



# Draft Genome Sequence of *Phyllobacterium endophyticum* mTS5, Isolated from *Lupinus micranthus* in Tunisia

Zoé Waller,<sup>a</sup> Mokhtar Rejili,<sup>b</sup> Mohamed Mars,<sup>b</sup> Andreas Brachmann,<sup>a</sup>  Macarena Marín<sup>a</sup>

<sup>a</sup>Genetics, Faculty of Biology, Ludwig-Maximilians-University Munich, Munich, Germany

<sup>b</sup>Laboratory of Biodiversity and Valorization of Arid Areas Bioresources (BVBA), Faculty of Sciences, Gabès University, Gabès, Tunisia

**ABSTRACT** We report here the draft genome sequence of *Phyllobacterium endophyticum* mTS5, isolated from a *Lupinus micranthus* root nodule. The genome consists of 5,454,168 bp, with a GC content of 57%, and contains 5,676 protein-coding sequences.

Lupines are emerging crops for human and livestock diets because of their protein- and fiber-rich seeds and their low requirements for nitrogen and phosphorus fertilization (1). Their ability to grow in nitrogen-poor soils is grounded in their symbiotic association with nitrogen-fixing bacteria (2). Thus, assessing the biodiversity of bacteria associated with lupines is of paramount importance for the use of these crops in sustainable agriculture.

*Phyllobacterium endophyticum* mTS5 was isolated from a single nodule on a *Lupinus micranthus* plant growing in the region of Takelsa, Tunisia. A whole nodule was dissected from a root and surface sterilized for 1 min with 95% ethanol and for 3 min with 4% sodium hypochlorite. The nodule was washed at least 5 times, and a 50- $\mu$ l aliquot of the last washing suspension was checked for sterility on yeast mannitol agar (YMA) plates (3). The nodule was then separately crushed in a 20- $\mu$ l aliquot of sterile water, and its content was streaked on YMA plates. These were incubated at 28°C under aerobic conditions for 5 to 7 days. Bacterial colonies were purified by repeated isolation at least 5 times.

Genomic DNA (gDNA) was extracted using the cetyltrimethylammonium bromide (CTAB) method (4) from a single-colony culture grown for 3 days in yeast mannitol broth. DNA was sheared on a Covaris M220 ultrasonicator with Covaris MicroCaps (50  $\mu$ l) to approximately 600 bp. Two libraries were prepared using the sparQ DNA library prep kit (QuantaBio), according to the manufacturer's instructions, with 600 ng and 107 ng of sheared gDNA, respectively. Libraries were quality controlled with a high-sensitivity DNA kit on a Bioanalyzer instrument (Agilent) for amplicon sizes between 300 and 800 bp and absence of adapter primers and then quantified on a Qubit 2.0 fluorometer (with a double-stranded DNA [dsDNA] high-sensitivity [HS] assay kit; Thermo Fisher Scientific). The genome was sequenced in the Genomics Service Unit (LMU Biocenter, Munich, Germany) on an Illumina MiSeq system with v3 chemistry (five partial rounds of paired-end sequencing with 2  $\times$  250 bp or 2  $\times$  300 bp), generating 5,765,152 paired-end reads. Adapter and quality trimming (trim limit, 0.01) resulted in 2,810,080 paired-end reads with an average length of 183 bp, from which the genome was assembled *de novo* using CLC Genomics Workbench 9.0 (Qiagen). Default parameters were used for all software unless otherwise specified. Quality control of the assembly was performed using QUAST 5.0.2 (5). In total, 93 contigs were obtained, with 5,454,168 bp of sequence information and 93-fold coverage. The average contig length was 58,081 bp, with the largest contig being 366,285 bp and the shortest being 511 bp. The  $N_{50}$  value was 165,103 bp, and the GC content 57%. The public version of the

**Citation** Waller Z, Rejili M, Mars M, Brachmann A, Marín M. 2019. Draft genome sequence of *Phyllobacterium endophyticum* mTS5, isolated from *Lupinus micranthus* in Tunisia. *Microbiol Resour Announc* 8:e00968-19. <https://doi.org/10.1128/MRA.00968-19>.

**Editor** David A. Baltrus, University of Arizona

**Copyright** © 2019 Waller et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Macarena Marín, [m.marin@biologie.uni-muenchen.de](mailto:m.marin@biologie.uni-muenchen.de).

This article is contribution 1 from the Chair of Genetics of the Ludwig-Maximilians-University Munich.

**Received** 21 August 2019

**Accepted** 14 October 2019

**Published** 7 November 2019

genome was annotated with the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (6), while the functional annotation of the in-house version was performed using the Rapid Annotations using Subsystems Technology (RAST) server for homology search against the SEED database (7). For the latter, in total, 5,676 protein-coding sequences and 49 RNA genes were predicted. A BLASTn search using the *nodC* and *nifH* orthologs of *Phyllobacterium sophorae* CCBAU 03422 (GenBank accession numbers [KJ685943](#) and [KJ685942](#), respectively) and *Mesorhizobium japonicum* MAFF303099 (accession number [BA000012](#)) against the *Phyllobacterium endophyticum* mT55 genome revealed the lack of the *nod* and *nif* symbiotic clusters in this strain.

A pairwise comparison based on the 16S rRNA and *recA* gene sequences was generated in CLC Main Workbench 7.0 (Qiagen). The type strain *Phyllobacterium endophyticum* PEPV15 was identified as the closest strain. Average nucleotide identity analysis was performed using the OrthoANIu algorithm (8). The obtained value, 98.56%, is above the species cutoff (9), which supports systematic placement of strain mT55 in the species *Phyllobacterium endophyticum*.

**Data availability.** The draft genome sequence has been deposited at NCBI/GenBank under BioProject number [PRJNA559855](#), BioSample number [SAMN12561461](#), accession number [VOWA00000000](#), and SRA numbers [SRR9985975](#) and [SRR9985976](#).

## ACKNOWLEDGMENTS

This study was supported by the Ministry of Higher Education and Research Development of Tunisia (MESRS) and by the German Federal Ministry of Education and Research (BMBF) in a joint grant (grant 01DH16008).

## REFERENCES

- Lucas MM, Stoddard FL, Annicchiarico P, Frias J, Martinez-Villaluenga C, Sussmann D, Duranti M, Seger A, Zander PM, Pueyo JJ. 2015. The future of lupin as a protein crop in Europe. *Front Plant Sci* 6:705. <https://doi.org/10.3389/fpls.2015.00705>.
- Evans J, Turner GL, O'Connor GE, Bergersen FJ. 1987. Nitrogen fixation and accretion of soil nitrogen by field-grown lupins (*Lupinus angustifolius*). *Field Crops Res* 16:309–322. [https://doi.org/10.1016/0378-4290\(87\)90069-4](https://doi.org/10.1016/0378-4290(87)90069-4).
- Vincent JM. 1970. The cultivation, isolation and maintenance of rhizobia, p. 1–13. *In* A manual for the practical study of root-nodule. Blackwell Scientific Publications, Oxford, United Kingdom.
- William S, Feil H, Copeland A. 2012. Bacterial genomic DNA isolation using CTAB. DOE Joint Genome Institute, Walnut Creek, CA.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44: 6614–6624. <https://doi.org/10.1093/nar/gkw569>.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the rapid annotation of microbial genomes using subsystems technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Lee I, Ouk Kim Y, Park S-C, Chun J. 2016. OrthoANI: an improved algorithm and software for calculating average nucleotide identity. *Int J Syst Evol Microbiol* 66:1100–1103. <https://doi.org/10.1099/ijs.0.000760>.
- Richter M, Rosselló-Móra R. 2009. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 106:19126–19131. <https://doi.org/10.1073/pnas.0906412106>.