





Involvement of astrocyte and oligodendrocyte gene sets in migraine

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Abstract

Background: Migraine is a common episodic brain disorder characterized by recurrent attacks of severe unilateral headache and additional neurological symptoms. Two main migraine types can be distinguished based on the presence of aura symptoms that can accompany the headache: migraine with aura and migraine without aura. Multiple genetic and environmental factors confer disease susceptibility. Recent genome-wide association studies (GWAS) indicate that migraine susceptibility genes are involved in various pathways, including neurotransmission, which have already been implicated in genetic studies of monogenic familial hemiplegic migraine, a subtype of migraine with aura.

Methods: To further explore the genetic background of migraine, we performed a gene set analysis of migraine GWAS data of 4954 clinic-based patients with migraine, as well as 13,390 controls. Curated sets of synaptic genes and sets of genes predominantly expressed in three glial cell types (astrocytes, microglia and oligodendrocytes) were investigated. Discussion: Our results show that gene sets containing astrocyte- and oligodendrocyte-related genes are associated with migraine, which is especially true for gene sets involved in protein modification and signal transduction. Observed differences between migraine with aura and migraine without aura indicate that both migraine types, at least in part, seem to have a different genetic background.

Keywords

Migraine, GWAS, pathway analysis, astrocytes, oligodendrocytes, synapse

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Introduction

Migraine is a common brain disorder affecting 15% of the population and characterized by recurrent attacks of severe, often unilateral headache associated with additional neurological symptoms such as nausea, vomiting and an increased sensitivity to light and sound (1). The presence of an aura accompanying the headache phase, consisting of transient neurological symptoms that are usually visual in nature, distinguishes between migraine with aura and migraine without aura.

Genome-wide association studies (GWAS), which have become a widely used approach to study the genetic background of common disorders, have already yielded 13 genome-wide significant loci for migraine (2–5). Several of the migraine candidate genes (i.e. MTDH, LRP1, PRDM16, MEF2D, ASTN2, PHACTR1, FHL5, MMP16) that have been brought up by these GWAS loci can be linked to (glutamatergic) neurotransmission (2–5). This seems in line with previous genetic findings in familial hemiplegic migraine, a monogenic subtype of migraine with aura (1), that revealed that all three causal genes identified so far (CACNA1A, ATP1A2 and SCN1A) play an important role in glutamatergic neurotransmission (6).

All currently known migraine GWAS loci combined explain only a minor fraction of the genetic heritability that has been estimated at 50% in twin studies (7), which is not different from GWAS findings in other common disorders (8). Part of the missing heritability may be explained by variants that do not reach the threshold for genome-wide significance, which is not unexpected given the small effect sizes that GWAS hits have and the large samples sizes that are needed to detect them. Such yet unidentified variants might be enriched in genes that play a role in disease-related pathways. Therefore, additional pathophysiological insights may be obtained by investigating GWAS data for specific disease-relevant gene sets. Here, we used three expert-curated glial cell-related gene lists (i.e. astrocyte, oligodendrocyte and microglia cell type-related gene lists) (9), and a synaptic gene list (10) to study the role of genes involved in synaptic mechanisms and glial cells in common migraine. The four gene lists were subdivided into functional gene sets and investigated for association with migraine. This approach has been applied successfully in, for example, schizophrenia, thus identifying a role for synaptic genes (11) as well as specific astrocyte and oligodendrocyte functions (9). For the analysis, we used GWAS data of 13,588 control individuals and 5037 migraine patients (2733 patients with migraine with aura and 2304 patients with migraine without aura) who were recruited via specialized headache clinics

(2,5). We used clinic-based migraine GWAS cohorts, as phenotypic information is more complete and accurate than in population-based cohorts. This might reduce the clinical and genetic heterogeneity within the cohorts, thereby increasing the likelihood of identifying associations with migraine. Our gene set analyses results indicated a role for astrocyte- and oligodendrocyte-specific genes in migraine pathophysiology with a different contribution in both migraine types.

Material and methods

Participants

In this study we investigated genetic data of 5037 migraine cases (2733 patients with migraine with aura and 2304 patients with migraine without aura) and 13,588 controls that were part of two previously published clinic-based GWA studies (2,5). Patients were collected via specialized headache centers within Europe (the Netherlands, Finland and Germany). Migraine diagnoses were established using a combination of questionnaires and individual interviews, based on the International Classification of Headache Disorders, third edition beta (ICHD-III beta) guidelines (1).

Genotyping and quality control (QC)

Patient and control samples were genotyped using Illumina arrays, except for the German control cohort that was used for the migraine without aura GWAS, which was genotyped partly using Illumina and partly using Affymetrix arrays (for details, see Anttila et al. (2) and Freilinger et al. (5)). Sample and genotype quality controls were performed specifically for this study using the raw genotyping data of the discovery cohorts of Anttila et al. (2) and Freilinger et al. (5), using the protocol described in Anderson et al. (12). In short, samples were removed in case of non-European ancestry, as measured with principal components analysis with EIGENSTRAT, sex discrepancies between genotypic and phenotypic data, missingness (> 3% of single nucleotide polymorphisms (SNPs)), outlying heterozygosity rate (> 5SD) and relatedness (identity by state (IBS) > 0.125). Samples were also removed if no valid principal components for stratification correction could be computed. We retained 4954 migraine cases and 13,390 controls after QC (Table 1). SNPs were removed from the analysis in case of: (i) a minor allele frequency < 1%; (ii) a deviation from Hardy-Weinberg equilibrium ($p < 10^{-5}$) in cases or controls; (iii) missing data for a SNP in > 3% of the individuals; (iv) a significantly different missing data rate between cases and controls ($p < 10^{-5}$); or (v) a non-unique genomic localization of SNPs.

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Table 1. Overview of the GWAS datasets that were used for gene-set analyses after QC.

Sample	# Cases (% females)	# Controls (% females)	SNPs						
Migraine with aura	Migraine with aura								
Dutch	629 (82)	4764 (59)	443,666						
German	992 (81)	1086 (61)	461,930						
Finnish	1031 (80)	3511 (53)	504,440						
Subtotal	2652	9361							
Migraine without au	ra								
Dutch	1118 (86)	2016 (54)	494,693						
German	1184 (87)	2013 (55)	425,411						
Subtotal	2302	4029							
Total migraine	4954	13,390							

GWAS: genome-wide association studies; QC: quality control; SNPs: single nucleotide polymorphisms.

Gene lists and gene sets

Four published, expert-curated gene lists, i.e. astrocytic, oligodendrocytic, microglia and synaptic, were used for the analysis (9,10). In brief, the glial gene lists (astrocytic, oligodendrocytic, microglial) contain genes with enriched expression in at least one of these cell types based on: (i) microarray data of fluorescentactivated cell sorting (FACS)-sorted oligodendrocytes, astrocytes and microglia from developing and adult mouse brain (13); (ii) genes that co-express with cell type markers in microarray data of human brain tissue (14,15); (iii) genes that are differentially expressed in microglia cell cultures after an inflammatory stimulus or in tissue of patients or animal models relevant to an inflammatory brain disorder (16–20); and (iv) a list of oligodendrocyte transcription factors (21). The synaptic gene list consists of: (i) a manually curated list of genes of presynaptic proteins with confirmation of proteomics data from isolated synapses (10,22–24), and (ii) a postsynaptic gene list based on proteomics data of synapses or the postsynaptic density (22–24). After combining the information from the abovementioned resources into three lists with cell typeenriched genes, the lists were further curated by removing genes that were present in more than one of these gene lists, or that were reported to be highly expressed in neurons. The final lists consisted of 1395 oligodendrocytic, 1688 astrocytic, 239 microglial and 914 synaptic genes.

The genes from each of the glial gene lists (astrocytic, oligodendrocytic, microglial) were categorized into functional gene sets based on Gene Ontology (GO) biological process annotations (9). Because of the hierarchical structure of GO, gene sets can be related and can contain overlapping genes.

The synaptic gene list was categorized into functional gene sets based on "expert knowledge" (10). This resulted in a total of 34 astrocytic, 29 oligodendrocytic, 19 microglial and 18 synaptic gene sets. Supplemental Table 1 shows the hierarchical structure of the functional gene sets.

Statistical analysis

Statistical analyses were performed using the in-house-developed software tool Joint Analysis of Genetic variants (JAG) (24) for: (i) the four gene lists, (ii) the 100 gene sets and (iii) the 4236 single genes in the four gene lists. JAG performs two types of statistical tests: the self-contained test and the competitive test. All p values in this study were calculated using JAG. For each GWAS dataset (Table 1), SNPs were assigned to genes if located between the transcription start and stop site of the respective gene, based on the National Center for Biotechnology Information (NCBI) human assembly build 37.3 and dbSNP release 135. Either 10 (for the Finnish and the German datasets) or 15 principal components (for the Dutch datasets) were used to correct for population stratification.

Self-contained p values (P_{SC}) were calculated for the four gene lists, the 100 gene sets and for 4236 single genes using at least 15,000 permutations of the case-control status (at $\alpha = 0.05$). The P_{SC} -values were calculated separately for each GWAS dataset, and then combined using Stouffer's Z-score method (25), weighted by the square root of the sample size. Bonferroni correction was used to account for multiple testing, correcting each gene set P_{SC} -value for the number of gene sets in each of the four gene lists. Hence the P_{SC} -values of the astrocytic, oligodendrocytic, microglial and synaptic gene sets were corrected for 34, 29, 19 and 18 tests, respectively.

A competitive test was performed for each of the four gene lists and for each of the 100 gene sets that was significant (at $\alpha = 0.05$) on the self-contained test. The competitive test calculates $P_{\rm SC}$ -values for 150 random gene sets containing the same number of genes as the original gene list or gene set, and calculates the competitive p value ($P_{\rm COMP}$) as the proportion of random gene sets with a lower $P_{\rm SC}$ -value. For gene sets found to be significant in the competitive test (at $\alpha = 0.05$), the contribution of each gene was calculated as the drop in $P_{\rm SC}$ -value when all SNPs from that gene were removed from the analysis.

The self-contained test in JAG tests the null hypothesis that a set of genes is not associated with migraine, while the competitive test provides insight into whether a set of genes is more strongly associated with migraine than other comparably sized sets of genes. For polygenic traits, self-contained tests are likely to show a

Gene list	Genes	SNPs	Migraine		Migraine with aura		Migraine without aura	
			P _{SC}	P _{COMP}	P _{SC}	P _{COMP}	P _{SC}	P _{COMP}
Oligodendrocytes	1395	25,559	< 0.001	< 0.001	< 0.001	0.040	< 0.001	0.033
Astrocytes	1688	28,035	< 0.001	0.25	< 0.001	0.17	< 0.001	0.42
Synapse	914	27,926	< 0.001	0.51	< 0.001	0.21	0.076	0.89

Table 2. Association of the four main gene lists with migraine.

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Association of the four gene lists with all migraine samples together, as well as the migraine with and migraine without aura separately. For each comparison a Bonferroni-corrected self-contained p value (indicated by P_{SC} -value) and a competitive p value (indicated by P_{COMP} -value) is shown.

0.45

significant *p* value simply because a gene set contains many genes. Competitive tests control for the polygenic nature of a trait by evaluating the association of a gene set relative to other gene sets. We thus interpreted only the results of the competitive test.

2730

0.018

Results

Microglia

Association analysis of the four gene lists with migraine

We performed a gene set analysis using migraine GWAS data of 4954 migraine patients (2652 patients with migraine with aura and 2302 patients with migraine without aura) and 13,390 controls to further explore the involvement of glial and synaptic genes in migraine pathophysiology. First, we conducted an overall analysis for the four gene lists, i.e. the three glial gene lists (astrocyte, oligodendrocyte and microglia) and the synaptic gene list (Table 2). A significant association ($P_{\rm SC}$ and $P_{\rm COMP}$ < 0.05) was found for oligodendrocyte-related genes with migraine, as well as with migraine with aura and migraine without aura, separately ($P_{\rm COMP} \le 0.001$, 0.040 and 0.033, respectively). No significant associations were found for the astrocytic, microglial and synaptic gene lists.

Association analysis of functional gene sets with migraine

Next, we studied specific functional gene sets that were defined within the four gene lists (Supplemental Table 1). Significant associations (P_{SC} and $P_{COMP} < 0.05$) with migraine were found for three astrocytic functional gene sets ("Protein transport," "Cell proliferation" and "Miscellaneous") and three oligodendrocyte functional gene sets ("Protein modification," "Signal transduction" and "Miscellaneous") (Table 3; Supplemental Table 1). Associations with functional gene sets were also calculated for migraine with and migraine without aura separately. Two functional gene sets showed a significant

association with migraine with aura (the astrocytic gene set "Cell proliferation" and the oligodendrocytic gene set "Miscellaneous") and two gene sets showed association with migraine without aura (the astrocytic gene set "Protein transport" and the oligodendrocytic gene set "Signal transduction") (Table 3). Interestingly, no overlap was found between functional gene sets that were associated with migraine with aura and those that were associated with migraine without aura.

0.084

0.49

Gene-based analysis

0.056

0.45

To understand whether the identified association of the functional gene sets with migraine was due to single genes or the gene set as a whole, a gene-based analysis was carried out. The association signal of the astrocytic gene sets "Protein transport" and "Cell proliferation" was mainly explained by the effect of *GLI3* (Supplemental Table 2). The associations of the other gene sets were based on a joint effect of the genes in the set (Supplemental Table 2).

Discussion

We performed a gene set analysis using glial and synaptic gene sets (9,10) combined with data from the two clinic-based migraine GWAS (2,5). The gene set analysis for the total oligodendrocyte gene list showed significant association with migraine. This suggests that in addition to neuronal dysfunction, oligodendrocytic dysfunction also seems to play a possibly modulating role in migraine pathophysiology. A role for oligodendrocytes in migraine has not yet been proposed. Studies have shown that oligodendrocytes communicate actively with neurons, probably to regulate their energy supply—and thereby their activity—or to stimulate their myelination (26,27). By modulating neuronal excitatory signaling, oligodendrocytes may play a role in migraine pathophysiology. Of note, a recent study identified myelin abnormalities in trigeminal nerves of migraine patients (28). Sensitization of the trigeminal Cephalalgia 36(7)

Table 3.	Association	of astrocytic and	doligodendrocytic	functional	gene sets with	migraine
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	Genes	SNPs	Migraine		Migraine with aura		Migraine without aura	
Functional gene set			P _{SC}	P _{COMP}	P _{SC}	P _{COMP}	P _{SC}	P _{COMP}
Astrocytes								
Protein transport	53	773	0.016	< 0.001	1.00	_	0.032	0.007
Cell proliferation	92	1337	< 0.001	0.007	< 0.001	< 0.001	1.00	_
Miscellaneous	499	8789	< 0.001	0.047	0.027	0.14	0.040	0.12
Transport and trafficking	266	4466	0.013	0.093	0.10	-	0.87	_
DNA metabolism	155	2287	0.039	0.10	1.00	_	0.062	_
Signal transduction	321	7507	0.062	_	0.032	0.093	1.00	_
Oligodendrocytes								
Miscellaneous	430	7689	< 0.001	0.007	< 0.001	< 0.001	0.39	_
Protein modification	157	3135	0.018	0.033	0.28	_	0.34	_
Signal transduction	250	6198	0.003	0.040	0.54	_	0.007	0.020
Protein metabolism	213	3661	0.031	0.087	0.53	-	0.28	-

Association of the functional gene sets with migraine, as well as with migraine with and migraine without aura separately. For each comparison a Bonferroni-corrected self-contained p value (indicated by P_{SC} -value) and a competitive p value (indicated by P_{COMP} -value) is shown. Only gene sets with P_{SC} < 0.05 are included in this table. SNPs: single nucleotide polymorphisms.

system and subsequent abnormal processing of pain signals is likely to play a causal role in generation of migraine headache (29).

Subsequently, we studied the 100 functional gene sets within the glial and synaptic gene lists and found that several astrocytic and oligodendrocytic gene sets were associated with migraine. Astrocyte dysfunction has already been implicated in migraine. For instance, the ATP1A2 protein, which is expressed in astrocytes in adults, is mutated in familial hemiplegic migraine type 2 (30), various sporadic hemiplegic migraine patients (31), and in two families with clustering of common migraine (32). Moreover, astrocytes have been shown to play a role in cortical spreading depression, the neurophysiological correlate of the migraine aura (33). For some of the astrocyte gene sets, the associations were mainly driven by the gene GLI3, for which the predominant expression in astrocytes (9) was confirmed by Zhang et al. in 2014 (34). GLI3 is involved in brain development, specifically the proliferation and cell fate specification of neural progenitors (35–37), and astrocyte function (37). Several migraine GWAS top hits pinpointed genes (i.e. MEF2D, ASTN2, PRDM16, PHACTR1 and TGFBR2) that are involved in brain development as well (38). Notably, GLI3 was recently shown to be differentially expressed in the cortex of two familial hemiplegic migraine type 1 mouse models (39), further supporting a relation between this gene and migraine pathophysiology.

A combined effect of all genes was observed for several astrocytic and oligodendrocytic gene sets. The oligodendrocytic gene sets "Signal transduction" and "Protein modification" might relate to the active role

that oligodendrocytes can play in the regulation of the energy supply and possibly the activity of neurons (26,27). Interestingly, the "Miscellaneous" gene sets contain a large percentage of membrane-associated and transmembrane genes. Furthermore, *PHACTR1*, one of the migraine GWAS top hits, contributes together with *PHACTR3* strongly to the association of the oligodendrocytic "Miscellaneous" gene set. Both genes regulate protein phosphatase 1, a protein with a main role in synaptic plasticity (40). This indicates that the genes from the "Miscellaneous" gene sets may have related functions involved in migraine that are not identified yet when classifying genes into GO-based functional categories.

There are limitations of the study design used. Public databases with gene expression information rarely reproduce the complete expression profile of a gene, thereby limiting the validity of gene sets that are built based on their data. By using well-defined, expert-curated gene sets (mostly based on experimental data), we tried to do our utmost to minimize this limitation. In addition, the validity of GO to classify glia cell and synaptic genes into gene sets based on their function is inherently limited to current, surely incomplete, knowledge about gene functions as well as the incomplete use of this knowledge within GO. As a consequence, for instance, the size of the "Miscellaneous" gene sets is quite large. Expected improvements and expansions of GO and similar public databases will increase the size and specificity of functional gene sets for future studies. A second limitation of this study is that by limiting our analyses to glia cell and synaptic gene sets, we potentially missed associations with other cell types, such as those involved in the

brain vasculature, that have been implicated in migraine (41). The reason for this omission is that we intended to increase the reliability of the results by using only well-defined, expert-curated gene sets, which are not available for other cell types.

Our separate analyses for migraine with and without aura showed different results. Of all functional gene sets that were associated with migraine, none were associated with both subtypes. This limited overlap in genetic risk for both migraine subtypes was previously observed in the migraine GWAS. For example, the only identified migraine with aura GWAS locus that contains the MTDH gene (2) showed no signal of association in the migraine without aura GWAS (3,5). This suggests that despite the clinical overlap, different and additional genes may play a role and may explain the aura phenomenon. Of note, the diagnosis of migraine with aura patients is based on ICHD-III beta guidelines and includes patients with migraine with aura who experience an aura in all of their attacks as well as patients who also experience attacks without an aura (1). Differences between migraine with aura and migraine without aura identified in the current study may be even more pronounced when including only migraine with aura patients who experience an aura

before each attack. At the moment, we lack genotyping data of a sufficiently large number of patients that exclusively have migraine with aura attacks to perform such an analysis.

For a similar reason we did not perform our analyses separately for males and females. Because of the low percentage of males in our cohorts (ranging from 13% in the German migraine without aura cohort and 20% in the Finnish migraine with aura cohort) (Table 1), the male subgroup would be underpowered for our gene set analyses. It is unlikely that such a gender analysis would have given different insights as: (i) heterogeneity analyses reported in the migraine GWAS meta-analysis (3) showed very low heterogeneity between female and male subgroups, and (ii) a SNP effect concordance analysis of migraine GWAS data (42) showed very high SNP concordance of GWAS signal between male and female subgroups.

In conclusion, our results indicate that astrocyteand oligodendrocyte-related gene sets may play a role in migraine susceptibility, especially those involved in protein modification and signal transduction. Furthermore, our results suggest that migraine with and migraine without aura may be conferred by—at least partly—different genetic factors.

Article highlights

- Gene-based analysis of genome-wide association studies (GWAS) data indicated that astrocytes and oligodendrocytes are genetically involved in migraine pathophysiology.
- The association of functional gene sets differed between migraine with aura and migraine without aura, indicating that both migraine types—at least in part—have a different genetic background.

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Declaration of conflicting interests

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References

- 1. Headache Classification Committee of the International Headache Society (IHS). The International Classification of Headache Disorders, 3rd edition (beta version). *Cephalalgia* 2013: 33: 629–808.
- 2. Anttila V, Stefansson H, Kallela M, et al. Genome-wide association study of migraine implicates a common susceptibility variant on 8q22.1. *Nat Genet* 2010; 42: 869–873.

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 Anttila V, Winsvold BS, Gormley P, et al. Genome-wide meta-analysis identifies new susceptibility loci for migraine. *Nat Genet* 2013; 45: 912–917.

- Chasman DI, Schurks M, Anttila V, et al. Genome-wide association study reveals three susceptibility loci for common migraine in the general population. *Nat Genet* 2011; 43: 695–698.
- Freilinger T, Anttila V, de Vries B, et al. Genome-wide association analysis identifies susceptibility loci for migraine without aura. *Nat Genet* 2012; 44: 777–782.
- van den Maagdenberg AMJM, Haan J, Terwindt GM, et al. Migraine: Gene mutations and functional consequences. Curr Opin Neurol 2007; 20: 299–305.
- Mulder EJ, Van Baal C, Gaist D, et al. Genetic and environmental influences on migraine: A twin study across six countries. Twin Res 2003; 6: 422–431.
- 8. Manolio TA, Collins FS, Cox NJ, et al. Finding the missing heritability of complex diseases. *Nature* 2009; 461: 747–753.
- Goudriaan A, de Leeuw C, Ripke S, et al. Specific glial functions contribute to schizophrenia susceptibility. Schizophr Bull 2013; 40: 925–935.
- Ruano D, Abecasis GR, Glaser B, et al. Functional gene group analysis reveals a role of synaptic heterotrimeric G proteins in cognitive ability. *Am J Hum Genet* 2010; 86: 113–125.
- Lips ES, Cornelisse LN, Toonen RF, et al. Functional gene group analysis identifies synaptic gene groups as risk factor for schizophrenia. *Mol Psychiatry* 2012; 17: 996–1006.
- Anderson CA, Pettersson FH, Clarke GM, et al. Data quality control in genetic case-control association studies. *Nat Protoc* 2010; 5: 1564–1573.
- 13. Cahoy JD, Emery B, Kaushal A, et al. A transcriptome database for astrocytes, neurons, and oligodendrocytes: A new resource for understanding brain development and function. *J Neurosci* 2008; 28: 264–278.
- Kang HJ, Kawasawa YI, Cheng F, et al. Spatio-temporal transcriptome of the human brain. *Nature* 2011; 478: 483–489.
- Oldham MC, Konopka G, Iwamoto K, et al. Functional organization of the transcriptome in human brain. *Nat Neurosci* 2008; 11: 1271–1282.
- Gebicke-Haerter PJ. Microarrays and expression profiling in microglia research and in inflammatory brain disorders. J Neurosci Res 2005; 81: 327–341.
- Paglinawan R, Malipiero U, Schlapbach R, et al. TGFbeta directs gene expression of activated microglia to an anti-inflammatory phenotype strongly focusing on chemokine genes and cell migratory genes. *Glia* 2003; 44: 219–231.
- Baker CA and Manuelidis L. Unique inflammatory RNA profiles of microglia in Creutzfeldt-Jakob disease. *Proc Natl Acad Sci U S A* 2003; 100: 675–679.
- 19. Glanzer JG, Enose Y, Wang T, et al. Genomic and proteomic microglial profiling: Pathways for neuroprotective inflammatory responses following nerve fragment clearance and activation. *J Neurochem* 2007; 102: 627–645.

 Re F, Belyanskaya SL, Riese RJ, et al. Granulocytemacrophage colony-stimulating factor induces an expression program in neonatal microglia that primes them for antigen presentation. *J Immunol* 2002; 169: 2264–2273.

- Fulton DL, Denarier E, Friedman HC, et al. Towards resolving the transcription factor network controlling myelin gene expression. *Nucleic Acids Res* 2011; 39: 7974–7991.
- 22. Li K, Hornshaw MP, van Minnen J, et al. Organelle proteomics of rat synaptic proteins: Correlation-profiling by isotope-coded affinity tagging in conjunction with liquid chromatography-tandem mass spectrometry to reveal post-synaptic density specific proteins. *J Proteome Res* 2005; 4: 725–733.
- 23. Li KW, Miller S, Klychnikov O, et al. Quantitative proteomics and protein network analysis of hippocampal synapses of CaMKIIalpha mutant mice. *J Proteome Res* 2007; 6: 3127–3133.
- Emes RD, Pocklington AJ, Anderson CN, et al. Evolutionary expansion and anatomical specialization of synapse proteome complexity. *Nat Neurosci* 2008; 11: 799–806
- 25. Whitlock MC. Combining probability from independent tests: The weighted Z-method is superior to Fisher's approach. *J Evol Biol* 2005; 18: 1368–1373.
- Stys PK. The axo-myelinic synapse. Trends Neurosci 2011; 34: 393–400.
- 27. Butt AM, Fern RF and Matute C. Neurotransmitter signaling in white matter. *Glia* 2014; 62: 1762–1779.
- Guyuron B, Yohannes E, Miller R, et al. Electron microscopic and proteomic comparison of terminal branches of the trigeminal nerve in patients with and without migraine headaches. *Plast Reconstr Surg* 2014; 134: 796e–805e.
- Bernstein C and Burstein R. Sensitization of the trigeminovascular pathway: Perspective and implications to migraine pathophysiology. *J Clin Neurol* 2012; 8: 89–99.
- 30. De Fusco M, Marconi R, Silvestri L, et al. Haploinsufficiency of *ATP1A2* encoding the Na+/K+ pump alpha2 subunit associated with familial hemiplegic migraine type 2. *Nat Genet* 2003; 33: 192–196.
- 31. Riant F, Ducros A, Ploton C, et al. De novo mutations in *ATP1A2* and *CACNA1A* are frequent in early-onset sporadic hemiplegic migraine. *Neurology* 2010; 75: 967–972.
- 32. Todt U, Dichgans M, Jurkat-Rott K, et al. Rare missense variants in *ATP1A2* in families with clustering of common forms of migraine. *Hum Mutat* 2005; 26: 315–321.
- 33. Charles A and Brennan K. Cortical spreading depression—new insights and persistent questions. *Cephalalgia* 2009; 29: 1115–1124.
- 34. Zhang Y, Chen K, Sloan SA, et al. An RNA-sequencing transcriptome and splicing database of glia, neurons, and vascular cells of the cerebral cortex. *J Neurosci* 2014; 34: 11929–11947.
- Wang H, Ge G, Uchida Y, et al. Gli3 is required for maintenance and fate specification of cortical progenitors. J Neurosci 2011; 31: 6440–6448.

36. Friedrichs M, Larralde O, Skutella T, et al. Lamination of the cerebral cortex is disturbed in Gli3 mutant mice. *Dev Biol* 2008; 318: 203–214.

- 37. Petrova R, Garcia AD and Joyner AL. Titration of GLI3 repressor activity by sonic hedgehog signaling is critical for maintaining multiple adult neural stem cell and astrocyte functions. *J Neurosci* 2013; 33: 17490–17505.
- 38. Eising E, de Vries B, Ferrari MD, et al. Pearls and pitfalls in genetic studies of migraine. *Cephalalgia* 2013; 33: 614–625.
- 39. de Vries B, Eising E, Broos LA, et al. RNA expression profiling in brains of familial hemiplegic migraine type 1 knock-in mice. *Cephalalgia* 2014; 34: 174–182.
- 40. Allen PB, Greenfield AT, Svenningsson P, et al. Phactrs 1–4: A family of protein phosphatase 1 and actin regulatory proteins. *Proc Natl Acad Sci U S A* 2004; 101: 7187–7192.
- 41. Bigal ME, Kurth T, Hu H, et al. Migraine and cardiovascular disease: Possible mechanisms of interaction. *Neurology* 2009; 72: 1864–1871.
- 42. Nyholt DR. Concordance of genetic risk across migraine subgroups: Impact on current and future genetic association studies. *Cephalalgia* 2015; 35: 489–499.