

16. Mouse cerebellar cultures were prepared and maintained by the method of K. Schilling, M. Dickinson, J. Connor, and J. Morgan (*Neuron*, in press) as previously described (6). Cultures used for recording had been maintained in vitro for 12 to 24 days. PNs were chosen for recording by distinct morphological criteria (large round soma, prominent dendritic arbor) as verified by staining with the PN-specific immunohistochemical markers PEP-19 [L. Sangameswaran, J. Hempstead, J. Morgan, *Proc. Natl. Acad. Sci. U.S.A.* **86**, 5651 (1989)] and calbindin-D<sub>28k</sub> [M. Thomasset *et al.*, *Dev. Pharmacol. Ther.* **7**, 6 (1984)]. Nystatin patch electrodes [R. Horn and A. Marty, *J. Gen. Physiol.* **92**, 145 (1988)] were attached to PN somata and were used to apply a holding potential of  $-80$  mV. Iontophoresis electrodes (tip diameter,  $<0.5$   $\mu$ m) were filled with excitatory amino acid receptor agonists and were positioned  $\sim 20$   $\mu$ m away from a large-caliber dendrite 10 to 20  $\mu$ m from the soma. Pulses of agonists (30 to 100 ms duration) were applied at a frequency of 0.05 Hz. After acquisition of 15 min of baseline responses, we induced LTD by pairing six successive glutamate test pulses with six 4-s depolarization steps to  $-10$  mV, timed so that the depolarization onset preceded the glutamate pulse by 500 ms. This treatment reliably induced a stable depression of the glutamate-mediated inward current (typically 50 to 65% of its baseline value) that was independent of alterations in input resistance or resting potential (6; D. J. Linden and J. A. Connor, in preparation). Cells were bathed in a solution (adjusted to pH 7.35 with NaOH) that contained NaCl (150 mM), KCl (5 mM), CaCl<sub>2</sub> (2 mM), MgCl<sub>2</sub> (0.8 mM), Hepes (10 mM), glucose (10 mM), tetrodotoxin (0.005 mM), and picrotoxin (0.1 mM). Tetrodotoxin and picrotoxin were added to the external saline to improve the resolution of the voltage clamp and to block spontaneous synaptic potentials, but the addition of these compounds was not required for the induction of LTD. For recordings of voltage-gated Ca<sup>2+</sup> currents (as in Fig. 1C), 10 mM TEA-Cl was added. The internal saline (adjusted to pH 7.35 with KOH) consisted of K<sub>2</sub>SO<sub>4</sub> (95 mM), KCl (15 mM), MgCl<sub>2</sub> (8 mM), and Hepes (10 mM). For Ca<sup>2+</sup> current recording, Cs<sub>2</sub>SO<sub>4</sub> and CsCl were substituted for K<sub>2</sub>SO<sub>4</sub> and KCl, respectively, in an equimolar fashion. Patch electrodes were pulled from N51A glass (Garner Glass Co., Claremont, CA) and polished on a microforge to yield a resistance of 1 to 3 megohms when measured with normal internal and external salines. Nystatin (250  $\mu$ g/ml) was added to the electrode filling saline. This concentration gave access resistance to the cell that was typically  $<10$  megohms. Only cells with stable resting potentials of  $> -45$  mV were used. Membrane currents induced by application of excitatory amino acids were stored on a chart recorder that imposed a 0- to 30-Hz bandpass. Membrane currents in response to voltage steps were filtered at 3 kHz, digitized with a 12-bit analog-to-digital converter operating at a sampling frequency of 20 kHz, and stored on the hard disk of an INDEC-LSI-11/23 computer. Leakage current was determined from 10-mV hyperpolarizing pulses and was digitally subtracted from records of voltage-gated currents. Experiments were conducted at 22° to 24°C. Imaging of intracellular free Ca<sup>2+</sup> with the indicator fura-2/AM was performed as described [J. A. Connor, *Proc. Natl. Acad. Sci. U.S.A.* **83**, 6179 (1986)].
17. Excitatory amino acid receptor agonists were delivered by iontophoresis and were prepared in 10 mM Hepes as follows: 10 mM glutamate (pH 7.1); 10 mM quisqualate (pH 7.2); 10 mM AMPA (pH 7.1); 100 mM NMDA (pH 7.3). These compounds were delivered through polished patch pipettes (resistance, 4 to 6 megohms when filled with normal internal saline) with negative current pulses. Braking currents were applied to prevent drug leakage.
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19. P. D. Davis *et al.*, *FEBS Lett.* **259**, 61 (1989). RO-31-8220 is referred to as compound 3 in this reference. The following inhibition concentration (IC<sub>50</sub>) values are reported for RO-31-8220 inhibition of purified protein kinases as measured in the presence of ATP (10  $\mu$ M), with a histone protein substrate: PKC, 0.01  $\mu$ M; cAMP-dependent kinase, 1.5  $\mu$ M; Ca<sup>2+</sup>/calmodulin dependent kinase, 17  $\mu$ M.
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22. D. J. Linden and J. A. Connor, unpublished observations.
23. Studies with exogenous agonists have shown a developmental decrease in the response of PNs to NMDA, resulting in a complete lack of sensitivity to this compound in the adult [J. L. Dupont, G. Gardette, F. Crepel, *Dev. Brain Res.* **34**, 59 (1987); G. Garthwaite, B. Yamani, J. Garthwaite, *ibid.* **36**, 288 (1987); E. Audinat, T. Knopfel, B. H. Gähwiler, *J. Physiol. (London)* **430**, 297 (1990)]. Our cultures of embryonic PNs showed NMDA responses typical of those seen in other neuronal types: that is, they were attenuated by DL-2-amino-5-phosphonopentanoic acid (AP5) and exhibited a Mg<sup>2+</sup> and voltage-dependent block. When the membrane potential was clamped to  $-80$  mV, the NMDA response became very small ( $<10\%$  of the glutamate current was blocked by 20  $\mu$ M AP5 at this potential). To produce NMDA currents of easily measurable amplitude (as in Fig. 3), membrane potential was clamped to  $-65$  mV, and NMDA pulses were typically applied for durations two to three times that of AMPA.
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27. We thank K. Schilling, who developed the cell culture technique used in this study; M. Smeyne and R. Sun for technical assistance; W. Muller, M. Dickinson, and J. Petrozzino for helpful discussion; and G. Lawton (Roche Welwyn, United Kingdom) for providing RO-31-8220.

8 July 1991; accepted 10 September 1991

## Targeting of the Master Receptor MOM19 to Mitochondria

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The targeting of proteins to mitochondria involves the recognition of the precursor proteins by receptors on the mitochondrial surface followed by insertion of the precursors into the outer membrane at the general insertion site GIP. Most mitochondrial proteins analyzed so far use a mitochondrial outer membrane protein of 19 kilodaltons (MOM19) as an import receptor. The gene encoding MOM19 has now been isolated. The deduced amino acid sequence predicts that MOM19 is anchored in the outer membrane by an NH<sub>2</sub>-terminal hydrophobic sequence, while the rest of the protein forms a hydrophilic domain exposed to the cytosol. MOM19 was targeted to the mitochondria via a pathway that is independent of protease-accessible surface receptors and controlled by direct assembly of the MOM19 precursor with GIP.

THE BIOGENESIS OF MITOCHONDRIA involves the translocation of cytosolically synthesized precursor proteins into or across the organellar membranes (1, 2). In *Neurospora crassa*, two mitochondrial outer membrane proteins (MOMs) were identified as import receptors for precursor proteins. The 19-kD protein MOM19 functions as a receptor for most precursors analyzed, including all precursors that carry an NH<sub>2</sub>-terminal signal sequence (3–5), and is referred to as the master receptor. The 72-

kD protein MOM72 is a receptor for the precursor of the adenosine diphosphate-adenosine triphosphate (ADP-ATP) carrier, which contains internal targeting signals (4, 6). Precursor proteins interact with these receptors and are inserted into the outer membrane at a common site, termed the general insertion site (general insertion protein = GIP) (7, 8). The 38-kD protein MOM38 forms part of the GIP and is assembled with MOM19 and MOM72 in the mitochondrial receptor complex (9). To determine the mechanism by which the master receptor is targeted to mitochondria, we investigated the biogenesis of MOM19.

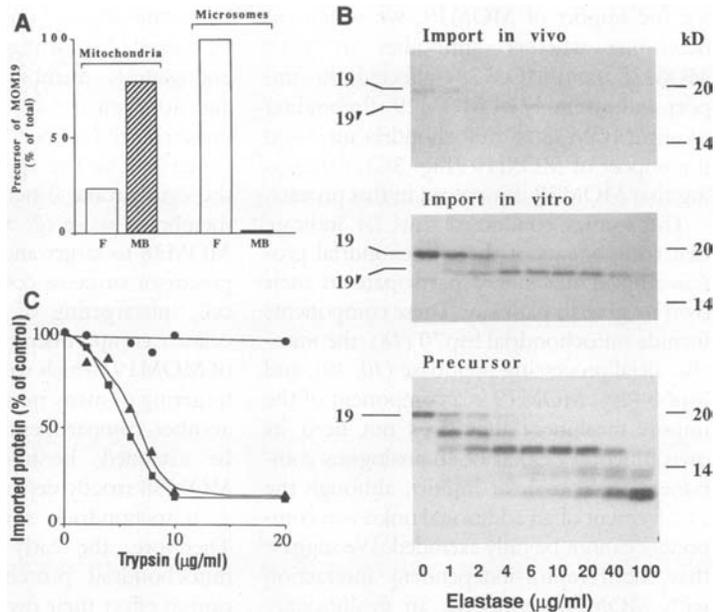
A full-length cDNA clone that encodes MOM19 from *N. crassa* was isolated (10) and used to obtain the complete MOM19 gene (11). The nucleotide sequences of both

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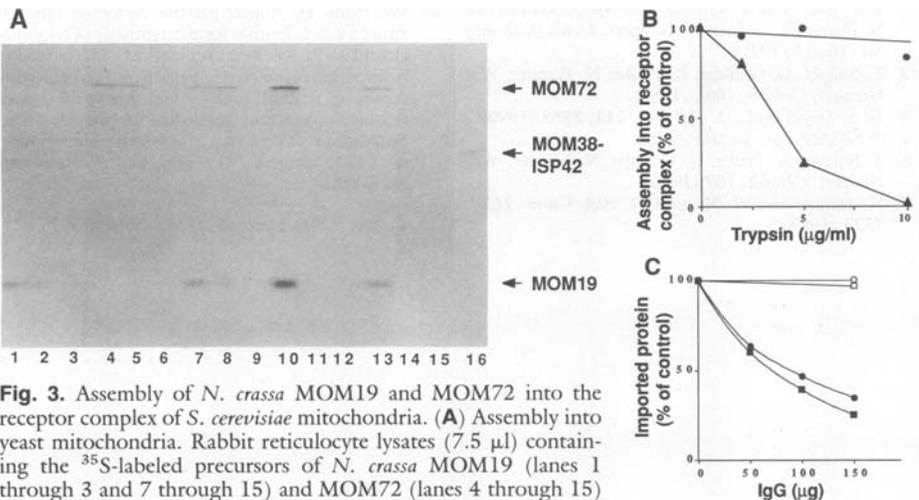
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**Fig. 2.** Lack of a requirement of a surface receptor for the import of MOM19 into *N. crassa* mitochondria. **(A)** The precursor of MOM19 efficiently binds to isolated mitochondria but not to microsomes. MOM19 (cDNA cloned into pGEM4) was synthesized in rabbit reticulocyte lysate in the presence of [<sup>35</sup>S]methionine (26). Lysate (2  $\mu$ l) was incubated with isolated mitochondria (10  $\mu$ g of protein) or microsomes (10  $\mu$ g of protein) for 15 min at 25°C in a final volume of 75  $\mu$ l (including 100 mM of KCl) as described (26). The membranes were reisolated by centrifugation and washed in 100 mM sucrose, 1 mM EDTA, 10 mM MOPS, pH 7.2, 100 mM KCl, and the proteins were analyzed as described in the caption of Fig. 1B. The values are averages of four determinations with SEMs less than 7%. The precursor of MOM19 showed the same apparent size as the mature protein, and its import into mitochondria did not require a membrane potential across the inner membrane, as is generally found with outer membrane proteins (2). F, free; MB, membrane bound. **(B)** Formation of a characteristic 17-kD fragment (MOM19') of imported MOM19. MOM19 synthesized in vitro was imported into isolated mitochondria for 7 min at 25°C. After treatment with elastase, the mitochondria were reisolated, washed, and analyzed by SDS-PAGE and fluorography. From parallel samples, mitochondria were transferred to nitrocellulose and treated with antibodies against MOM19 (3) (import in vivo). Rabbit reticulocyte lysate containing the precursor of MOM19 was treated with elastase under the conditions used for the treatment of mitochondria (3) (precursor). **(C)** Import of MOM19 into mitochondria does not require a surface receptor. Mitochondria were pretreated with trypsin (26) and incubated with reticulocyte lysate containing the precursors of MOM19 (●), F<sub>1</sub>B (▲), or AAC (■) for 5 min at 15°C, followed by a treatment with elastase. The amount of protein imported into mitochondria not treated with trypsin was set to 100% (control). The values are averages of five



determinations with SEMs less than 5%. The rate of import linearly depended on the time of incubation and the amount of mitochondria. Preincubation of the mitochondria with antibodies to MOM19 (3) also did not inhibit the import of newly synthesized MOM19.



**Fig. 3.** Assembly of *N. crassa* MOM19 and MOM72 into the receptor complex of *S. cerevisiae* mitochondria. **(A)** Assembly into yeast mitochondria. Rabbit reticulocyte lysates (7.5  $\mu$ l) containing the <sup>35</sup>S-labeled precursors of *N. crassa* MOM19 (lanes 1 through 3 and 7 through 15) and MOM72 (lanes 4 through 15) were incubated with isolated yeast mitochondria (25  $\mu$ g of protein) for 5 min at 25°C (26) (lanes 10 through 12 received mitochondria lysed with buffer containing 0.5% digitonin). The mitochondria of lanes 1 through 9 and 13 through 15 were reisolated, washed in 100 mM sucrose, 1 mM EDTA, 10 mM MOPS, pH 7.2, and solubilized in buffer containing 0.5% digitonin (lanes 1 through 9) (9) or in SDS-containing buffer (lanes 13 through 15) (3, 9). Immunoprecipitations were performed with antibodies directed to *N. crassa* MOM19 (lanes 1, 7, 10, and 13), MOM72 (lanes 4, 7, 10, and 13), yeast MOM38 (=ISP42) (lanes 2, 5, 8, 11, and 14), and preimmune antibodies (lanes 3, 6, 9, 12, and 15) (9). The specificity of the antibodies to yeast MOM38 was shown by immunoprecipitation of SDS-lysed yeast mitochondria labeled with <sup>35</sup>S (lane 16). **(B)** Assembly of MOM19 does not require a known surface receptor. Yeast mitochondria were pretreated with trypsin (26). *N. crassa* MOM19 (●) and MOM72 (▲) were imported, and coimmunoprecipitations with anti-MOM38-ISP42 antibodies were performed as described above. The amount of protein that coprecipitated with mitochondria that were not treated with trypsin was set to 100% (control) (similar results were obtained when the import was performed at lower temperature, 15°C or 0°C). The values are averages of five determinations with SEMs less than 6%. **(C)** MOM38 (=ISP42) is needed for import of MOM19. Isolated yeast mitochondria (10  $\mu$ g) were preincubated with IgGs to MOM38 (closed symbols) or preimmune IgGs (open symbols) (3, 6, 26). Precursors of MOM19 (●) and AAC (■) were imported, and the mitochondria were treated with elastase as described above. For import of MOM19, the mitochondria were pretreated with trypsin (5  $\mu$ g/ml). The amount of protein imported into mitochondria not carrying IgGs was set to 100% (control). The values are averages of at least three determinations with SEMs less than 8%.

nizing the native protein are not available (9), rendering a direct assay with *N. crassa* mitochondria impossible. The *Saccharomyces cerevisiae* equivalent of MOM38, also termed ISP42 (15, 16), is more easily accessible to antibodies, probably as a result of an extra sequence of about 20 amino acid residues at the NH<sub>2</sub> terminus (9). We prepared monospecific antibodies to yeast MOM38 (Fig. 3A) that efficiently recognized the native protein (anti-MOM38). MOM19 from *N. crassa* was imported into yeast mitochondria and assembled into the yeast receptor complex, as shown by the coprecipitation of MOM19 with anti-MOM38 (Fig. 3A, lanes 2 and 8). Assembly with MOM38 did not occur when reticulocyte lysate containing the radioactively labeled precursor was incubated with lysed mitochondria (Fig. 3A, lane 11), indicating that import of the MOM19 precursor into intact mitochondria is a prerequisite for the assembly of MOM19 with MOM38. MOM72 from *N. crassa* also correctly assembled into the receptor complex of yeast mitochondria (Fig. 3A, lanes 5 and 8), showing a high conservation of the mitochondrial protein import machinery between *N. crassa* and *S. cerevisiae*.

Yeast mitochondria were pretreated with trypsin so that surface receptors, but not MOM38, were degraded, leading to inhibition of import of various precursor proteins (Fig. 3B) (4, 17). Import and assembly of MOM19, however, were still possible. To further analyze whether MOM38 is needed

for the import of MOM19, we sought to determine whether antibodies to yeast MOM38 (anti-MOM38) affected the import and assembly of MOM19. Prebinding of anti-MOM38 to mitochondria inhibited the import of MOM19 (Fig. 3C), confirming that MOM38 is involved in this process.

The studies conducted thus far indicate that components of the mitochondrial protein import machinery participate in their own biogenesis pathway. These components include mitochondrial hsp70 (18), the mitochondrial processing peptidase (10, 19), and hsp60 (20). MOM19 is a component of the import machinery that does not need its own function or that of an analogous component (receptor) for import, although the involvement of an additional unknown component cannot be fully excluded. We suggest that the receptor-independent interaction with MOM38 represents an evolutionary remnant form of mitochondrial protein import, allowing specific import without a need for surface receptors. This finding sheds new light on a previous unexpected result, namely, the import of a number of authentic mitochondrial precursor proteins into mitochondria that lack surface receptors (21). This so-called bypass import occurred with a low efficiency in most but not all cases; its mechanism and specificity remained unclear. The import of MOM19 shows that the interaction with MOM38 allows a specific and highly efficient import in the absence of surface receptors, raising a similar possibility for other precursor proteins [this pathway has to be distinguished

from the unique import pathway of the precursor of cytochrome c that possesses an endogenous membrane insertion activity and does not use any of the known MOM proteins; the import of this precursor is controlled by the specific interaction with the cytochrome c heme lyase in the intermembrane space (2, 3, 11)]. The ability of MOM38 to target and insert mitochondrial precursor proteins could be harmful to the cell; mistargeting of MOM38 to another cellular compartment might lead to import of MOM19, which would allow the efficient targeting of many mitochondrial proteins to another compartment. This possibility can be excluded, because the biogenesis of MOM38 strictly depends on the presence of a mitochondrial surface receptor (22). Therefore, the early components of the mitochondrial protein import machinery cannot effect their own import, so an effective control system exists for the specific assembly of the protein import apparatus only at the appropriate location.

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27. We thank H. Bujard for the vector pUHE, G. Arnold and I. Leitner for the synthesis of oligonucleotides, R. Zimmermann and M. Sagstetter for isolated microsomes, H. Feldmann for help with computer analysis, and U. Hanemann for expert technical assistance. Supported by the Deutsche Forschungsgemeinschaft (Sonderforschungsbereich 184, project B1) and the Genzentrum München.

11 July 1991; accepted 20 September 1991