Review

Neutrophil Extracellular Traps in Atherosclerosis and Atherothrombosis

Yvonne Döring, Oliver Soehnlein, Christian Weber

<u>Abstract</u>: Neutrophil extracellular traps expelled from suicidal neutrophils comprise a complex structure of nuclear chromatin and proteins of nuclear, granular, and cytosolic origin. These net-like structures have also been detected in atherosclerotic lesions and arterial thrombi in humans and mice. Functionally, neutrophil extracellular traps have been shown to induce activation of endothelial cells, antigen-presenting cells, and platelets, resulting in a proinflammatory immune response. Overall, this suggests that they are not only present in plaques and thrombi but also they may play a causative role in triggering atherosclerotic plaque formation and arterial thrombosis. This review will focus on current findings of the involvement of neutrophil extracellular traps in atherogenesis and atherothrombosis. (*Circ Res.* 2017;120:736-743. DOI: 10.1161/CIRCRESAHA.116.309692.)

Key Words: atherosclerosis ■ coronary artery disease ■ extracellular trap ■ neutrophils ■ thrombosis

Phagocytes and especially neutrophils belong to the first line of defense against invading pathogens and use a broad spectrum of weaponry to eliminate microorganisms and further activate adaptive immunity.^{1,2} Besides the discharge of granule proteins (eg, antimicrobial peptides), proteases, and reactive oxygen metabolites, neutrophils have also been recognized to respond to infectious challenges via the formation of neutrophil extracellular traps (NETs). NETs are defined as large web-like structures composed of decondensed chromatin and neutrophil-derived nuclear, cytoplasmatic, and granular proteins, which are capable of ensnaring and killing pathogens. In this context, NET structures may function as a platform providing high local concentrations of effector molecules eliminating the intruder.3-5 The process of NET formation is termed NETosis and was introduced to discriminate this route of dying from other forms of cell death, eg, necrosis and apoptosis.³ Notably, other cell types such as macrophages⁶ or mast cells⁷ can also release decondensed chromatin, which is referred to as ETosis. Hence, NETs (and ETs) are believed to represent an important defense tool of innate immunity fighting pathogens8 but have also been implicated in the pathophysiology of cancer,9 autoimmunity,10 and chronic inflammation.¹¹ The latter also encompasses a variety of cardiovascular diseases and syndromes including coronary artery disease and stroke as the most frequent entities. The underlying pathophysiology termed atherosclerosis refers to a lipid-driven inflammatory disease of arteries, which develops at predilection sites with disturbed flow, where endothelial activation facilitates intimal retention of lipoproteins. Modified lipoproteins, eg, oxidized low-density lipoprotein, augment

endothelial damage and trigger recruitment of leukocytes, which eventually fail to clear lipoproteins, undergo cell death, and maintain inflammation. Ultimately, growing lesions will lead to vessel occlusion and subsequent ischemia or (arterial) thrombosis.^{12,13} Neutrophils and their specific contribution to the pathophysiology of atherosclerosis have long been denied; however, during the last years, we¹⁴⁻¹⁸ and others¹⁹⁻²⁶ have provided substantial evidence underlining presence and actions of neutrophils in early and established human and murine atherosclerotic lesions. In a nutshell, neutrophils aggravate endothelial dysfunction, attract leukocytes, in particular monocytes, to atherosclerotic lesions and promote foam cell formation. Advanced plaques are exposed to neutrophil-derived proteases and reactive oxygen species leading to plaque destabilization.^{17,27,28} NET formation may occur at all stages of disease progression and particularly high local concentrations of effector molecules are suspected to be proinflammatory²⁹ and therefore also atherogenic.

This review will focus on current findings of the involvement of NETs in atherosclerosis, arterial thrombosis (atherothrombosis), and myocardial infarction.

Mechanisms and Initiators of NET Formation

Among the 2 types of NETosis (lytic and vital) defined to date, lytic or suicidal NETosis is best known as a slow (hours) active form of cell death distinct from, for example, necrosis or apoptosis.⁸ Most of the mechanistic insight on lytic NETosis stems from studies investigating NETosis by treating isolated neutrophils with phorbol 12-myristate 13-acetate in vitro, which activates the neutrophil respiratory burst leading to the assembly

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Nonstandard Abbreviations and Acronyms	
IL	interleukin
NADPH	nicotinamide adenine dinucleotide phosphate (reduced form)
NETs	neutrophil extracellular traps
NE	neutrophil elastase
PAD4	peptidyl arginine deiminase 4
PR3	proteinase 3
r-tPA	recombinant tissue-type plasminogen activator

of multicomponent nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and subsequent reactive oxygen species formation.³⁰ Reactive oxygen species can liberate nuclear, granular, and cytoplasmic contents, including neutrophil elastase (NE) and myeloperoxidase, which degrade linker histones and enhance chromatin decondensation.³¹ Furthermore, peptidyl arginine deiminase 4 (PAD4) mediates histone deimination by citrullinating histones, which then mix with granule proteins and are expelled from the neutrophil.³² However, phorbol 12-myristate 13-acetate is an artificial stimulus, and although in vivo NET formation in mice seems to indeed depend on NE and PAD4 activity, it may not rely on reactive oxygen species generated by NADPH oxidase.³³

More recently the term vital NETosis has been introduced, describing a fast (minutes) process in vivo (humans and mice) where cell viability and functions are retained. Discharge of nuclear material is enabled by bleb formation and vesicular exportation. Vital NETosis can be specifically induced by microbial-specific molecular patterns (eg, lipopolysaccharides), which bind to host pattern recognition receptors and does not necessarily involve NADPH oxidase. Remaining anuclear neutrophils show integrity of the plasma membrane and still contain granules.³⁴⁻³⁷ Hence, the question remains, whether vital NETosis might even be the more physiologically relevant of the 2 processes because conclusions on lytic NETosis are mainly based on phorbol 12-myristate 13-acetate-stimulated in vitro assays. On the contrary, recent study suggests that NETosis in general may have been confused or misleadingly recognized as leukotoxic hypercitrullination and defective mitophagy leaving us with the notion that real NETosis is dependent on NADPH oxidase but not associated with hypercitrullination.38

Another phenomenon solely observed in vitro seems to be the formation of (N)ETs composed of mitochondrial DNA,^{39,40} whereas in vivo mitochondrial DNA released by neutrophils, for example, after major surgery, rather triggers NET formation.37,41,42 Besides the well-documented NET-inducers (phorbol 12-myristate 13-acetate, lipopolysaccharide, and mitochondrial DNA), many other stimuli have been reported to initiate NET formation in vitro and in vivo (human and mouse), including pathogens such as Gram-positive and Gram-negative bacteria,^{3,8,34,43} fungi,⁴⁴ parasites,⁴⁵ and viruses.⁴⁶ Moreover, NET formation may also be propagated by intrinsic mediators, such as hydrogen peroxide,8 cytokines,47,48 chemokines,3,49 cholesterol,⁵⁰ monosodium urate crystals,⁵¹ autoantibodies,⁵² and antibody-antigen complexes.53 Initiators of NET formation that have already been described in the context of atherosclerosis/atherothrombosis comprise antibody-antigen complexes,54 cholesterol crystals,50 or activated platelets.55

NET Content

Decondensed nuclear chromatin consisting of nucleic acids (DNA and RNA) and proteins (predominantly positively charged histones) constitute the backbone of a NET. Histones (accounting for $\approx 70\%$ of the NET protein content) facilitate the adhesion of (negatively charged) microbial and viral pathogens and both; histones⁵⁶ and nucleic acids⁵⁷ can per se exert bactericidal activity. Other NET-associated antimicrobial peptides comprise granule, cytoplasmic and cytoskeletal proteins, and metabolic enzymes of neutrophil origin.31,58,59 In addition, it is conceivable that NETs contain effector molecules or constituents released by neighboring cells, as evidenced by a study showing trapping and degradation of proinflammatory mediators by aggregated NET structures in vitro and in vivo (mouse).⁵¹ Thus, not only the stimuli inducing NET formation are manifold but also the NET proteome may differ depending on the localization (tissue versus circulation) and disease (acute versus chronic).^{29,43,60,61} Nevertheless ≈20 (mostly neutrophil-derived) proteins have been proposed as a core NET proteome including histones, myeloperoxidase, NE, proteinase 3, cathepsin G, and α -defensins.^{61–63} Yet, disease-specific NETome patterns await to be properly defined.

NETs in Innate Immune Defense

Initially, NETs were discovered and described as an important tool to fight invading pathogens, in particular bacteria.³ On the contrary, NET-associated microbes have also been found to remain alive and pathogenic.64,65 Nevertheless, restoration of NET formation in a patient with chronic granulomatous disease (strong decrease in NADPH oxidase function) and Aspergillus infection significantly improved the immune response against the fungi.66 Accordingly, PAD4-deficient mice, which are incapable of generating NETs,32 are more susceptible to bacterial infection, as shown in a mouse model of infectious necrotizing fasciitis.⁶⁷ However, another study using PAD4 knockout mice did not reveal differences in morbidity or mortality compared with wild-type mice in a model of peritonitis (cecal ligation puncture).68 Similarly, patients having Papillon-Lefèvre Syndrome are unable to generate NETs-except in saliva-because of the lack of all neutrophil serine proteases but do not exhibit severe immunodeficiencies. Instead, they have pronounced juvenile periodontitis caused by fungi and bacteria although saliva (containing NETs) is supposed to prevent periodontal disease.⁶⁹⁻⁷¹ In conclusion, the overall importance of NET-mediated killing of microbes may differ depending on the type of pathogen and the specific location versus a more systemic affliction; it is hence conceivable that NETs deficiency may be compensated by other antimicrobial mechanisms during inflammatory immune responses.

NETs in Atherosclerosis

Atherosclerosis, widely recognized as a lipid-driven inflammatory disease of the arterial vessel wall, results in intimal lesion growth. Progressing lesions may eventually rupture, thereby inducing intraluminal thrombosis leading to acute cardiac events and ischemic stroke.⁷² Of note, extracellular DNA (eg, NET derived) exerts cytotoxic and prothrombotic effects, possibly providing a causative link between inflammation and coagulation. In line with this, NETs were identified in luminal location in murine and human atherosclerotic lesions^{50,73,74} and implicated in arterial thrombosis (see subsequent section).

In apolipoprotein E-deficient knock-out (Apoe-/-) mice, we could elucidate a mechanism of NET-driven atherogenesis, involving the autoimmune activation of plasmacytoid dendritic cells. Mechanistically, complexes of self-DNA (presumably NET-borne DNA but also self-DNA from dying cells) and neutrophil-derived granule proteins (eg, cathelicidin) stimulate plasmacytoid dendritic cells in the vessel wall, resulting in a strong type I interferon response, which drives atherogenesis.54 Depletion of plasmacytoid dendritic cells instead resulted in reduced plaque burden and type I interferon response.^{54,75} Moreover, NETs have been suggested to directly induce endothelial dysfunction (as a starting point for atherosclerosis) by activation and damage of endothelial cells.76-78 Findings by Knight et al⁷⁹ further underscore the importance of NETs in atherosclerotic lesion development by showing that inhibition of PAD4 (described to be important in NET formation³²) by chloramidine treatment prevents NET formation, thereby decreasing atherosclerotic lesion size and delaying carotid artery thrombosis in a mouse model of atherosclerosis. Comparable effects could not be reproduced in mice treated with a neutrophil-depleting antibody or in mice lacking a functional type I interferon receptor. In summary, these data suggest an important role of NETs in the instigation of a type I interferon response driving atherogenesis.⁷⁹ On the contrary, the importance of a NET-driven type I interferon response in atherogenesis is called into doubt by Warnatsch et al,50 instead showing that cholesterol crystals, as a sterile danger signal, induce lesional NET formation in Apoe-/- mice after a high-fat diet for only 8 weeks. These NETs are thought to subsequently prime macrophages for production of interleukin (IL)- 1β , resulting in activation of a Th17 response, which further amplifies immune cell recruitment into atherosclerotic lesions. To render Apoe-/- mice devoid of NETs, the authors used different approaches; however, in their hands, only inhibition of NADPH oxidase with diphenylene iodonium or blocking of the neutrophil-specific proteases NE and proteinase 3 (PR3)³¹ abrogated NET formation, whereas chloramidine treatment was ineffective. As a consequence, they used Apoe-/- mice lacking NE and PR3 (Apoe-/-Elane-/-Prtn3-/-) to study atherosclerosis in the absence of NETs, revealing reduced lesions size in Apoe-/-Elane-/-Prtn3-/- animals after 8 but not after 4 weeks of high-fat diet feeding. Likewise, they found diminished plaque growth in Apoe-/- mice treated with deoxyribonuclease to degrade NETs. Excluding the possibility that NE/PR3 deficiency causes intrinsic defects in neutrophil chemotaxis or extravasation, which would add to reduced lesions sizes in Apoe-'-Elane-'-Prtn3-'-animals, the authors conclude that NET-mediated priming of macrophages induces a strong IL-1β/Th17 response driving atherogenesis.⁵⁰ In our hands,⁸⁰ however, Apoe-/-Elane-/-Prtn3-/- animals showed reduced atherosclerotic lesion size only at early but not at advanced stages of lesion development and lack of NE alone did not affect plaque growth. In addition, we could not observe any effect on lesion size after NET degradation through repetitive deoxyribonuclease injection. Hence, our findings do not support the

conclusion that the absence of elastase-provoked NET release or deoxyribonuclease treatment alters atherosclerotic lesion development. In summary, these results point toward PR3 rather than toward NE triggering early atherosclerosis, giving rise to the notion that PR3-mediated cytokine maturation (in particular cleavage of pro–IL-1 β^{81}) seems to be more important for lesion formation than NET-mediated macrophage priming.⁸⁰

At a diagnostic or prognostic level, NETs (or their components) have been suggested to serve as biomarkers predicting the severity of atherosclerosis and the risk of future cardiovascular events.82 In this study, 282 human individuals with suspected coronary artery disease were examined about the prevalence of double-stranded DNA, nucleosomes, citrullinated histone H4, and myeloperoxidase-DNA complexes in coronary atherosclerosis. Furthermore, plasma markers of coagulation activation and inflammation were determined. The results demonstrated that plasma levels of double-stranded DNA, nucleosomes, and myeloperoxidase-DNA complexes were positively associated with thrombin generation and significantly elevated in patients with severe coronary atherosclerosis or extremely calcified coronary arteries. In addition, high plasma nucleosome levels were found to be an independent risk factor of severe coronary stenosis, and the load of myeloperoxidase-DNA complexes correlated with the number of atherosclerotic coronary vessels and the occurrence of major adverse cardiac events.82 A synopsis of potential NETmediated mechanisms driving atherosclerosis has been visualized in a Figure. However, NET localization in atherosclerosis (mouse and human) was mostly luminal or-in a lesional context-somewhat associative by identifying single NET relevant structures/proteins rather than a real NET-releasing neutrophil. Clearly, further in-depth studies will be required to detail the role of lesional NETs and their interactions with potential immune cells (plasmacytoid dendritic cells and macrophages) triggering proinflammatory immune responses such as type I interferon and Th17 responses, which can fuel atherosclerosis.

NETs in Atherothrombosis and Myocardial Infarction

Studies specifically investigating the role of NETs in the context of atherothrombosis are scarce, whereas more detailed knowledge is available about their involvement in venous thrombosis.^{83–85} However, circulating leukocytes, in particular monocytes, play a crucial role in atherothrombosis,^{86,87} and systemic neutrophil counts are robust predictors of acute coronary events⁸⁸ and impact outcomes⁸⁹ in humans. Furthermore, neutrophils are present in coronary thrombi^{90,91} and have been detected in surgical thrombectomies and abdominal aortic aneurysms.⁹² Following on these findings, recent research specifically addressed whether and how NETs (and NETosis) may affect arterial thrombus formation and its complications.

Riegger et al⁹¹ analyzed 253 samples from patients with stent thrombosis after percutaneous coronary intervention. Around 23% of the specimens investigated contained NETs, but no differences in the number of NETs could be detected within these thrombi with respect to the timing of stent



Figure. Emerging roles of neutrophil extracellular traps (NETs) in atherosclerosis and atherothrombosis. (A) Luminally netting neutrophils activate leukocytes, platelets, and endothelial cells creating a proinflammatory milieu presumably resulting in endothelial dysfunction, the initial trigger of lesion development. (B/C) Lesional NETs may initiate a interleukin-1 β /TH17 (T helper 17) and type I interferon response, which leads to further activation of lesional leukocytes, releasing more proinflammatory mediators. (D/E) Furthermore, it may be assumed that NET-driven proinflammatory responses will cause an inflammatory environment that favors plaque destabilization and rupture. During atherothrombosis, NETs may trigger activation of the coagulation cascade and increase thrombus stability thus orchestrating arterial occlusion.

thrombosis (early versus late), the stent type, or in comparison to samples from patients with spontaneous myocardial infarction. Nevertheless, analysis of thrombi from 111 patients with ST-segment elevation as a sign of acute coronary syndrome and subjected to the primary percutaneous coronary intervention revealed highly activated neutrophils at culprit lesional sites compared with femoral neutrophils. Moreover, the NET burden in these thrombi correlated positively with infarct size and negatively with ST-segment resolution, whereas deoxyribonuclease activity in these lesions showed a negative correlation with infarct size but positive with ST-segment resolution. Notably, ex vivo addition of deoxyribonuclease to these thrombi accelerated their lysis. In conclusion, the authors postulate that the balance of NETing neutrophils at rupture-prone lesion sites and endogenous deoxyribonuclease activity is predictors of ST-segment resolution and myocardial infarct size.93

Along these lines, Maugeri et al⁹⁴ performed histology on 26 thrombectomies from patients after acute myocardial infarction, revealing that activated platelets present high-mobility group box 1 protein to neutrophils thereby inducing NET formation. The authors speculate that these NETs may have contributed to plaque rupture with subsequent thrombus formation. In accordance with these findings, platelet-derived high-mobility group box 1 protein has also been shown to facilitate NET formation and coagulation in a mouse model of venous thrombosis.⁹⁵ Another study examined 45 coronary thrombectomy specimens (15 fresh, 15 lytic, and 15 organized thrombi) obtained from patients after acute myocardial infarction and detected NETs in fresh and lytic, but not in organized thrombi.⁹⁶ Notably, these NETs were found to be coated with IL-17A and IL-17F, which has been described to promote thrombosis by enhancing platelet aggregation.⁹⁶

As the main initiator of coagulation critically involved in arterial thrombosis,⁹⁷ tissue factor has also been investigated in patients with acute ST-segment–elevation myocardial infarction. Samples of thrombotic material and surrounding blood from the infarct-related coronary artery and the noninfarcted area of 18 patients were collected during the primary

percutaneous revascularization. Analysis of these specimens revealed local accumulation of tissue factor-bearing NETs at sites of coronary thrombosis and blood neutrophils releasing NETs and exposing tissue factor in the infarct-related area, but not in the noninfarcted area of these patients. In addition, neutrophil islets and NETs decorated with tissue factor were detected in thrombi obtained from the infarcted region.98 As a conclusion, the authors state that the interactions of activated platelets with neutrophils at sites of plaque rupture during acute myocardial infarction result in local NET formation and delivery of active tissue factor altogether fostering thrombus formation.98 Interestingly, another study investigating carotid culprit plaque samples from 157 patients showed the presence of periodontal bacteria especially in hemorrhagic atherosclerotic carotid plaques, correlating with an increased prevalence of activated neutrophils, as evidenced for example by measuring myeloperoxidase-DNA complexes, which would be indicative of NET release.99 Taken together, these data suggest a potential role of periodontal microorganisms, inducing NET formation and subsequent plaque rupture.99

In mice, neutrophil-derived externalized nucleosomes, which may turn into NETs, have been studied in an arterial vessel injury model induced by ferric chloride application. Infusion of the anti-H2A-H2B-DNA antibody (neutralizing histones as major components of these nucleosomes) into ferric chloride-treated wild-type mice leads to prolonged time to occlusion and lower thrombus stability in carotid arteries. Notably, no effect of antibody infusion is observed in the NE/cathepsin G-deficient mice after induction of vessel injury. Mechanistically, externalized nucleosomes can enable the coassembly of NE and its substrate tissue factor pathway inhibitor on the surface of activated neutrophils triggering thrombosis. Hence, in sterile inflammation, neutrophil-derived serine proteases and nucleosomes may contribute to large-vessel thrombosis, leading to myocardial infarction and stroke.55

The role of NETs has also been examined in an alternative model of myocardial ischemia-reperfusion and myocardial noreflow. Male Wistar rats were treated with deoxyribonuclease, recombinant tissue-type plasminogen activator (r-tPA), a combination of deoxyribonuclease and r-tPA or left untreated for 45 minutes after induction of myocardial ischemia. Comparing control rats with those treated with a combination of deoxyribonuclease and r-tPA revealed reduced NET density and noflow area in the ischemic region, as well as reduced infarct size in deoxyribonuclease/r-tPA-treated animals. In addition, deoxyribonuclease/r-tPA treatment significantly ameliorated ischemia-reperfusion injury-induced left ventricular remodeling, as compared to controls. The authors conclude that NETs have a detrimental effect in ischemia-reperfusion-challenged myocardium and suggest deoxyribonuclease treatment regimens in patients with myocardial ischemia reperfusion injury and coronary no-reflow.100 Similarly and again using a mouse model of myocardial ischemia reperfusion, Savchenko et al101 showed a significant cardioprotective effect of deoxyribonuclease I treatment on myocardial ischemia reperfusion injury. In the same study, PAD4-deficient mice, which are not capable of producing NETs, were significantly protected from myocardial ischemia reperfusion injury and deoxyribonuclease I treatment had no added beneficial effect in these animals. Deoxyribonuclease I treatment also protects mice from cerebral ischemia reperfusion injury,¹⁰² suggesting that chromatin/ NET degradation by deoxyribonuclease and consequently removal of extracellular histones significantly alleviates tissue damage after ischemia-reperfusion injury. Hence, removal of extracellular chromatin generated at the sites of infarction not only implicates NETs in the myocardial damage but may also offer promising therapeutic options.

Summary, Outlook, and Conclusion

As evident from the studies highlighted and discussed above, NETs clearly do matter and significantly affect the initiation and progression of atherosclerotic lesions. Moreover, NETs can induce and contribute to arterial thrombus formation, stability, and growth (Figure). Although deoxyribonuclease treatment, likely enhancing thrombus lysis, seems to harbor a relevant therapeutic potential in acute cardiac events,¹⁰³ its utility and applicability in preventing NET formation or in digesting already established NETs to reduce atherosclerotic lesion growth is at least debatable and will remain controversial. Undoubtedly, more in-depth studies are needed to meticulously dissect the exact mechanisms of in vivo NET formation³⁶ and to clarify the importance of histone citrullination for NETosis.³⁸ A better understanding of in vivo NETosis, both with regards to the structural constituents and to their context-specific functional decoration, will be a prerequisite to further elucidate the role of NETs in the development of atherosclerotic plaques and will be of paramount importance to identify, validate, and enable the best molecular candidates for therapeutic targeting.

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Disclosures

None.

References

- Soehnlein O, Lindbom L. Phagocyte partnership during the onset and resolution of inflammation. *Nat Rev Immunol.* 2010;10:427–439. doi: 10.1038/nri2779.
- Nauseef WM, Borregaard N. Neutrophils at work. Nat Immunol. 2014;15:602–611. doi: 10.1038/ni.2921.
- Brinkmann V, Reichard U, Goosmann C, Fauler B, Uhlemann Y, Weiss DS, Weinrauch Y, Zychlinsky A. Neutrophil extracellular traps kill bacteria. *Science*. 2004;303:1532–1535. doi: 10.1126/science.1092385.
- Brinkmann V, Zychlinsky A. Beneficial suicide: why neutrophils die to make NETs. *Nat Rev Microbiol.* 2007;5:577–582. doi: 10.1038/ nrmicro1710.
- Branzk N, Lubojemska A, Hardison SE, Wang Q, Gutierrez MG, Brown GD, Papayannopoulos V. Neutrophils sense microbe size and selectively release neutrophil extracellular traps in response to large pathogens. *Nat Immunol.* 2014;15:1017–1025. doi: 10.1038/ni.2987.
- Boe DM, Curtis BJ, Chen MM, Ippolito JA, Kovacs EJ. Extracellular traps and macrophages: new roles for the versatile phagocyte. *J Leukoc Biol*. 2015;97:1023–1035. doi: 10.1189/jlb.4RI1014-521R.

- Möllerherm H, von Köckritz-Blickwede M, Branitzki-Heinemann K. Antimicrobial activity of mast cells: role and relevance of extracellular DNA traps. *Front Immunol*. 2016;7:265. doi: 10.3389/fimmu.2016.00265.
- Fuchs TA, Abed U, Goosmann C, Hurwitz R, Schulze I, Wahn V, Weinrauch Y, Brinkmann V, Zychlinsky A. Novel cell death program leads to neutrophil extracellular traps. *J Cell Biol*. 2007;176:231–241. doi: 10.1083/jcb.200606027.
- Olsson AK, Cedervall J. NETosis in cancer platelet-neutrophil crosstalk promotes tumor-associated pathology. *Front Immunol.* 2016;7:373. doi: 10.3389/fimmu.2016.00373.
- Grayson PC, Kaplan MJ. At the bench: neutrophil extracellular traps (NETs) highlight novel aspects of innate immune system involvement in autoimmune diseases. *J Leukoc Biol.* 2016;99:253–264. doi: 10.1189/ jlb.5BT0615-247R.
- Pinegin B, Vorobjeva N, Pinegin V. Neutrophil extracellular traps and their role in the development of chronic inflammation and autoimmunity. *Autoimmun Rev.* 2015;14:633–640. doi: 10.1016/j.autrev.2015.03.002.
- Weber C, Noels H. Atherosclerosis: current pathogenesis and therapeutic options. Nat Med. 2011;17:1410–1422. doi: 10.1038/nm.2538.
- Lippi G, Franchini M, Targher G. Arterial thrombus formation in cardiovascular disease. *Nat Rev Cardiol.* 2011;8:502–512. doi: 10.1038/ nrcardio.2011.91.
- Döring Y, Drechsler M, Wantha S, Kemmerich K, Lievens D, Vijayan S, Gallo RL, Weber C, Soehnlein O. Lack of neutrophil-derived CRAMP reduces atherosclerosis in mice. *Circ Res.* 2012;110:1052–1056. doi: 10.1161/CIRCRESAHA.112.265868.
- Drechsler M, Megens RT, van Zandvoort M, Weber C, Soehnlein O. Hyperlipidemia-triggered neutrophilia promotes early atherosclerosis. *Circulation*. 2010;122:1837–1845. doi: 10.1161/ CIRCULATIONAHA.110.961714.
- Wantha S, Alard JE, Megens RT, van der Does AM, Döring Y, Drechsler M, Pham CT, Wang MW, Wang JM, Gallo RL, von Hundelshausen P, Lindbom L, Hackeng T, Weber C, Soehnlein O. Neutrophil-derived cathelicidin promotes adhesion of classical monocytes. *Circ Res.* 2013;112:792–801. doi: 10.1161/CIRCRESAHA.112.300666.
- Soehnlein O. Multiple roles for neutrophils in atherosclerosis. *Circ Res.* 2012;110:875–888. doi: 10.1161/CIRCRESAHA.111.257535.
- Zernecke A, Bot I, Djalali-Talab Y, Shagdarsuren E, Bidzhekov K, Meiler S, Krohn R, Schober A, Sperandio M, Soehnlein O, Bornemann J, Tacke F, Biessen EA, Weber C. Protective role of CXC receptor 4/CXC ligand 12 unveils the importance of neutrophils in atherosclerosis. *Circ Res.* 2008;102:209–217. doi: 10.1161/CIRCRESAHA.107.160697.
- Ionita MG, van den Borne P, Catanzariti LM, Moll FL, de Vries JP, Pasterkamp G, Vink A, de Kleijn DP. High neutrophil numbers in human carotid atherosclerotic plaques are associated with characteristics of rupture-prone lesions. *Arterioscler Thromb Vasc Biol.* 2010;30:1842–1848. doi: 10.1161/ATVBAHA.110.209296.
- Marino F, Tozzi M, Schembri L, Ferraro S, Tarallo A, Scanzano A, Legnaro M, Castelli P, Cosentino M. Production of IL-8, VEGF and elastase by circulating and intraplaque neutrophils in patients with carotid atherosclerosis. *PLoS One*. 2015;10:e0124565. doi: 10.1371/journal.pone.0124565.
- Naruko T, Ueda M, Haze K, van der Wal AC, van der Loos CM, Itoh A, Komatsu R, Ikura Y, Ogami M, Shimada Y, Ehara S, Yoshiyama M, Takeuchi K, Yoshikawa J, Becker AE. Neutrophil infiltration of culprit lesions in acute coronary syndromes. *Circulation*. 2002;106:2894–2900.
- Nencioni A, da Silva RF, Fraga-Silva RA, et al. Nicotinamide phosphoribosyltransferase inhibition reduces intraplaque CXCL1 production and associated neutrophil infiltration in atherosclerotic mice. *Thromb Haemost*. 2014;111:308–322. doi: 10.1160/TH13-07-0531.
- Pende A, Artom N, Bertolotto M, Montecucco F, Dallegri F. Role of neutrophils in atherogenesis: an update. *Eur J Clin Invest*. 2016;46:252–263. doi: 10.1111/eci.12566.
- Quinn KL, Henriques M, Tabuchi A, et al. Human neutrophil peptides mediate endothelial-monocyte interaction, foam cell formation, and platelet activation. *Arterioscler Thromb Vasc Biol.* 2011;31:2070–2079. doi: 10.1161/ATVBAHA.111.227116.
- Rotzius P, Thams S, Soehnlein O, Kenne E, Tseng CN, Björkström NK, Malmberg KJ, Lindbom L, Eriksson EE. Distinct infiltration of neutrophils in lesion shoulders in ApoE-/- mice. *Am J Pathol.* 2010;177:493– 500. doi: 10.2353/ajpath.2010.090480.
- van Leeuwen M, Gijbels MJ, Duijvestijn A, Smook M, van de Gaar MJ, Heeringa P, de Winther MP, Tervaert JW. Accumulation of myeloperoxidase-positive neutrophils in atherosclerotic lesions in LDLR-/- mice. Arterioscler Thromb Vasc Biol. 2008;28:84–89. doi: 10.1161/ ATVBAHA.107.154807.

- Döring Y, Drechsler M, Soehnlein O, Weber C. Neutrophils in atherosclerosis: from mice to man. *Arterioscler Thromb Vasc Biol.* 2015;35:288– 295. doi: 10.1161/ATVBAHA.114.303564.
- Moreno JA, Ortega-Gómez A, Delbosc S, Beaufort N, Sorbets E, Louedec L, Esposito-Farèse M, Tubach F, Nicoletti A, Steg PG, Michel JB, Feldman L, Meilhac O. In vitro and in vivo evidence for the role of elastase shedding of CD163 in human atherothrombosis. *Eur Heart J.* 2012;33:252–263. doi: 10.1093/eurheartj/ehr123.
- Sørensen OE, Borregaard N. Neutrophil extracellular traps the dark side of neutrophils. J Clin Invest. 2016;126:1612–1620. doi: 10.1172/ JCI84538.
- Karlsson A, Dahlgren C. Assembly and activation of the neutrophil NADPH oxidase in granule membranes. *Antioxid Redox Signal*. 2002;4:49–60. doi: 10.1089/152308602753625852.
- Papayannopoulos V, Metzler KD, Hakkim A, Zychlinsky A. Neutrophil elastase and myeloperoxidase regulate the formation of neutrophil extracellular traps. J Cell Biol. 2010;191:677–691. doi: 10.1083/jcb.201006052.
- 32. Wang Y, Li M, Stadler S, Correll S, Li P, Wang D, Hayama R, Leonelli L, Han H, Grigoryev SA, Allis CD, Coonrod SA. Histone hypercitrullination mediates chromatin decondensation and neutrophil extracellular trap formation. J Cell Biol. 2009;184:205–213. doi: 10.1083/jcb.200806072.
- 33. Kolaczkowska E, Jenne CN, Surewaard BG, Thanabalasuriar A, Lee WY, Sanz MJ, Mowen K, Opdenakker G, Kubes P. Molecular mechanisms of NET formation and degradation revealed by intravital imaging in the liver vasculature. *Nat Commun.* 2015;6:6673. doi: 10.1038/ncomms7673.
- 34. Pilsczek FH, Salina D, Poon KK, Fahey C, Yipp BG, Sibley CD, Robbins SM, Green FH, Surette MG, Sugai M, Bowden MG, Hussain M, Zhang K, Kubes P. A novel mechanism of rapid nuclear neutrophil extracellular trap formation in response to Staphylococcus aureus. *J Immunol.* 2010;185:7413–7425. doi: 10.4049/jimmunol.1000675.
- Byrd AS, O'Brien XM, Johnson CM, Lavigne LM, Reichner JS. An extracellular matrix-based mechanism of rapid neutrophil extracellular trap formation in response to *Candida albicans. J Immunol.* 2013;190:4136– 4148. doi: 10.4049/jimmunol.1202671.
- Yipp BG, Kubes P. NETosis: how vital is it? *Blood*. 2013;122:2784–2794. doi: 10.1182/blood-2013-04-457671.
- 37. Yipp BG, Petri B, Salina D, Jenne CN, Scott BN, Zbytnuik LD, Pittman K, Asaduzzaman M, Wu K, Meijndert HC, Malawista SE, de Boisfleury Chevance A, Zhang K, Conly J, Kubes P. Infection-induced NETosis is a dynamic process involving neutrophil multitasking in vivo. *Nat Med.* 2012;18:1386–1393. doi: 10.1038/nm.2847.
- Konig MF, Andrade F. A critical reappraisal of neutrophil extracellular traps and netosis mimics based on differential requirements for protein citrullination. *Front Immunol.* 2016;7:461. doi: 10.3389/fimmu.2016.00461.
- Yousefi S, Mihalache C, Kozlowski E, Schmid I, Simon HU. Viable neutrophils release mitochondrial DNA to form neutrophil extracellular traps. *Cell Death Differ*. 2009;16:1438–1444. doi: 10.1038/cdd.2009.96.
- Yousefi S, Gold JA, Andina N, Lee JJ, Kelly AM, Kozlowski E, Schmid I, Straumann A, Reichenbach J, Gleich GJ, Simon HU. Catapult-like release of mitochondrial DNA by eosinophils contributes to antibacterial defense. *Nat Med.* 2008;14:949–953. doi: 10.1038/nm.1855.
- 41. Itagaki K, Kaczmarek E, Lee YT, Tang IT, Isal B, Adibnia Y, Sandler N, Grimm MJ, Segal BH, Otterbein LE, Hauser CJ. Mitochondrial DNA released by trauma induces neutrophil extracellular traps. *PLoS One*. 2015;10:e0120549. doi: 10.1371/journal.pone.0120549.
- McIlroy DJ, Jarnicki AG, Au GG, Lott N, Smith DW, Hansbro PM, Balogh ZJ. Mitochondrial DNA neutrophil extracellular traps are formed after trauma and subsequent surgery. *J Crit Care*. 2014;29:1133.e1–1133. e5. doi: 10.1016/j.jcrc.2014.07.013.
- Yang H, Biermann MH, Brauner JM, Liu Y, Zhao Y, Herrmann M. New insights into neutrophil extracellular traps: mechanisms of formation and role in inflammation. *Front Immunol.* 2016;7:302. doi: 10.3389/fimmu. 2016.00302.
- 44. Urban CF, Ermert D, Schmid M, Abu-Abed U, Goosmann C, Nacken W, Brinkmann V, Jungblut PR, Zychlinsky A. Neutrophil extracellular traps contain calprotectin, a cytosolic protein complex involved in host defense against Candida albicans. *PLoS Pathog*. 2009;5:e1000639. doi: 10.1371/ journal.ppat.1000639.
- 45. Rochael NC, Guimarães-Costa AB, Nascimento MT, DeSouza-Vieira TS, Oliveira MP, Garcia e Souza LF, Oliveira MF, Saraiva EM. Classical ROSdependent and early/rapid ROS-independent release of neutrophil extracellular traps triggered by Leishmania parasites. *Sci Rep.* 2015;5:18302. doi: 10.1038/srep18302.
- Saitoh T, Komano J, Saitoh Y, Misawa T, Takahama M, Kozaki T, Uehata T, Iwasaki H, Omori H, Yamaoka S, Yamamoto N, Akira S. Neutrophil

extracellular traps mediate a host defense response to human immunodeficiency virus-1. *Cell Host Microbe*. 2012;12:109–116. doi: 10.1016/j. chom.2012.05.015.

- Martinelli S, Urosevic M, Daryadel A, Oberholzer PA, Baumann C, Fey MF, Dummer R, Simon HU, Yousefi S. Induction of genes mediating interferon-dependent extracellular trap formation during neutrophil differentiation. *J Biol Chem.* 2004;279:44123–44132. doi: 10.1074/jbc. M405883200.
- 48. Khandpur R, Carmona-Rivera C, Vivekanandan-Giri A, Gizinski A, Yalavarthi S, Knight JS, Friday S, Li S, Patel RM, Subramanian V, Thompson P, Chen P, Fox DA, Pennathur S, Kaplan MJ. NETs are a source of citrullinated autoantigens and stimulate inflammatory responses in rheumatoid arthritis. *Sci Transl Med.* 2013;5:178ra40. doi: 10.1126/ scitranslmed.3005580.
- Rossaint J, Herter JM, Van Aken H, Napirei M, Döring Y, Weber C, Soehnlein O, Zarbock A. Synchronized integrin engagement and chemokine activation is crucial in neutrophil extracellular trap-mediated sterile inflammation. *Blood.* 2014;123:2573–2584. doi: 10.1182/ blood-2013-07-516484.
- Warnatsch A, Ioannou M, Wang Q, Papayannopoulos V. Inflammation. Neutrophil extracellular traps license macrophages for cytokine production in atherosclerosis. *Science*. 2015;349:316–320. doi: 10.1126/science. aaa8064.
- Schauer C, Janko C, Munoz LE, et al. Aggregated neutrophil extracellular traps limit inflammation by degrading cytokines and chemokines. *Nat Med.* 2014;20:511–517. doi: 10.1038/nm.3547.
- Kessenbrock K, Krumbholz M, Schönermarck U, Back W, Gross WL, Werb Z, Gröne HJ, Brinkmann V, Jenne DE. Netting neutrophils in autoimmune small-vessel vasculitis. *Nat Med.* 2009;15:623–625. doi: 10.1038/ nm.1959.
- 53. Garcia-Romo GS, Caielli S, Vega B, Connolly J, Allantaz F, Xu Z, Punaro M, Baisch J, Guiducci C, Coffman RL, Barrat FJ, Banchereau J, Pascual V. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci Transl Med.* 2011;3:73ra20. doi: 10.1126/scitranslmed.3001201.
- Döring Y, Manthey HD, Drechsler M, et al. Auto-antigenic protein-DNA complexes stimulate plasmacytoid dendritic cells to promote atherosclerosis. *Circulation*. 2012;125:1673–1683. doi: 10.1161/ CIRCULATIONAHA.111.046755.
- Massberg S, Grahl L, von Bruehl ML, et al. Reciprocal coupling of coagulation and innate immunity via neutrophil serine proteases. *Nat Med.* 2010;16:887–896. doi: 10.1038/nm.2184.
- Brinkmann V, Zychlinsky A. Neutrophil extracellular traps: is immunity the second function of chromatin? *J Cell Biol.* 2012;198:773–783. doi: 10.1083/jcb.201203170.
- Halverson TW, Wilton M, Poon KK, Petri B, Lewenza S. DNA is an antimicrobial component of neutrophil extracellular traps. *PLoS Pathog*. 2015;11:e1004593. doi: 10.1371/journal.ppat.1004593.
- Kolaczkowska E, Kubes P. Neutrophil recruitment and function in health and inflammation. *Nat Rev Immunol.* 2013;13:159–175. doi: 10.1038/nri3399.
- 59. Jaillon S, Peri G, Delneste Y, Frémaux I, Doni A, Moalli F, Garlanda C, Romani L, Gascan H, Bellocchio S, Bozza S, Cassatella MA, Jeannin P, Mantovani A. The humoral pattern recognition receptor PTX3 is stored in neutrophil granules and localizes in extracellular traps. *J Exp Med.* 2007;204:793–804. doi: 10.1084/jem.20061301.
- Urban CF, Ermert D, Schmid M, Abu-Abed U, Goosmann C, Nacken W, Brinkmann V, Jungblut PR, Zychlinsky A. Neutrophil extracellular traps contain calprotectin, a cytosolic protein complex involved in host defense against Candida albicans. *PLoS Pathog*. 2009;5:e1000639. doi: 10.1371/ journal.ppat.1000639.
- 61. Csomós K, Kristóf E, Jakob B, Csomós I, Kovács G, Rotem O, Hodrea J, Bagoly Z, Muszbek L, Balajthy Z, Csősz É, Fésüs L. Protein crosslinking by chlorinated polyamines and transglutamylation stabilizes neutrophil extracellular traps. *Cell Death Dis.* 2016;7:e2332. doi: 10.1038/ cddis.2016.200.
- Knight JS, Carmona-Rivera C, Kaplan MJ. Proteins derived from neutrophil extracellular traps may serve as self-antigens and mediate organ damage in autoimmune diseases. *Front Immunol.* 2012;3:380. doi: 10.3389/ fimmu.2012.00380.
- Rahman S, Gadjeva M. Does NETosis contribute to the bacterial pathoadaptation in cystic fibrosis? *Front Immunol.* 2014;5:378. doi: 10.3389/ fimmu.2014.00378.
- Menegazzi R, Decleva E, Dri P. Killing by neutrophil extracellular traps: fact or folklore? *Blood*. 2012;119:1214–1216. doi: 10.1182/ blood-2011-07-364604.

- 65. Xu Z, Cai J, Gao J, White GC 2nd, Chen F, Ma YQ. Interaction of kindlin-3 and β2-integrins differentially regulates neutrophil recruitment and NET release in mice. *Blood.* 2015;126:373–377. doi: 10.1182/ blood-2015-03-636720.
- Bianchi M, Hakkim A, Brinkmann V, Siler U, Seger RA, Zychlinsky A, Reichenbach J. Restoration of NET formation by gene therapy in CGD controls aspergillosis. *Blood.* 2009;114:2619–2622. doi: 10.1182/ blood-2009-05-221606.
- Li P, Li M, Lindberg MR, Kennett MJ, Xiong N, Wang Y. PAD4 is essential for antibacterial innate immunity mediated by neutrophil extracellular traps. *J Exp Med.* 2010;207:1853–1862. doi: 10.1084/jem.20100239.
- Martinod K, Fuchs TA, Zitomersky NL, Wong SL, Demers M, Gallant M, Wang Y, Wagner DD. PAD4-deficiency does not affect bacteremia in polymicrobial sepsis and ameliorates endotoxemic shock. *Blood*. 2015;125:1948–1956. doi: 10.1182/blood-2014-07-587709.
- Mohanty T, Sjögren J, Kahn F, Abu-Humaidan AH, Fisker N, Assing K, Mörgelin M, Bengtsson AA, Borregaard N, Sørensen OE. A novel mechanism for NETosis provides antimicrobial defense at the oral mucosa. *Blood*. 2015;126:2128–2137. doi: 10.1182/blood-2015-04-641142.
- Pham CT, Ivanovich JL, Raptis SZ, Zehnbauer B, Ley TJ. Papillon-Lefèvre syndrome: correlating the molecular, cellular, and clinical consequences of cathepsin C/dipeptidyl peptidase I deficiency in humans. J Immunol. 2004;173:7277–7281.
- Sørensen OE, Clemmensen SN, Dahl SL, Østergaard O, Heegaard NH, Glenthøj A, Nielsen FC, Borregaard N. Papillon-Lefèvre syndrome patient reveals species-dependent requirements for neutrophil defenses. *J Clin Invest.* 2014;124:4539–4548. doi: 10.1172/JCI76009.
- Finn AV, Nakano M, Narula J, Kolodgie FD, Virmani R. Concept of vulnerable/unstable plaque. Arterioscler Thromb Vasc Biol. 2010;30:1282– 1292. doi: 10.1161/ATVBAHA.108.179739.
- Megens RT, Vijayan S, Lievens D, Döring Y, van Zandvoort MA, Grommes J, Weber C, Soehnlein O. Presence of luminal neutrophil extracellular traps in atherosclerosis. *Thromb Haemost*. 2012;107:597–598. doi: 10.1160/TH11-09-0650.
- Quillard T, Araújo HA, Franck G, Shvartz E, Sukhova G, Libby P. TLR2 and neutrophils potentiate endothelial stress, apoptosis and detachment: implications for superficial erosion. *Eur Heart J.* 2015;36:1394–1404. doi: 10.1093/eurheartj/ehv044.
- Macritchie N, Grassia G, Sabir SR, Maddaluno M, Welsh P, Sattar N, Ialenti A, Kurowska-Stolarska M, McInnes IB, Brewer JM, Garside P, Maffia P. Plasmacytoid dendritic cells play a key role in promoting atherosclerosis in apolipoprotein E-deficient mice. *Arterioscler Thromb Vasc Biol.* 2012;32:2569–2579. doi: 10.1161/ATVBAHA.112.251314.
- Carmona-Rivera C, Zhao W, Yalavarthi S, Kaplan MJ. Neutrophil extracellular traps induce endothelial dysfunction in systemic lupus erythematosus through the activation of matrix metalloproteinase-2. *Ann Rheum Dis.* 2015;74:1417–1424. doi: 10.1136/annrheumdis-2013-204837.
- Villanueva E, Yalavarthi S, Berthier CC, et al. Netting neutrophils induce endothelial damage, infiltrate tissues, and expose immunostimulatory molecules in systemic lupus erythematosus. *J Immunol.* 2011;187:538– 552. doi: 10.4049/jimmunol.1100450.
- Gupta AK, Joshi MB, Philippova M, Erne P, Hasler P, Hahn S, Resink TJ. Activated endothelial cells induce neutrophil extracellular traps and are susceptible to NETosis-mediated cell death. *FEBS Lett.* 2010;584:3193– 3197. doi: 10.1016/j.febslet.2010.06.006.
- Knight JS, Luo W, O'Dell AA, Yalavarthi S, Zhao W, Subramanian V, Guo C, Grenn RC, Thompson PR, Eitzman DT, Kaplan MJ. Peptidylarginine deiminase inhibition reduces vascular damage and modulates innate immune responses in murine models of atherosclerosis. *Circ Res.* 2014;114:947–956. doi: 10.1161/CIRCRESAHA.114.303312.
- Soehnlein O, Ortega-Gómez A, Döring Y, Weber C. Neutrophilmacrophage interplay in atherosclerosis: protease-mediated cytokine processing versus NET release. *Thromb Haemost.* 2015;114:866–867. doi: 10.1160/TH15-08-0623.
- 81. Joosten LA, Netea MG, Fantuzzi G, Koenders MI, Helsen MM, Sparrer H, Pham CT, van der Meer JW, Dinarello CA, van den Berg WB. Inflammatory arthritis in caspase 1 gene-deficient mice: contribution of proteinase 3 to caspase 1-independent production of bioactive interleukin-1beta. *Arthritis Rheum*. 2009;60:3651–3662. doi: 10.1002/art.25006.
- 82. Borissoff JI, Joosen IA, Versteylen MO, Brill A, Fuchs TA, Savchenko AS, Gallant M, Martinod K, Ten Cate H, Hofstra L, Crijns HJ, Wagner DD, Kietselaer BL. Elevated levels of circulating DNA and chromatin are independently associated with severe coronary atherosclerosis and a pro-thrombotic state. *Arterioscler Thromb Vasc Biol.* 2013;33:2032–2040. doi: 10.1161/ATVBAHA.113.301627.

- Kimball AS, Obi AT, Diaz JA, Henke PK. The emerging role of NETs in venous thrombosis and immunothrombosis. *Front Immunol*. 2016;7:236. doi: 10.3389/fimmu.2016.00236.
- von Brühl ML, Stark K, Steinhart A, et al. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. J Exp Med. 2012;209:819–835. doi: 10.1084/jem.20112322.
- Fuchs TA, Brill A, Duerschmied D, Schatzberg D, Monestier M, Myers DD Jr, Wrobleski SK, Wakefield TW, Hartwig JH, Wagner DD. Extracellular DNA traps promote thrombosis. *Proc Natl Acad Sci U S A*. 2010;107:15880–15885. doi: 10.1073/pnas.1005743107.
- Libby P, Ridker PM, Hansson GK. Progress and challenges in translating the biology of atherosclerosis. *Nature*. 2011;473:317–325. doi: 10.1038/ nature10146.
- Wyss CA, Neidhart M, Altwegg L, Spanaus KS, Yonekawa K, Wischnewsky MB, Corti R, Kucher N, Roffi M, Eberli FR, Amann-Vesti B, Gay S, von Eckardstein A, Lüscher TF, Maier W. Cellular actors, Tolllike receptors, and local cytokine profile in acute coronary syndromes. *Eur Heart J*. 2010;31:1457–1469. doi: 10.1093/eurheartj/ehq084.
- Horne BD, Anderson JL, John JM, Weaver A, Bair TL, Jensen KR, Renlund DG, Muhlestein JB; Intermountain Heart Collaborative Study Group. Which white blood cell subtypes predict increased cardiovascular risk? J Am Coll Cardiol. 2005;45:1638–1643. doi: 10.1016/j.jacc.2005.02.054.
- Distelmaier K, Winter MP, Dragschitz F, Redwan B, Mangold A, Gleiss A, Perkmann T, Maurer G, Adlbrecht C, Lang IM. Prognostic value of culprit site neutrophils in acute coronary syndrome. *Eur J Clin Invest*. 2014;44:257–265. doi: 10.1111/eci.12228.
- Distelmaier K, Adlbrecht C, Jakowitsch J, Winkler S, Dunkler D, Gerner C, Wagner O, Lang IM, Kubicek M. Local complement activation triggers neutrophil recruitment to the site of thrombus formation in acute myocardial infarction. *Thromb Haemost.* 2009;102:564–572. doi: 10.1160/ TH09-02-0103.
- 91. Riegger J, Byrne RA, Joner M, et al.; Prevention of Late Stent Thrombosis by an Interdisciplinary Global European Effort (PRESTIGE) Investigators. Histopathological evaluation of thrombus in patients presenting with stent thrombosis. A multicenter European study: a report of the prevention of late stent thrombosis by an interdisciplinary global European effort consortium. Eur Heart J. 2016;37:1538–1549. doi: 10.1093/eurheartj/ehv419.
- Oklu R, Albadawi H, Watkins MT, Monestier M, Sillesen M, Wicky S. Detection of extracellular genomic DNA scaffold in human thrombus: implications for the use of deoxyribonuclease enzymes in thrombolysis. J Vasc Interv Radiol. 2012;23:712–718. doi: 10.1016/j.jvir.2012.01.072.
- Mangold A, Alias S, Scherz T, et al. Coronary neutrophil extracellular trap burden and deoxyribonuclease activity in ST-elevation acute coronary syndrome are predictors of ST-segment resolution and infarct size. *Circ Res.* 2015;116:1182–1192. doi: 10.1161/CIRCRESAHA.116.304944.

- 94. Maugeri N, Campana L, Gavina M, Covino C, De Metrio M, Panciroli C, Maiuri L, Maseri A, D'Angelo A, Bianchi ME, Rovere-Querini P, Manfredi AA. Activated platelets present high mobility group box 1 to neutrophils, inducing autophagy and promoting the extrusion of neutrophil extracellular traps. J Thromb Haemost. 2014;12:2074–2088. doi: 10.1111/jth.12710.
- Stark K, Philippi V, Stockhausen S, et al. Disulfide HMGB1 derived from platelets coordinates venous thrombosis in mice. *Blood*. 2016;128:2435– 2449. doi: 10.1182/blood-2016-04-710632.
- 96. de Boer OJ, Li X, Teeling P, Mackaay C, Ploegmakers HJ, van der Loos CM, Daemen MJ, de Winter RJ, van der Wal AC. Neutrophils, neutrophil extracellular traps and interleukin-17 associate with the organisation of thrombi in acute myocardial infarction. *Thromb Haemost*. 2013;109:290–297. doi: 10.1160/TH12-06-0425.
- Darbousset R, Thomas GM, Mezouar S, Frère C, Bonier R, Mackman N, Renné T, Dignat-George F, Dubois C, Panicot-Dubois L. Tissue factor-positive neutrophils bind to injured endothelial wall and initiate thrombus formation. *Blood.* 2012;120:2133–2143. doi: 10.1182/blood-2012-06-437772.
- Stakos DA, Kambas K, Konstantinidis T, Mitroulis I, Apostolidou E, Arelaki S, Tsironidou V, Giatromanolaki A, Skendros P, Konstantinides S, Ritis K. Expression of functional tissue factor by neutrophil extracellular traps in culprit artery of acute myocardial infarction. *Eur Heart J*. 2015;36:1405–1414. doi: 10.1093/eurheartj/ehv007.
- Rangé H, Labreuche J, Louedec L, Rondeau P, Planesse C, Sebbag U, Bourdon E, Michel JB, Bouchard P, Meilhac O. Periodontal bacteria in human carotid atherothrombosis as a potential trigger for neutrophil activation. *Atherosclerosis*. 2014;236:448–455. doi: 10.1016/j. atherosclerosis.2014.07.034.
- 100. Ge L, Zhou X, Ji WJ, Lu RY, Zhang Y, Zhang YD, Ma YQ, Zhao JH, Li YM. Neutrophil extracellular traps in ischemia-reperfusion injury-induced myocardial no-reflow: therapeutic potential of DNase-based reperfusion strategy. *Am J Physiol Heart Circ Physiol*. 2015;308:H500–H509. doi: 10.1152/ajpheart.00381.2014.
- 101. Savchenko AS, Borissoff JI, Martinod K, De Meyer SF, Gallant M, Erpenbeck L, Brill A, Wang Y, Wagner DD. VWF-mediated leukocyte recruitment with chromatin decondensation by PAD4 increases myocardial ischemia/reperfusion injury in mice. *Blood.* 2014;123:141–148. doi: 10.1182/blood-2013-07-514992.
- 102. De Meyer SF, Suidan GL, Fuchs TA, Monestier M, Wagner DD. Extracellular chromatin is an important mediator of ischemic stroke in mice. *Arterioscler Thromb Vasc Biol*. 2012;32:1884–1891. doi: 10.1161/ ATVBAHA.112.250993.
- Vogel B, Shinagawa H, Hofmann U, Ertl G, Frantz S. Acute DNasel treatment improves left ventricular remodeling after myocardial infarction by disruption of free chromatin. *Basic Res Cardiol.* 2015;110:15. doi: 10.1007/s00395-015-0472-y.