

Prevalence and influence on outcome of *HER2/neu*, *HER3* and *NRG1* expression in patients with metastatic colorectal cancer

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Our aim was to explore the impact of the *HER2/neu*, *HER3* receptor as well as their ligands' neuregulin (*NRG1*) expression on the outcome of patients with metastatic colorectal cancer (mCRC). *NRG1*, *HER2/neu* and *HER3* expression was evaluated in 208 patients with mCRC receiving 5-FU/LV plus irinotecan or irinotecan plus oxaliplatin as the first-line treatment. Biomarker expression was correlated with the outcome of patients. *NRG1* (low: 192 vs. high: 16), *HER2/neu* (low: 201 vs. high: 7) and *HER3* (low: 69 vs. high: 139) expressions were assessed in 208 patients. High versus low *NRG1* expression significantly affected progression-free survival (PFS) [4.7 vs. 8.2 months, hazard ratio (HR): 2.45; 95% confidence interval (CI): 1.45–4.13; $P=0.001$], but not overall survival (OS) (15.5 vs. 20.7 months, HR: 1.33; 95% CI: 0.76–2.35; $P=0.32$). High versus low *HER3* expression (PFS: 7.1 vs. 8.8 months, HR: 1.11; 95% CI: 0.82–1.50; $P=0.50$; OS: 19.8 vs. 21.1 months, HR: 0.95; 95% CI: 0.70–1.30; $P=0.75$) and high compared with low *HER2/neu* expression (PFS: 7.7 vs. 8.0 months, HR: 1.07; 95% CI: 0.71–1.60; $P=0.75$; OS: 16.6 vs. 21.1 months, HR: 1.13; 95% CI: 0.75–1.71; $P=0.57$) did not influence outcome. High *NRG1* expression was associated with

inferior PFS in the FIRE-1 trial. We did not detect a prognostic impact of *HER2/neu* and *HER3* overexpression in mCRC. The frequency of overexpression was comparable with other studies. *Anti-Cancer Drugs* 28:717–722 Copyright © 2017 Wolters Kluwer Health, Inc. All rights reserved.

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Introduction

Treatment of metastatic colorectal cancer (mCRC) has improved since molecular biomarkers are being evaluated for their predictive and prognostic information, such as *RAS* mutations for epidermal growth factor receptor (EGFR)-targeted treatment [1,2]. Together with the *HER2/neu*, *HER3* and *HER4* receptor, the EGFR belongs to the HER receptor family [3]. These receptors have tyrosine kinase activity. When activated, the receptors engage intracellular signalling pathways leading to proliferation [4–8].

HER2/neu overexpression was identified as valuable target in subpopulations of breast cancer and gastric cancer [9–11]. The HERACLES trial has further identified

HER2/neu overexpression as a targetable structure in a subset of patients with *KRAS* wild-type mCRC [12]. Besides this potential predictive relevance, the unfavourable outcome of colorectal cancer (CRC) patients has been associated with *HER2/neu* overexpression in previous analyses [13,14].

Potentially treatment-relevant expression of the *HER3* receptor has been reported in several solid cancer types [15]. If neuregulin (*NRG1*) binds to the *HER3* receptor, *HER3* forms heterodimers with the *HER2/neu* receptor [6]. Subsequently, the *PI3K-AKT* and *MAPK* pathways are activated, which stimulate tumour proliferation [6,15]. In the literature, *HER3* expression rates in CRC cells range from 34 to 90% [15]. However, it is not clear if

HER3 expression is associated with patients outcome in colorectal cancer. *HER3* overexpression was reported to be a negative prognostic marker for patients with CRC without distant metastases [16–18].

In addition, previous investigations focused on the role of *NRG1* in CRC as being the activating ligand of the *HER3* receptor. De Boeck *et al.* [19] found tumour progression to be highly influenced by bone marrow-derived mesenchymal stem cells releasing *NRG1* *in vitro* and *in vivo*. Furthermore, two investigations indicated a potential role for predicting lymph node involvement and the occurrence of distant metastases [20–22]. Nevertheless, two cohorts of advanced CRC and one study of a cohort of CRC patients with distant metastases did not confirm effects on outcome [23–25]. Coalteration of *HER2/neu* and *HER3* expression was found rarely, also without impact on the outcome of patients with CRC [23].

This analysis was designed to confirm the prevalence and prognostic impact of *HER2/neu*, *HER3* and *NRG1* expression in a chemotherapy-based study cohort of 208 patients with mCRC (FIRE-1 trial) receiving either 5-FU/LV plus irinotecan or irinotecan plus oxaliplatin as first-line therapy. To our knowledge, the FIRE-1 trial is the first randomized-controlled trial to investigate the impact of *HER2/neu* and *HER3* receptor overexpression in relation to *NRG1* expression [16–18,20–24].

Methods

Study design and treatment schedule

FIRE-1 was a multicentre phase III study. The protocol, primary results and characteristics of patients have been published previously [26]. Also, details on the sub-population evaluable for translational research have been reported [27]. Information on *RAS* mutation status and EGFR ligand expression was available for the cohort as described previously [27].

Patients

Our analysis included 208 of 479 patients with available tumour material [27] for the analysis of *HER2/neu*, *HER3* and *NRG1* expression.

Ethics

The trial was conducted in accordance with the Declaration of Helsinki (1996). All patients provided written informed consent to be treated within a clinical trial. This investigation was performed as a retrospective evaluation with the approval of the local ethics committee of the University of Munich (registry-number: 545-11).

End points

For this manuscript, overall survival (OS) (time from randomization to death), progression-free survival (PFS) (interval between randomization and death or progression) and response rate (WHO classification: complete remission, partial remission, no change, progressive

disease) were used to correlate molecular characteristics with the outcome of patients of the FIRE-1 trial.

Immunohistochemistry

Immunohistochemistry was performed using 5 µm whole standard tissue sections of FFPE tumour samples. For the detection of *HER2/neu*, a prediluted anti-*HER2/neu* rabbit monoclonal antibody (clone 4B5; Ventana Medical Systems, Oro Valley, Arizona, USA) was used as the primary antibody. The staining was performed on a Ventana Benchmark XT autostainer using the XT UltraView diaminobenzidine kit (Ventana Medical Systems) following the manufacturer's protocols. Staining of *HER3* and *NRG1* was performed using the Vectastain ABC-Kit Elite Universal detection system (Vector Laboratories, Peterborough, UK). For *HER3* immunohistochemistry, a monoclonal rabbit antibody was used as the primary antibody (ab93739; Abcam, Cambridge, UK). *NRG1* staining was performed using a polyclonal human antibody (HPA010964; Atlas Antibodies, Stockholm, Sweden).

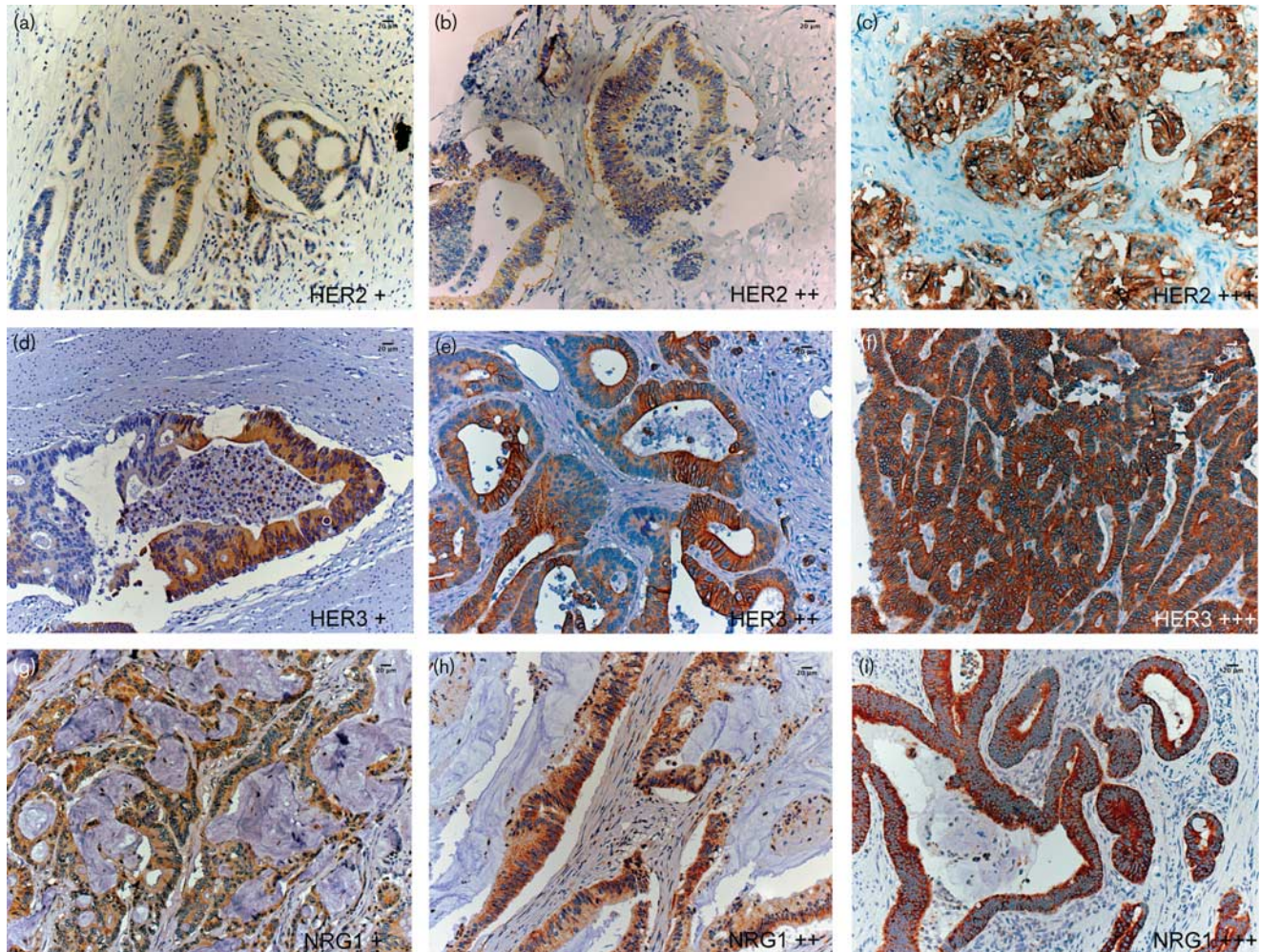
Scoring of high and low expression, FISH

As no evident and standardized method for dividing high and low expression of *HER2/neu*, *HER3* and *NRG1* existed at the time of evaluation, we used the *HER2/neu* score of Rüschoff and colleagues in gastric cancer for scoring complete biomarker expression. Therefore, membrane staining was graduated by intensity (0: none, 1+: weak, 2+: moderate, 3+: strong) and percentage of stained tumour cells (Fig. 1). High expression was defined by a percentage of more than 10% stained tumour cells and at least moderate (2+ or 3+) membrane staining versus no or weak staining (0 or 1+) for low expression. In addition, two-colour fluorescence in-situ hybridization (FISH) was performed in patients showing moderate (2+) *HER2/neu* staining. Chromosome 17 centromere signals (green) as well as *HER2* gene signals (red) were counted in at least 20 nuclei of colorectal tumour cells. Thus, a red to green ratio of at least 2 indicated amplification of *HER2*. Primary tumour slides were evaluated by two independent observers (A.S. and J.N.) using a light microscope. Disagreements (<5%) were reviewed together, followed by conclusive judgement.

Statistical analysis

OS and PFS stratified by the molecular markers were estimated using Kaplan–Meier analysis. Significant differences were evaluated using the log-rank test and Cox regression analysis. Univariate Cox regression was performed in subgroups. The correlation of clinicopathologic parameters with biomarker expression was assessed using the χ^2 -test and the Fisher exact test for nominal variables. All *P*-values of less than 0.05 (two sided) were considered significant. SPSS PASW 18.0 (SPSS Inc., Chicago, Illinois, USA) was used for statistical analysis.

Fig. 1



Immunohistochemical staining intensity of *HER2/neu* expression [+ : weak (a), ++ : intermediate (b), +++ : strong (c)], *HER3* expression [+ : weak (d), ++ : intermediate (e), +++ : strong (f)] and *NRG1* expression [+ : weak (g), ++ : intermediate (h), +++ : strong (i)]. NRG, neuregulin.

Results

Study population

HER2/neu, *HER3* and *NRG1* analyses were carried out in 208 tumours. Characteristics of the entire patient population and the evaluable subpopulation have been published before [27]. According to baseline and tumour characteristics as well as PFS and OS, the subpopulation was well comparable with the entire study population [27].

Prevalence of high *NRG1*, *HER3* and *HER2/neu* expression

Of 208 metastatic colorectal tumours in total, high *NRG1* expression was detected in 16 (7.7%) specimens. 139 of 208 tumours (67%) were diagnosed to have *HER3* overexpression. Twenty-three (11.1%) patients showed moderate *HER2/neu* staining. A subsequent FISH analysis, however, showed a missing gene amplification in all

of these 23 patients (*HER2/neu*: chromosome 17 ratio <2.0). Therefore, only strong (3+) stainings in seven (3.3%) patients were accepted as high *HER2/neu* expression.

Correlation of *NRG1*, *HER3* and *HER2/neu*

A significant correlation of biomarkers with each other could not be detected for *NRG1* and *HER2/neu* ($P=1.00$) or for *NRG1* and *HER3* ($P=0.41$). High *HER2/neu* expression also did not correlate with *HER3* overexpression ($P=0.43$; Table 1).

Association of *NRG1*, *HER3* and *HER2/neu* expression with *RAS* mutations and EGFR-ligand expression

HER3 overexpression was significantly correlated with the presence of *RAS* mutations ($P=0.02$; Table 2). *HER3* overexpression showed a trend towards an association

Table 1 Correlation of *HER2/neu* and *HER3* overexpression with *NRG1* expression and coalteration of *HER2/neu* and *HER3* expression

Correlation of <i>HER2/neu</i> and <i>HER3</i> overexpression with <i>NRG1</i> expression				
	Neuregulin 1 [n (%)]		Total [n (%)]	P (two sided)
	Low	High		
<i>HER2/neu</i>				
Low	185 (89.0)	16 (7.7)	201 (96.7)	1.00
High	7 (3.3)	0 (0.0)	7 (3.3)	
Total	192 (92.3)	16 (7.7)	208 (100.0)	
<i>HER3</i>				
Low	62 (29.8)	7 (3.4)	69 (33.2)	0.41
High	130 (62.5)	9 (4.3)	139 (66.8)	
Total	192 (92.3)	16 (7.7)	208 (100.0)	
Coalteration of <i>HER2/neu</i> and <i>HER3</i> expression				
	<i>HER2/neu</i> [n (%)]		Total [n (%)]	P (two sided)
	Low	High		
<i>HER3</i>				
Low	68 (32.7)	1 (0.5)	69 (33.2)	0.43
High	133 (64.0)	6 (2.8)	139 (66.8)	
Total	201 (96.7)	7 (3.3)	208 (100.0)	

P-values calculated using the χ^2 -test.
NRG, neuregulin.

Table 2 Correlation of *HER3* expression and *RAS* status

<i>RAS</i>	<i>HER3</i> [n (%)]		Total [n (%)]	P (two sided)
	Low	High		
Wild type	42 (20.2)	59 (28.4)	101 (48.6)	0.02
Mutation	27 (12.9)	80 (38.5)	107 (51.4)	
Total	69 (33.1)	139 (66.9)	208 (100.0)	

P-values calculated using the χ^2 -test.

with high *EREG* expression. ($P=0.07$). All other combinations did not show associations.

Survival analysis

High versus low *NRG1* expression significantly affected PFS (4.7 vs. 8.2 months, hazard ratio: 2.45; 95% confidence interval: 1.45–4.13; $P=0.001$), but not OS (15.5 vs. 20.7 months, hazard ratio: 1.33; 95% confidence interval: 0.76–2.35; $P=0.32$). *HER3* and *Her2/neu* expression did not influence outcome (Fig 2a–d).

Discussion

To our knowledge, the FIRE-1 trial is the first randomized-controlled trial to investigate the impact of *HER2/neu* and *HER3* receptor overexpression in relation to *NRG1* expression and *RAS* status in mCRC [16–18, 20–24]. Previous investigations focused mostly on analysing *HER2/neu* and *HER3* expression in advanced colorectal or rectal tumours with patients receiving adjuvant radiotherapy or chemotherapy [17,18,20–22]. The FIRE-1 treatment schedule consisted of 5-FU/LV plus irinotecan or oxaliplatin plus irinotecan as first-line therapy, following a recommended second-line therapy

with the respective crossover study regimen. With 208 patients enrolled in the analysis, our trial represents a robust investigation [16–18,20–24].

Evaluation of *HER2/neu* expression has been established in gastric cancer by Rüschoff *et al.* [9] using immunohistochemical staining and FISH in intermediate cases. Therefore, we decided to use these validated methods for patients with mCRC in accordance with the literature. *HER3* expression was also evaluated by immunohistochemical stainings using a modified Rüschoff semiquantitative scoring system, as in previous investigations, defining overexpression by cytoplasmatic or membrane staining intensity [16,18,23, 24]. As scoring of *NRG1* expression is not yet standardized for any tumour type [19], we also used an adapted Rüschoff score.

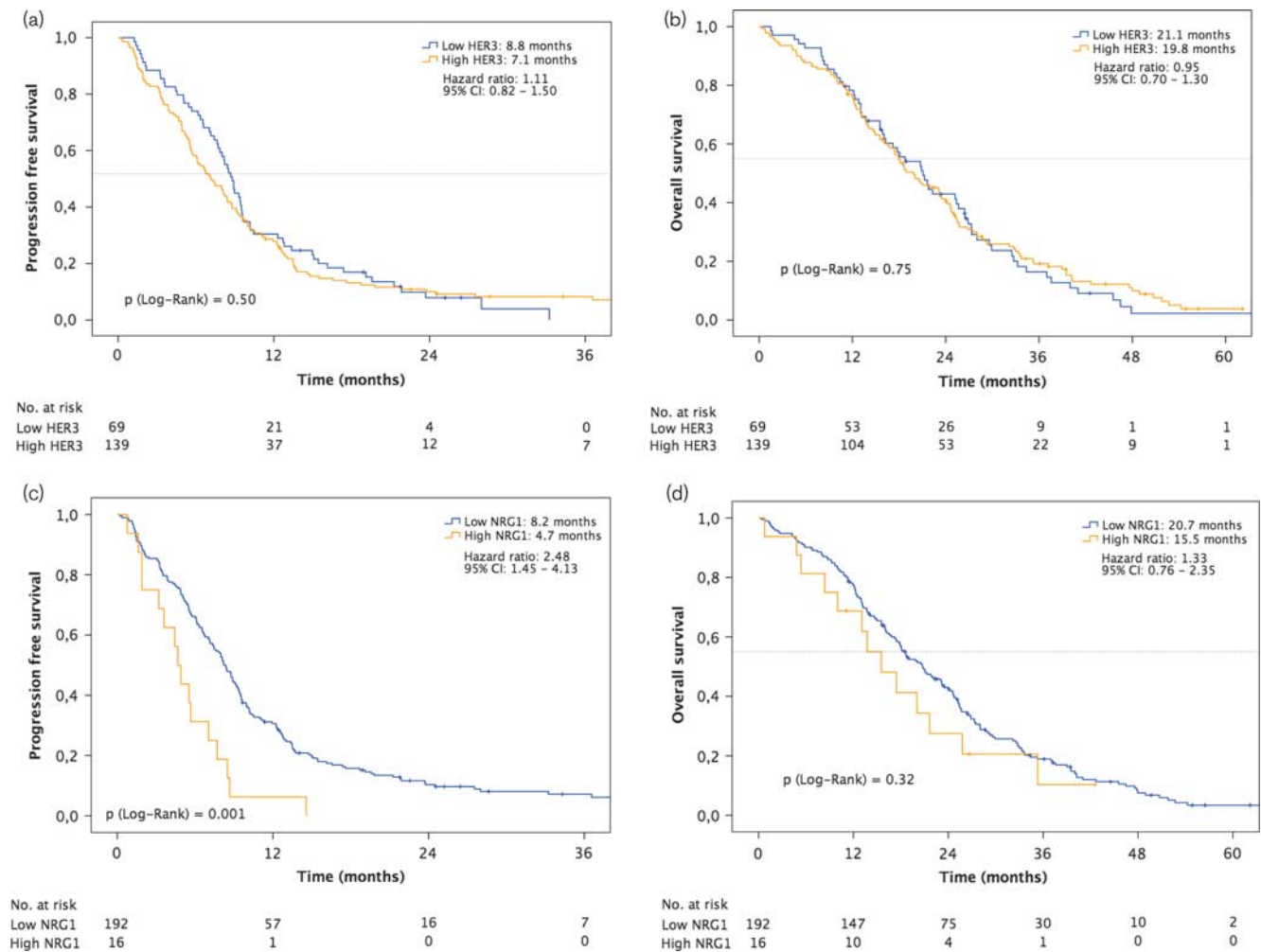
In FIRE-1, 3.3% of tumours showed high *HER2/neu* expression. This compares favourably to the average expression rate of *HER2/neu* in the literature of ~5.0% [12,13,23]. In FIRE-1, 67% of primary tumours showed high *HER3* expression, comparing favourably to recent other cohorts (Seo *et al.* [23]: 69%; Lédél *et al.* [18,22]: 70%). One trial evaluating transmembrane *NRG1* expression in CRC reported a frequency of high expression of 76%, in contrast to 8% in FIRE-1 [19]. However, a standardized scoring system is missing in this case, which may explain the discrepancy.

We attempted to correlate expression rates of *HER* receptors with *HER* ligands (*NRG1*) as well as downstream molecules (*RAS*). Unlike a previous investigation, we could not associate *HER3* expression with *NRG1* expression [19]. In our investigation, regular simultaneous expression of *HER2/neu* and *HER3* was also not detected, although coexpression of *HER2/neu* and *HER3* has been described in cohort of 364 surgically resected CRC patients [28]. The latter discrepancy might be caused in part by the different clinical backgrounds of patients as well as by the different diagnostic methods used.

By contrast, in our cohort, *HER3* expression correlated with *RAS* mutations, although *HER3* expression could not be associated with *KRAS* mutations previously [21]. This observation might have resulted from a higher number of patients enrolled in our investigation in addition to an extended analysis of *RAS* mutations.

In our study, high *NRG1* expression led to a significant decrease in PFS. This finding is supported by a previous investigation that also observed significantly worse 5-year PFS in patients with mCRC and high *NRG1* expression [19]. Because of the small numbers of patients showing high *NRG1* expression, conclusions are limited. The sample size of patients with *HER2/neu* overexpression in our cohort does not allow for conclusions on the prognostic impact. It is noteworthy that *HER3* overexpression

Fig. 2



Outcomes according to subgroups in FIRE-1; (a): PFS of patients in FIRE-1 comparing low and high *HER3* expression. (b): OS of patients in FIRE-1 comparing low and high *HER3* expression. (c): PFS of patients in FIRE-1 comparing low and high *NRG1* expression. (d): OS of patients in FIRE-1 comparing low and high *NRG1* expression. CI, confidence interval; NRG, neuregulin; OS, overall survival; PFS, progression-free survival.

was not associated with an unfavourable outcome in FIRE-1, although conflicting data may exist [16–18,24, 28]. However, the small number of patients enrolled in some trials limits conclusions.

Our investigation had several strengths. FIRE-1 was a randomized-controlled trial with irinotecan-based treatment, which had a rather small likelihood of bias in terms of the outcome and follow-up information. Unfortunately, although 208 patients were enrolled in this investigation, which represented a rather robust sample size, numbers in subgroups (*NRG1*, *Her2/neu*) became small. It might also be argued that treatment in FIRE-1 does not comply with the latest recommendations. In addition, FIRE-1, as well as previous investigations, lacked a validation collective for a proof-of-principle analysis. Therefore, further research is warranted to evaluate prognostic effects.

Conclusion

A significant unfavourable impact on PFS was observed in patients with mCRC with a high *NRG1* expression in the FIRE-1 trial. We did not detect a prognostic impact of *HER2/neu* and *HER3* overexpression in mCRC with respect to PFS and OS.

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Conflicts of interest

D.P.M. received a research grant from the Weigand-Bohnewand-Gravenhorst-Fonds for this project. For the remaining authors there are no conflicts of interest.

References

- 1 Douillard JY, Siena S, Cassidy J, Tabernero J, Burkes R, Barugel M, *et al.* Randomized, phase III trial of panitumumab with infusional fluorouracil, leucovorin, and oxaliplatin (FOLFOX4) versus FOLFOX4 alone as first-line treatment in patients with previously untreated metastatic colorectal cancer: the PRIME study. *J Clin Oncol* 2010; **28**:4697–4705.
- 2 Van Cutsem E, Kohne CH, Lang I, Folprecht G, Nowacki MP, Cascinu S, *et al.* Cetuximab plus irinotecan, fluorouracil, and leucovorin as first-line treatment for metastatic colorectal cancer: updated analysis of overall survival according to tumor KRAS and BRAF mutation status. *J Clin Oncol* 2011; **29**:2011–2019.
- 3 Leahy DJ. Structure and function of the epidermal growth factor (EGF/ErbB) family of receptors. *Adv Protein Chem* 2004; **68**:1–27.
- 4 Pinkas-Kramarski R, Shelly M, Guarino BC, Wang LM, Lyass L, Alroy I, *et al.* ErbB tyrosine kinases and the two neuregulin families constitute a ligand-receptor network. *Mol Cell Biol* 1998; **18**:6090–6101.
- 5 Kani K, Warren CM, Kaddis CS, Loo JA, Landgraf R. Oligomers of ERBB3 have two distinct interfaces that differ in their sensitivity to disruption by heregulin. *J Biol Chem* 2005; **280**:8238–8247.
- 6 Hellyer NJ, Kim MS, Koland JG. Heregulin-dependent activation of phosphoinositide 3-kinase and Akt via the ErbB2/ErbB3 co-receptor. *J Biol Chem* 2001; **276**:42153–42161.
- 7 Dawson JP, Berger MB, Lin CC, Schlessinger J, Lemmon MA, Ferguson KM. Epidermal growth factor receptor dimerization and activation require ligand-induced conformational changes in the dimer interface. *Mol Cell Biol* 2005; **25**:7734–7742.
- 8 Berger MB, Mendrola JM, Lemmon MA. ErbB3/HER3 does not homodimerize upon neuregulin binding at the cell surface. *FEBS Lett* 2004; **569**:332–336.
- 9 Rüschoff J, Hanna W, Bilous M, Hofmann M, Osamura RY, Penault-Llorca F, *et al.* HER2 testing in gastric cancer: a practical approach. *Mod Pathol* 2012; **25**:637–650.
- 10 Rüschoff J, Dietel M, Baretton G, Arbogast S, Walch A, Monges G, *et al.* HER2 diagnostics in gastric cancer-guideline validation and development of standardized immunohistochemical testing. *Virchows Arch* 2010; **457**:299–307.
- 11 Gown AM, Goldstein LC, Barry TS, Kussick SJ, Kandalaf PL, Kim PM, *et al.* High concordance between immunohistochemistry and fluorescence in situ hybridization testing for HER2 status in breast cancer requires a normalized IHC scoring system. *Mod Pathol* 2008; **21**:1271–1277.
- 12 Valtorta E, Martino C, Sartore-Bianchi A, Penault-Llorca F, Viale G, Riso M, *et al.* Assessment of a HER2 scoring system for colorectal cancer: results from a validation study. *Mod Pathol* 2015; **28**:1481–1491.
- 13 Ingold Heppner B, Behrens HM, Balschun K, Haag J, Kruger S, Becker T, *et al.* HER2/neu testing in primary colorectal carcinoma. *Br J Cancer* 2014; **111**:1977–1984.
- 14 Lee WS, Park YH, Lee JN, Baek JH, Lee TH, Ha SY. Comparison of HER2 expression between primary colorectal cancer and their corresponding metastases. *Cancer Med* 2014; **3**:674–680.
- 15 Sithanandam G, Anderson LM. The ERBB3 receptor in cancer and cancer gene therapy. *Cancer Gene Ther* 2008; **15**:413–448.
- 16 Beji A, Horst D, Engel J, Kirchner T, Ullrich A. Toward the prognostic significance and therapeutic potential of HER3 receptor tyrosine kinase in human colon cancer. *Clin Cancer Res* 2012; **18**:956–968.
- 17 Scartozzi M, Mandolesi A, Giampieri R, Bittoni A, Pierantoni C, Zaniboni A, *et al.* The role of HER-3 expression in the prediction of clinical outcome for advanced colorectal cancer patients receiving irinotecan and cetuximab. *Oncologist* 2011; **16**:53–60.
- 18 Lédel F, Hallstrom M, Ragnhammar P, Ohrling K, Edler D. HER3 expression in patients with primary colorectal cancer and corresponding lymph node metastases related to clinical outcome. *Eur J Cancer* 2014; **50**:656–662.
- 19 De Boeck A, Pauwels P, Hensen K, Rummens JL, Westbroek W, Hendrix A, *et al.* Bone marrow-derived mesenchymal stem cells promote colorectal cancer progression through paracrine neuregulin 1/HER3 signalling. *Gut* 2013; **62**:550–560.
- 20 Grivas PD, Antonacopoulou A, Tzelepi V, Sotiropoulou-Bonikou G, Kefalopoulou Z, Papavassiliou AG, *et al.* HER-3 in colorectal tumorigenesis: from mRNA levels through protein status to clinicopathologic relationships. *Eur J Cancer* 2007; **43**:2602–2611.
- 21 Ho-Pun-Cheung A, Assenat E, Bascoul-Mollevi C, Bibeau F, Boissiere-Michot F, Cellier D, *et al.* EGFR and HER3 mRNA expression levels predict distant metastases in locally advanced rectal cancer. *Int J Cancer* 2011; **128**:2938–2946.
- 22 Lédel F, Stenstedt K, Hallstrom M, Ragnhammar P, Edler D. HER3 expression in primary colorectal cancer including corresponding metastases in lymph node and liver. *Acta Oncol* 2015; **54**:480–486.
- 23 Seo AN, Kwak Y, Kim DW, Kang SB, Choe G, Kim WH, *et al.* HER2 status in colorectal cancer: its clinical significance and the relationship between HER2 gene amplification and expression. *PLoS one* 2014; **9**:e98528.
- 24 Kountourakis P, Pavlakis K, Psyrris A, Rontogianni D, Xiros N, Patsouris E, *et al.* Prognostic significance of HER3 and HER4 protein expression in colorectal adenocarcinomas. *BMC cancer* 2006; **6**:46.
- 25 Uner A, Ebinc FA, Akyurek N, Unsal D, Menten BB, Dursun A. Vascular endothelial growth factor, c-erbB-2 and c-erbB-3 expression in colorectal adenoma and adenocarcinoma. *Exp Oncol* 2005; **27**:225–228.
- 26 Fischer von Weikersthal L, Schalhorn A, Stauch M, Quietzsch D, Maubach PA, Lambert H, *et al.* Phase III trial of irinotecan plus infusional 5-fluorouracil/folinic acid versus irinotecan plus oxaliplatin as first-line treatment of advanced colorectal cancer. *Eur J Cancer* 2011; **47**:206–214.
- 27 Stahler A, Heinemann V, Giessen-Jung C, Crispin A, Schalhorn A, Stintzing S, *et al.* Influence of mRNA expression of epiregulin and amphiregulin on outcome of patients with metastatic colorectal cancer treated with 5-FU/LV plus irinotecan or irinotecan plus oxaliplatin as first-line treatment (FIRE 1-trial). *Int J Cancer* 2016; **138**:739–746.
- 28 Seo AN, Kwak Y, Kim WH, Kim DW, Kang SB, Choe G, *et al.* HER3 protein expression in relation to HER2 positivity in patients with primary colorectal cancer: clinical relevance and prognostic value. *Virchows Arch* 2015; **466**:645–654.