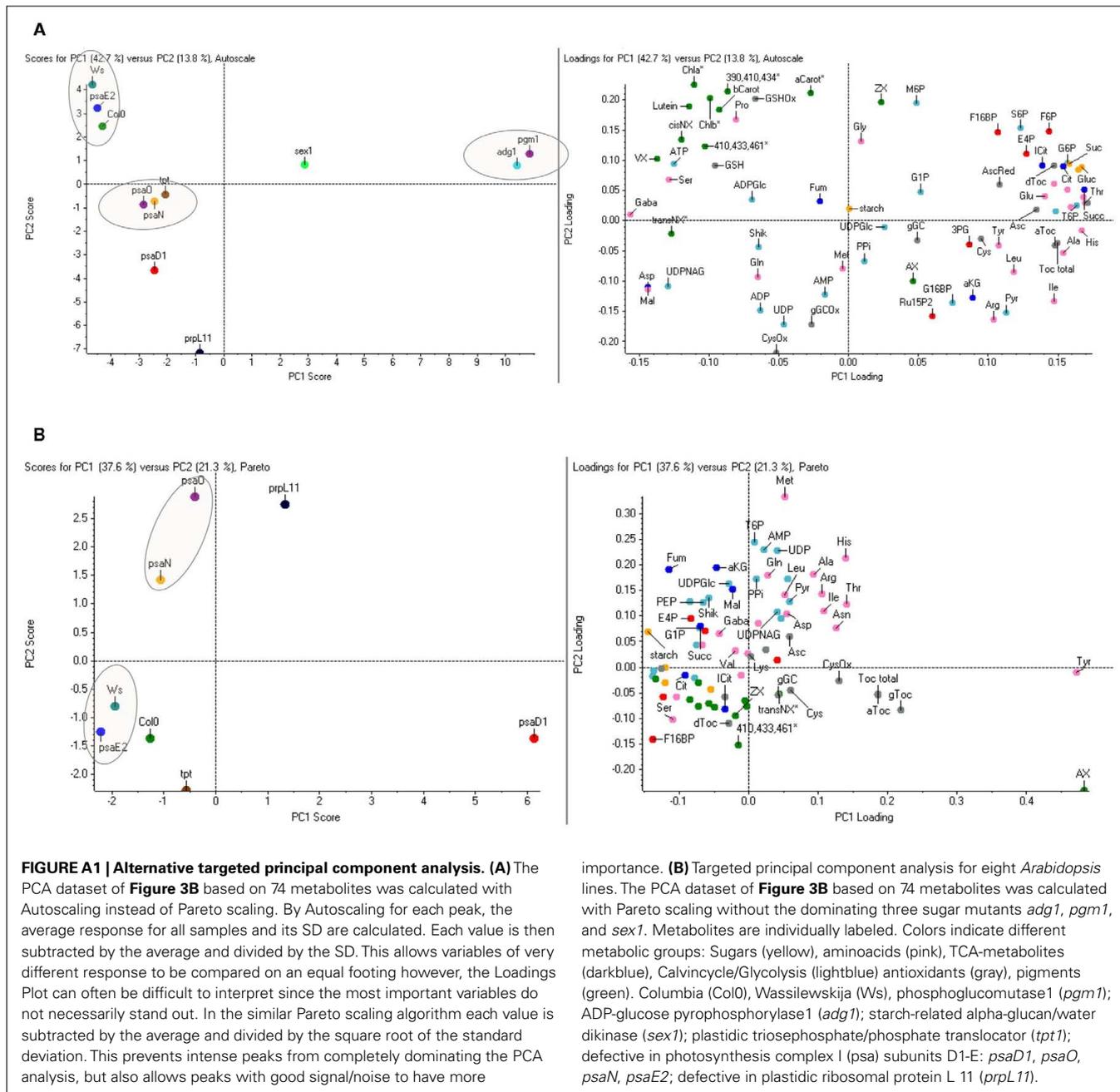
Received: 20 October 2011; accepted: 02 November 2011; published online: 23 November 2011.

Citation: Hofmann J, Börnke F, Schmiedel A, Kleine T and Sonnewald U (2011) Detecting functional groups of *Arabidopsis* mutants by metabolic profiling and evaluation of pleiotropic responses. *Front. Plant Sci.* 2:82. doi: 10.3389/fpls.2011.00082

This article was submitted to *Frontiers in Plant Physiology*, a specialty of *Frontiers in Plant Science*.

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APPENDIX



REFERENCE

Junker, B. H., Klukas, C., and Schreiber, F. (2006). VANTED: A system for advanced data analysis and visualization in the context of biological networks. *BMC Bioinformatics* 7:109.