An Amphitropic cAMP-Binding Protein in Yeast Mitochondria. 1. Synergistic Control of the Intramitochondrial Location by Calcium and Phospholipid

Günter Müller* and Wolfhard Bandlows†
Laboratorium für molekulare Zellbiologie und allgemeine Pathologie, Institut für Pathologie und Rechtsmedizin der Universität Ulm, Neuherbergstrasse 11, D-8000 München 45, Federal Republic of Germany, and Institut für Genetik und Mikrobiologie der Universität München, Maria-Ward-Strasse 1a, D-8000 München 19, Federal Republic of Germany

Received March 3, 1989; Revised Manuscript Received July 14, 1989

ABSTRACT: A cAMP-binding protein is found to be integrated into the inner mitochondrial membrane of the yeast Saccharomyces cerevisiae under normal conditions. It resists solubilization by high salt and chaotropic agents. The protein is, however, converted to a soluble form which then resides in the intermembrane space, when isolated mitochondria are incubated with low concentrations of calcium. Phospholipids or diacylglycerol (or analogues) dramatically increases the efficiency of receptor release from the inner membrane, whereas these compounds alone are ineffective. Also, cAMP does not effect or enhance liberation from the membrane of the cAMP-binding protein. Photoaffinity labeling with 8-N3-[125I]cAMP followed by mitochondrial subfractionation and sodium dodecyl sulfate-polyacrylamide gel electrophoresis does not reveal differences in the apparent molecular weight between the membrane-bound and the soluble form of the cAMP receptor. The two forms differ, however, in their partitioning behavior in Triton X-114 as well as in their protease resistance, indicating that the release from the membrane is accompanied by a change in lipophilicity and conformation of the receptor protein. Evidence is presented that a change of the intramitochondrial location of the yeast cAMP-binding protein also occurs in vivo and leads to the activation of a mitochondrial cAMP-dependent protein kinase. The cAMP-binding protein is the first example of a mitochondrial protein with amphitropic character; i.e., it has the property to occur in two different locations, as a membrane-embedded and a soluble form.

Several proteins, previously described as soluble cytoplasmic components, such as α-actinin, gelsolin, profilin, spectrin, vinculin, and protein kinase C have been found in recent years to occur in a membrane-associated form as well [e.g., Niggli and Burger (1987); for a review, see Burn (1988a)]. The reversible interaction of these proteins with the plasma membrane plays an important role in the fulfillment of their function in proliferation, the cell cycle, cell division and motility [for a review, see Geiger (1985) and Burridge (1986)]. It is controlled by the turnover of certain phospholipids in the membrane (Burn & Burger, 1987; Burn, 1988a). For instance, upon hydrolytic generation of diacylglycerol in the plasma membrane the soluble Ca2+- and phosphatidylinerine-dependent protein kinase C becomes associated with the cytosolic side of the plasma membrane, with a concomitant decrease of the Ca2+ requirement for its activation [Kishimoto et al., 1980; for a review, see Nishizuka (1986)].

On the other hand, a variety of enzymes have been described, the greatest portion of which is tightly anchored in the membrane from which they can be released, most commonly, by a chemical modification or hydrolytic cleavage step. In this group are found, e.g., CTP:phosphocholine cytidylyltransferase (Kraft & Anderson, 1983), calmodulin-dependent protein kinase type II (Bennet et al., 1983), and the self-processing protease calpain (Imajoh et al., 1986). These two groups of proteins belong to the newly defined class of amphitropic proteins.

Amphitropism of a protein is not only related to the regulation of its enzymic activity but may also alter its accessibility for other enzymes or substrates (e.g., calpain; Kishimoto et al., 1983) or may induce reorganization of the cytoskeletal architecture during elapse of the cell cycle (e.g., cytoskeletal linker proteins; Lassing & Lindberg, 1985, 1988; Rotman et al., 1982). Thus, amphitropic proteins are assumed to be the heart piece of various regulatory events in transmembrane signaling by association with or dissociation from a membrane, thereby transmitting information from one compartment to the other.

There are only a few amphitropic proteins known which shuttle between the cytoplasm and cellular membranes other than the plasma membrane. CTP:phosphocholine cytidylyltransferase is recovered from both the cytosolic fraction and the endoplasmic reticulum of rat liver homogenates (Hershenson & Ernst-Funberg, 1983), being inactive in the cytosol and active at the endoplasmic reticulum membrane (Terce et al., 1988). A cAMP-dependent protein kinase type II in MDBK cells is associated with centrosomes as well as with...
Golgi stacks (Nigg et al., 1985a), whereas the catalytic subunits, after dissociation from the regulatory subunits, migrate to the nucleus (Nigg et al., 1985b). The Sec4 protein of yeast is a ras-related, GTP-binding protein, synthesized as a soluble cytoplasmic protein. It becomes associated with the cytoplasmic face of both the plasma membrane and secretory vesicles during transit to the cell surface (Goud et al., 1988).

Up to now no amphitropic protein has been described in both the cytoplasm and mitochondria or in two different compartments of this organelle. In addition, little is known about signal transmission from the cytoplasm to mitochondria or within this organelle. Such a transfer of information would be expected to coordinate the biosynthetic activities of the nucleo/cytoplasmic and mitochondrial system both cooperate in the biogenesis of mitochondria (Sechat & Mason, 1974).

Here we describe an amphitropic protein in mitochondria of the yeast Saccharomyces cerevisiae. This protein (apparent molecular weight about 46000) was previously demonstrated to bind cAMP specifically with high affinity (Rödel et al., 1985). It is tightly embedded in the inner membrane, the cAMP-binding domain facing the intermembrane space. In this study we provide evidence that phospholipids or diacylglycerol (or analogues) in the presence of Ca2+, at concentrations which do not affect the integrity of the inner mitochondrial membrane, causes the conversion of the membrane-bound cAMP-binding protein into a soluble form which is then localized in the intermembrane space. The releasing process is accompanied by a conformational change and a significant decrease in lipophilicity and is unlikely to involve proteolytic processing. The soluble form of the cAMP receptor may constitute the regulatory subunit of the cAMP-dependent protein kinase recently identified in the intermembrane space of yeast mitochondria (Müller & Bandlow, 1987a).

**Materials and Methods**

**Materials.** Creatine kinase, cytochrome c, protease inhibitor E-64, α2-macroglobulin, 3',5'-cAMP, 3',5'-cGMP, ADP, AMP, creatine phosphate, MEGA 10, DTT, Triton X-114, PI(3), and IP_2 were obtained from Boehringer, Mannheim: Sephadex G-10 and G-25, Ficoll 400, Percoll, and ethylene glycol were provided by Pharmacia, Freiburg; Zymolyase 20000 was from Seikagaku Kogyo Co., Tokyo; acrylamide, N',N'-methylenebis(acrylamide), ammonium persulfate, TEMED, SDS, POP0, PPO, theophylline, bacitracin, PPH, PHA, TPA, PS, PC, and DAG were obtained from Serva, Heidelberg; [5,8-H]cAMP (52 Ci/mmol) and [γ-32P]ATP (10-25 Ci/mmol) were provided by Amersham-Buchler, Braunschweig; 8-N_{2}PcAMP (20 Ci/mmol) was purchased from ICN, Eschwege; antipain dihydrochlordim, aprotinin, leupeptin hemisulfate, peptatin, PI, IP_2, IP_3, PEG 4000, benzamidine hydrochloride, BSA, HEPES, MES, MOPS, TEA, Tris, and IBMX were from Sigma, Taufkirchen; laetic acid was from Fluka, Neu-Ulm; mixed-ester cellular filters (HAWP, 24-mm diameter) were obtained from Millipore, Eschborn; all other chemicals were bought from Merck, Darmstadt.

**Yeast Strains and Growth Conditions.** The experiments throughout this work were carried out with the protease-deficient yeast strains ABYS-66 (haploid) (Achatz et al., 1984) and PEP4-3 (diploid) (Hemmings et al., 1981) with the wild-type strain D273-10B as indicated. Cells, grown at 30 °C in lactate medium (Daum et al., 1982) in the presence of 0.5 mg/mL adenine, 20 mg/mL uracil, and 250 mg/mL ampicillin, were harvested at a titer of 3 × 10^6/mL.

**Preparation of Mitochondria.** Mitochondria were prepared as published previously (Rödel et al., 1985) with the following modifications: Cells were washed with 1.25 M sorbitol, 20 mM KPi, and 2 mM EDTA, treated with 100 mM Tris/SO_4 and 5 mM DTT in the presence of 1.25 M sorbitol, and converted to spheroplasts with 1.5 mg of Zymolyase 20000/g wet weight (20 min, 30 °C). After having been washed once with 1.25 M sorbitol, the spheroplasts were spun through a 10-mL cushion of 0.8 M sucrose, 1.5% Ficoll, and 20 mM HEPES/KOH (pH 7.2) per gram wet weight (15 min, 4 °C, 3000 rpm; Sorval, HB4 rotor). After removal of the supernatant with sucition, the pellet was resuspended in 30 mL of 0.6 M mannitol, 20 mM Tris-HCl (pH 7.2), and 1 mM EDTA per gram wet weight by ten strokes in a loosely fitting Dounce homogenizer. Crude mitochondria were obtained after five differential centrifugation cycles. They were resuspended at 1 mg of protein/mL in SEM buffer (0.25 M sucrose, 0.5 mM EDTA, 20 mM MOPS/KOH, pH 7.4) and layered on top of a 2.5-mL cushion of 28% Percoll, 0.25 M sucrose, 20 mM MOPS/KOH (pH 7.2), and 0.5 mM EDTA per milligram of protein. After centrifugation (30 min, 4 °C, 18000 rpm; Sorval, SS34 rotor) mitochondria were withdrawn with a Pasteur pipet from the lower third of the gradient and diluted with four volumes of SEM buffer. Purified mitochondria were pelleted (10 min, 4 °C, 15000 rpm; Sorval, SS34 rotor) and resuspended in SEM buffer at a final concentration of 10 mg of protein/mL. All experimental steps following homogenization of the spheroplasts were performed in the presence of a cocktail of protease inhibitors. Final concentrations were 0.25 mM PMSF, 0.1 mg/mL aprotinin, 50 μg/mL peptatin, 10 μg/mL antipain, 1 mM leupeptin, 0.1 mg/mL benzamidine, 0.1 mg/mL soybean trypsin inhibitor, 200 μM E-64, 0.4 mg/mL bacitracin, and 10 μg/mL α2-macroglobulin.

**Subfractionation of Mitochondria.** Purified mitochondria were treated with 20 mM creatine phosphate, 0.5 mg/mL creatine kinase, and 0.5 mM ADP (5 min, 25 °C). Then, up to 15 mg of mitochondria (7.5 mg/mL in SEM buffer) was incubated with 50 μM Ca2+, 0.5 μg/mg TPA, or other compounds as indicated (the phospholipids and diacylglycerol or analogues were dispersed immediately prior to addition by sonication). The releasing reaction was stopped by addition of EGTA (final concentration 100 μM). Mitochondria were washed by centrifugation through a 1-mL cushion of 0.4 M sucrose, 25 mM MOPS/KOH (pH 7.2), and 1 mM EGTA (10 min, 4 °C, 30000 rpm; Beckman TL-100, swing-out rotor). The pellet was resuspended in the same volume of SEM buffer. "Shocked mitochondria" were obtained after dilution of the suspension with five volumes of 20 mM MOPS/KOH (pH 7.2) and gently stirring for 10 min at 4 °C. After five strokes in a loosely fitting Dounce homogenizer, 0.5 mL of shocked mitochondria was layered on top of a two-step sucrose gradient (1 mL of 1.5 M sucrose/1 mL of 0.4 M sucrose in 25 mM HEPES/KOH, pH 7.4, 150 mM KCl, 5 mM DTT, 5 mM MgCl_2, 0.5 mM EDTA, 0.5 mg/mL BSA) and centrifuged (15 min, 4 °C, 50000 rpm; Beckman TL-100, swing-out rotor). The upper layer on top of the sucrose (500 μL) (intermembrane space) and the interface between the sucrose layers (200 μL) (mitoplasts) were removed with a Pasteur pipet and diluted with an equal volume of assay buffer (25 mM HEPES/KOH, pH 7.4, 125 mM KCl, 0.25 mM sucrose, 0.5 mM 5'-AMP, 0.1 mM EDTA, 7.5 mM MgCl_2, 15% glycerol, 0.5 mM DTT, 100 μM PMSF). The supernatant fraction was precipitated by addition of an equal volume of 60% PEG 4000 in 20 mM MOPS/KOH (pH 7.2) and 1 mM EDTA (15 min, 0 °C/10 min, 4 °C, 10000g) and subsequently dissolved in binding buffer (25 mM MOPS/KOH, pH 7.2, 100 mM sodium citrate, 5 mM DTT, 5 mM MgCl_2, 100 mM sucrose,
Amphitropic cAMP-Binding Protein

Biochemistry, Vol. 28, No. 26, 1989 9959

7.5% ethylene glycol, 1 mg/mL BSA, 2 mM theophylline, 1 mM IBMX) at 2.5 mg of protein/mL. The pellet fraction was layered on top of 0.5 mL of 0.4 M sucrose in 25 mM HEPES/KOH, pH 7.4, and centrifuged (10 min, 4 °C, 25000 rpm; Beckman TL-100, fixed-angle rotor). The pellet was resuspended in binding buffer at 2.5 mg/mL protein.

**cAMP-Binding Assay.** Specific binding of cAMP to mitochondria or mitochondrial subfractions was determined by a filter-binding assay (Walton & Garren, 1970). A total of 250 µg of mitochondrial protein in a maximum volume of 125 µL of binding buffer was supplemented with 5% MEGA 10 (final concentration 0.5%) and incubated (5 min, 10 °C). After centrifugation (15 min, 4 °C, 80 000 rpm; Beckman TL-100, fixed-angle rotor) the supernatant was removed, diluted with 50 µL of 25 mM MOPS/KOH (pH 7.2), 0.5 mM EDTA, 25 mM benzoaminide, 0.1 mM 5'-AMP, 25 mM DTT, 5 mM/mL BSA, 0.5 mg/mL aprotinin, 5 mg/mL antipain, 5 mM leupeptin, 2.5 mg/mL pepstatin, and 5 mM IBMX and 15 µL of [3H]cAMP (50 nCi), and adjusted with H2O to a total volume of 250 µL. After incubation (15 min, 0 °C), 750 µL of ice-cold washing buffer (20 mM MOPS/KOH, pH 7.2, 100 mM sodium citrate, 1 mM EDTA, 0.5 mM EGTA, 0.2 mM S'-AMP, 0.5 mM MgCl2, 5 mM DTT, 200 µM PMSF, 1 mM theophylline, 2 mM IBMX, 0.1% MEGA 10, 10% glycerol) was added and the mixture was applied to a cellulose column of Sephadex G-10 (equilibrated with labeling buffer). After incubation (15 min, 0 °C), the reaction was terminated by addition of ATP (final concentration 4 mM), cooling to 0 °C, and incubation with 0.5% mercaptoethanol, 50 µM unlabeled cAMP and 50 µM unlabeled cAMP were added, and the mixtures were centrifuged through a 1-mL column of Sephadex G-10 (equilibrated with labeling buffer). To the eluate (300 µL), an equal volume of 10% TCA was added (30 min, 0 °C). After centrifugation (15 min, 4 °C, microfuge) the pellet was washed twice with 1 mL of ice-cold acetone and once with 0.5 mL of 70% ethanol, dried under vacuum, resuspended in 50 µL of sample buffer (Laemmli, 1970), and analyzed by SDS–PAGE.

**Hydrophobic/Hydrophilic Partitioning.** The sample was supplemented with an equal volume of 2% Triton X-114 (in 0.3 M NaCl and 0.1 M sorbitol/TEA), incubated (30 min, 0 °C), and centrifuged (2 min, 0 °C, microfuge) (Bordier, 1981). The clear supernatant was warmed up for 2 min at 4 °C and then for 10 min at 37 °C and spun for 1 min in a microfuge at room temperature to separate the detergent and aqueous phases.

**Analytical Procedures.** Published procedures were used for the determination of the following enzyme activities: succinate:cytochrome c oxidoreductase (Lang et al., 1974), fumarase (Racker, 1950), and cytochrome c peroxidase (Dia- vadi-Ohanian et al., 1978). Cytochrome c was assayed by immunodetection with specific antibodies and ³⁵Clabeled protein A (Towbin, 1979; Müller & Bandlow, 1987a) and densitometry of autoradiograms with an LKB Ultrascan XL laser densitometer. Protein was determined by the method of Bradford (1976). Polyaclrylamide gel electrophoresis [5% acrylamide (w/v) stacking gels and 12% or 20% acrylamide (w/v) separating gels] was carried out in the presence of SDs and urea (Laemmli, 1970; Ito et al., 1980; Müller & Zimmermann, 1987). For fluorography [³²H-containing gels were soaked in sodium salicylate (Chamberlain, 1979), dried under vacuum, and exposed to X-ray films (Kodak X-Omat AR) at 12-72 h at –80 °C.

**RESULTS**

The mitochondrial cAMP receptor (molecular weight 46 000) which binds cAMP at 0.5–1 pmol/mg of protein with high affinity (Kd = 10⁻⁹ M⁻¹) was found to be located in the inner mitochondrial membrane of the yeast Saccharomyces cerevisiae. The cAMP-binding domain of the receptor faces the intermembrane space (Rödel et al., 1985). cAMP-dependent protein kinase activity was, on the other hand, observed exclusively in the soluble intermembrane space compartment of mitochondria (Müller & Bandlow, 1987a). In order to resolve this discrepancy in the occurrence of cAMP-binding and of cAMP-mediated effects in different intramitochondrial compartments, we studied whether, under certain conditions, the membrane-bound mitochondrial cAMP-binding protein is converted into a soluble form as has been described for some eucaryotic plasma membrane associated proteins.

**Calcium, Phospholipids, and Diacylglycerol (or Analogues), but Not cAMP, Cause a Specific Change in the Intramitochondrial Location of cAMP-Binding Activity.** We examined the effects of calcium and various lipids or lipid derivatives on the intramitochondrial distribution of cAMP-binding activity, since it has been reported repeatedly that such compounds affect the intracellular distribution of certain amphitropic proteins. Yeast mitochondria were purified by Percoll gradient centrifugation, which ensures the removal of contaminating cAMP-binding proteins from both the plasma membrane (Jaynes et al., 1980) and the cytoplasm (Sy &
The purity of mitochondria was tested routinely by marker enzyme assays (Rödel et al., 1985; Müller & Bandlow, 1987a). Typically, isolated mitochondria contained only 3–5% of total cellular hexokinase activity (a cytoplasmic enzyme), 6–8% of total cellular catalase activity (a peroxisomal enzyme), and 10–15% of total membrane-associated alkaline phosphatase or adenylate cyclase activity (enzymes of the plasma membrane) but 75–85% of total membrane-associated cyclic AMP-binding activity. The specific cyclic AMP-binding activity was about 7-fold higher in mitochondria isolated from Percoll gradients than in purified plasma membrane preparations [see Müller and Bandlow (1989b)]. These fractionation data confirm the intracellular location of the mitochondrial cyclic AMP receptor protein, which has been demonstrated previously (Rödel et al., 1985). For studying the intramitochondrial location of the binding activity, mitochondria were incubated with the compounds to be tested and subsequently converted to “shocked mitochondria” (mitochondria having disrupted outer membranes). These were separated by centrifugation through a two-step sucrose gradient into mitoplasts (inner-membrane vesicles with adhering outer membranes) and the released soluble intermembrane space proteins. This procedure guarantees the integrity of the mitoplasts as sealed vesicles and their purification from soluble proteins other than those of the mitochondrial matrix. The two mitochondrial subfractions were assayed for cyclic AMP-binding activity by assaying protein-bound \(^{3}H\)cAMP retained on nitrocellulose filters.

It can be seen from Figure 1 that, in the absence of Ca\(^{2+}\), none of the phospholipids was really efficient in altering the intramitochondrial distribution of the specific cyclic AMP-binding activity (open bars). However, as soon as 50 \(\mu\)M Ca\(^{2+}\) was also present, a 3–4-fold enrichment of cyclic AMP-binding activity in the intermembrane space was observed (solid bars). Only small differences in the capacity to stimulate specific cyclic AMP-binding activity in the intermembrane space could be detected between the five phospholipids tested (PC > CL > PS > PI > PIP\(_{2}\)). On the other hand, Ca\(^{2+}\) alone led only to a 2-fold increase of specific cyclic AMP-binding activity in that compartment (H\(_{2}\)O). Of the hydrolysis products of phospholipids (DAG, IP\(_{3}\), P) only diacylglycerol and the diacylglycerol analogue TPA exhibited a Ca\(^{2+}\)-dependent increase of cyclic AMP binding in the intermembrane space, the latter being the most effective of all compounds. Physiologically inactive TPA derivatives (phorbol, phorbol acetate) showed no Ca\(^{2+}\)-dependent effect in addition to that observed with Ca\(^{2+}\) alone. These findings underline the specificity of TPA in mimicking the phospholipid-induced change in topology of the cyclic AMP-binding activity. Therefore, TPA in the presence or absence of Ca\(^{2+}\) was used in the following experiments. Interestingly, cyclic AMP alone did not affect the intramitochondrial distribution of cyclic AMP-binding activity either alone or in combination with Ca\(^{2+}\) and/or TPA.

The necessity of incubating isolated yeast mitochondria with phospholipids or TPA to yield maximal soluble cyclic AMP-binding activity led to the question whether this could be due to solubilization of the inner mitochondrial membrane. To test this possibility, we assayed the intermembrane space and mitoplast fractions from isolated mitochondria, incubated with or without Ca\(^{2+}\) and/or TPA, for marker proteins of the matrix (fumarase), inner membrane (sucinate-cytochrome c oxidoreductase, cytochrome \(c_{1}\)), and intermembrane space (cytochrome \(c\) peroxidase). The DeDuve plots in Figure 2 show that the membrane structure remained undisturbed by the treatment with Ca\(^{2+}\) and TPA at the concentrations used. The bulk of both matrix and inner-membrane marker proteins as well as of cytochrome c peroxidase activity copurified with the mitoplast and intermembrane space fractions, respectively, regardless of whether Ca\(^{2+}\) and TPA were present or not. By contrast, the relative specific and total cyclic AMP-binding activities increased in the intermembrane space of mitochondria incubated with Ca\(^{2+}\) and—in a more dramatic fashion—in the presence of Ca\(^{2+}\) and TPA. This increase in the soluble fraction was accompanied by a corresponding decrease in total binding activity in the inner membrane. This inverse relationship of cyclic AMP-binding activity in both compartments provides the first evidence that the membrane-bound cyclic AMP-binding protein is released into the intermembrane space. The total recovery of cyclic AMP-binding activity in mitoplasts and intermembrane space after Ca\(^{2+}\)/TPA treatment of mitochondria was only about 60%. This loss is indicative of either incomplete recovery after the incubation and subfractionation or
Amphitropic cAMP-Binding Protein

Biochemistry, Vol. 28, No. 26, 1989 9961

FIGURE 2: Distribution of mitochondrial marker enzymes. The experiment was carried out as described for Figure 1, but in addition to the cAMP-binding activity, the activities of fumarase, cytochrome c peroxidase, and succinate-cytochrome c oxidoreductase were determined. The activities of only two marker enzymes and CAMP-binding protein in the mitoplast or intermembrane space fraction, respectively. CAMP-binding activities originate from the same protein. In untreated mitochondria, the areas of the filled rectangles represent the total activities of the marker enzymes and cAMP-binding protein in the mitoplast or intermembrane space fraction, respectively.

or a reduced binding affinity ($K_a$) or a low stability of the cAMP receptor in the presence of Ca$^{2+}$ and TPA (see below).

Membrane-Bound and Released cAMP-Binding Protein Exhibit the Same Apparent Molecular Weight. Next we wanted to prove that the soluble and membrane-bound cAMP-binding activities originate from the same protein. In the first experiment, we compared the molecular weights of both cAMP-binding proteins. Isolated mitochondria were incubated with various compounds and subsequently separated into mitoplast and intermembrane space fractions, both of which were photoaffinity labeled with 8-N3-[32P]cAMP and analyzed by SDS-PAGE and fluorography. The data shown in Figure 3A are in good agreement with those from the filter-binding assay (Figure 1). In untreated mitochondria (H$_2$O), a single photolabeled protein of $M_c$ 46,000 was observed which was exclusively associated with the mitoplast fraction. No such signal could be detected at higher molecular weight in particular $M_c$ 52,000 (Hixson & Krebs, 1980), which documents the absence of cross contamination by the cytoplasmic R subunit in these mitochondrial preparations. In these untreated mitochondria no photoaffinity labeled material could be detected in the intermembrane space (lane 3). In the presence of Ca$^{2+}$ alone, only a small portion of cAMP-binding protein was identified by photoaffinity labeling intermembrane space proteins (lane 6), whereas Ca$^{2+}$ and TPA together led to significant labeling in this compartment (lane 15) with a concomitant complete loss of label in the mitoplasts (lane 14). The specificity of the combination Ca$^{2+}$/TPA in releasing the cAMP receptor was again emphasized by the lack of effects by TPA alone or phorbol (lanes 9, 12, and 18). The apparent molecular weights of the cAMP-binding proteins recovered from the two different compartments did not differ. [The electrophoretic system would have resolved molecular weight differences of only about 0.5K in proteins of about 40K (Zimmermann & Mollay, 1986).]

In the second experiment, we studied the molecular relationship between the two cAMP-binding proteins. Whole mitochondria were labeled with 8-N3-[32P]cAMP in the presence or absence of excess unlabeled cAMP and subsequently treated with or without Ca$^{2+}$ and TPA. After the mitochondria had been fractionated into mitoplasts and intermembrane space proteins, the samples were analyzed by SDS-PAGE (Figure 3B). The mitochondrial cAMP receptor from both compartments bound CAMP specifically as evident from the complete competition of labeling with azido-cAMP by an excess of unlabeled cAMP (lanes 4-6, 10-12, and 16-18). In the presence of Ca$^{2+}$ alone, most of the photoaffinity labeled protein was recovered from the mitoplast fraction (lane 8). Only a minor portion could be observed in the intermembrane space (lane 9). The latter compartment was, however, completely devoid of cAMP receptor in the absence of Ca$^{2+}$ (lane 3). Ca$^{2+}$ in the presence of TPA caused a dramatic increase of photoaffinity labeled material in the intermembrane space (lane 15) with a concomitant decrease in the mitoplasts (lane 14). Photoaffinity labeling prior to incubation with Ca$^{2+}$/TPA led to some additional labeled bands, caused probably by proteolytic degradation of the 46K cAMP-binding protein despite the presence of a multitude of protease inhibitors. This proteolytic sensitivity would also explain the loss of total CAMP-binding activity in mitochondria treated with Ca$^{2+}$/TPA (shown above). Interestingly, the degradation patterns of the labeled fragments were identical in both compartments, inferring a common origin. Again, a signal which could be due to a contaminating cytoplasmic cAMP-binding protein was absent. These findings together with the identical apparent molecular weight (46K) of the cAMP receptor in both mitochondrial compartments suggest that, in the presence of Ca$^{2+}$/TPA and—to a lesser extent—of Ca$^{2+}$ alone, the cAMP-binding protein is released from the inner membrane into the intermembrane space. Thus, the
mitochondrial cAMP receptor of the yeast *S. cerevisiae* exhibits properties of an amphitropic protein.

The conversion of the membrane-bound form into the soluble form of the cAMP-binding protein under preservation of the apparent molecular weight could be most easily explained by a loose association of the receptor with the outer face of the inner membrane as a peripheral membrane protein. To test the nature of the membrane association, isolated mitochondria were photoaffinity labeled, subjected to various extraction procedures, and centrifuged. The resulting pellets and supernatant fractions were counted for radioactivity. Figure 4 shows that the cAMP-binding protein was almost completely released into the supernatant in the presence of detergent. However, extraction by agents capable of releasing peripheral membrane proteins (urea, hydroxylamine, sodium carbonate, or high salt) was ineffective. In a control incubation with Ca²⁺/TPA, the cAMP receptor was solubilized to an extent similar to that after detergent treatment. These findings indicate that the membrane-bound cAMP receptor behaves as an integral rather than as a peripheral membrane protein. Since most integral membrane proteins are anchored in the membrane via a stretch of hydrophobic amino acids and since, at the same time, no proteolytic removal of such a structure seemed to have had occurred, we examined how the membrane-embedded form of the amphitropic cAMP-binding protein is rendered soluble.

**Two Versions of the cAMP-Binding Protein Differ in Their Conformation and Detergent-Binding Characteristics.** The membrane-bound and soluble forms of amphitropic proteins are assumed to differ in their structure due to posttranslational modification and/or in their conformation. We tested the last possibility and examined the relative susceptibility of the released and membrane-associated receptor toward proteolytic digestion by endoprotease V8. Different conformations of a protein should exhibit different protease sensitivities. Photoaffinity labeled released or membrane-bound receptor was dissolved in buffer plus detergent and digested with limiting amounts of protease V8 in both the native (minus SDS/urea) and the denatured state (plus SDS/urea). The electrophoretic analysis of the labeled degradation products is shown in Figure 5. In the absence of SDS and urea the peptide patterns obtained with up to 10 µg/mL protease differed significantly between the membrane-bound form solubilized by detergent (lanes 1–3) and the released form (lanes 6–8), suggesting different conformational states of these proteins. The identity of the peptide patterns generated after denaturation of both forms with SDS/urea supports this view and again emphasizes the similarity or perhaps identity of their primary structures (lanes 11–16). The fact that, after complete digestion of the denatured 46K protein, two peptide fragments were observed in about stoichiometric amounts which have retained the photoaffinity label (lanes 13 and 16) may be taken as an
Amphitropic cAMP-Binding Protein

Mitochondria were isolated, incubated with or without Ca\(^{2+}\) plus TPA, and fractionated into mitoplasts containing membrane-bound receptor and intermembrane space containing soluble receptor. Both forms were incubated at 0 °C in a detergent-containing buffer and subsequently assayed for cAMP-binding activity. In all three strains the soluble receptor had a shorter half-life than the membrane-bound one, the protein from the wild-type strain being most sensitive (data not shown). This argues that the released receptor has a less compactly folded structure which is more accessible to protease digestion as compared to the solubilized membrane protein. Obviously, liberation of the receptor from the membrane by detergent does not per se result in the conformation of the Ca\(^{2+}\)/TPA-released receptor. This is the first hint at a posttranslational modification causing the conformational change (Müller & Bandlow, 1989b).

An altered conformation may guarantee the compatibility of the previously membrane-embedded cAMP receptor with the soluble compartment. Therefore, we studied the hydrophobicity of both amphitropic versions by examining their partition equilibrium between an aqueous phase and TX-114. Photoaffinity labeled mitochondria were incubated with phospholipids or lipid derivatives in the presence or absence of Ca\(^{2+}\). Subsequently, the separated mitoplast and intermembrane space fractions were adjusted to 1% TX-114 and centrifuged. Both pellet and supernatant were assayed for radioactivity. It can be seen from Figure 6 that the membrane-bound cAMP-binding protein (H\(_2\)O) was found almost exclusively in the detergent phase, whereas the bulk of the released receptor (Ca\(^{2+}\) plus phospholipids, TPA, diacetyl-glycerol) was recovered from the aqueous phase. Surprisingly, Ca\(^{2+}\) alone, although solubilizing about 20% of the total cAMP-binding activity, did not increase the proportion of receptor in the aqueous phase. These results argue that the Ca\(^{2+}\) plus phospholipid-controlled release of the cAMP-binding protein from the inner membrane is accompanied by a conformational change and decrease in lipophilicity, whereas Ca\(^{2+}\) alone is inefficient in this respect.

![FIGURE 4: Extraction behavior of the cAMP-binding protein. Whole mitochondria (500 µg) were labeled with 8-N\(_3\){\[^{32}P\]}cAMP, converted to shocked mitochondria, and divided into ten aliquots. Two aliquots were incubated with Ca\(^{2+}\) in the presence or absence of TPA. All the other aliquots were centrifuged through 200-µL cushions of 0.25 M sucrose. The resulting pellets were resuspended in either 0.1 M sorbitol/20 mM TEA (pH 7.0) (control) or 6 M urea, 1 M KCl, 1 M NaCl, 1% TX-100, and 0.5% MEGA 10 made in 0.1 M sorbitol/TEA or 0.2 M sodium carbonate (in water, pH 11.5) or in 1 M hydroxylamine (in water, pH 7.0). After incubation for 15 min on ice or 60 min at 37 °C (hydroxylamine) or 10 min at 25 °C (Ca\(^{2+}\)/TPA), the samples were spun (60 min, 4 °C, 25000g). The supernatants and pellets were boiled in 0.5% SDS, supplemented with scintillation cocktail, and counted for radioactivity. 100% of photoaffinity label refers to the cpm measured for 50 µg of photoaffinity labeled total mitochondrial protein before the extraction procedure.](https://example.com/f4)

![FIGURE 5: Partial proteolysis of membrane-bound and soluble cAMP-binding protein. Aliquots of photoaffinity labeled mitochondria (3.2 mg) from the yeast strain PEP4-3 were incubated with H\(_2\)O or Ca\(^{2+}\) plus TPA. Reisolated mitochondria were separated into mitoplast and intermembrane space fractions. The mitoplasts derived from the untreated mitochondria were diluted 4-fold with 25 mM MOPS/KOH (pH 7.2), collected by centrifugation, and resuspended at 0.5 mg of protein/ml in 50 mM (NH\(_4\))\(_2\)CO\(_3\) buffer (pH 7.8) plus 0.5% MEGA 10 (m). The intermembrane space proteins derived from the Ca\(^{2+}\)/TPA-treated mitochondria were concentrated 10-fold by ultrafiltration through an Amicon XM-300 membrane and supplemented with (NH\(_4\))\(_2\)CO\(_3\). All samples were divided into two halves, one of which received urea (6 M final concentration) and SDS (0.2% final concentration) and the other H\(_2\)O. Aliquots of each half were incubated with 2, 5, 10, 15, or 20 µg/mL protease V8 from Staphylococcus aureus (60 min, 25 °C) in a total volume of 250 µL. After addition of a 2-fold molar excess of α-macroglobulin (5 min, 0 °C), the samples were precipitated with 5% TCA, washed twice with acetone, and analyzed by SDS-PAGE (20%) and fluorography.](https://example.com/f5)
Protein Changes Also In Vivo. Yeast cells, grown at 30 °C to midlog phase, were cooled to 10 °C. After various incubation periods mitochondria were prepared and separated into mitoplasts and intermembrane space, and both fractions were assayed for CAMP-binding activity and specific marker proteins. In addition, the partitioning behavior of photoaffinity labeled CAMP-binding protein was analyzed. Figure 7 demonstrates that the intramitochondrial distribution of the CAMP receptor, in fact, changed during prolonged incubation of cells at 10 °C. After 5 h the protein was nearly equally distributed between mitoplasts and intermembrane space as well as between the detergent and aqueous phases, respectively. By contrast, after growth at 30 °C, the bulk of the receptor was membrane bound. Inherent instability of mitochondria isolated from nongrowing yeast cells was excluded since the major portion of fumarase and succinate-cytochrome c oxidoreductase was retained within the mitoplasts for more than 6 h after the temperature shift (data not shown). It remains unclear whether the altered intramitochondrial distribution of the CAMP receptor is ultimately related to a change in lipid composition of the mitochondrial membranes caused by the temperature shift. Nevertheless, these data suggest that the amphitropism of the CAMP receptor observed in vitro may also play a role in vivo.

Release of the CAMP-Binding Protein Is Accompanied by an Increase in Mitochondrial CAMP-Dependent Protein Kinase Activity. Having established the dual intramitochondrial location of the mitochondrial CAMP-binding protein in vitro and in vivo, the question concerning the functional relevance of this amphitropism arose. In a previous study we observed a CAMP-dependent protein kinase activity in yeast mitochondria located exclusively in the intermembrane space (Müller & Bandlow, 1987a). However, CAMP-dependent phosphorylation of a 40K inner-membrane protein could be demonstrated only after incubation of an excess of isolated intermembrane space proteins with isolated inner-membrane vesicles but not in whole mitochondria. In light of the data presented in this paper it was tempting to speculate that, under physiological conditions, mitochondria would contain only a low steady-state concentration of CAMP receptor in the intermembrane space, only in this compartment it might function as the regulatory subunit of the CAMP-dependent kinase. Consequently, the membrane-bound form of the receptor would be a kind of inactive precursor. To test this possibility we studied the CAMP-dependent protein kinase activity in intact yeast mitochondria in the presence of Ca²⁺ and TPA.
Amphitropic cAMP-Binding Protein

Mitochondria were incubated with Ca²⁺ in the presence or absence of TPA, reisolated by centrifugation, and subsequently incubated with [γ-³²P]ATP in the presence or absence of cAMP. The total incubation mixtures or the separated soluble and particulate fractions were analyzed by SDS-PAGE and autoradiography. Figure 8 shows the complex pattern of phosphoproteins obtained. Surprisingly, we observed a dramatic decrease in total phosphate incorporation in the presence of 20 nM cAMP whether or not the mitochondria had been pretreated with TPA (lanes 3 and 6). Most of the ³²P-labeled proteins seen were likely to be phosphorylated by extra- and intramitochondrial protein kinases other than the cAMP-dependent species. At least four such enzymes had been described in mitochondria previously, among them a casein kinase II (Müller & Bandlow, 1987a). In addition, the cAMP-dependent phosphorylation of a 50K protein and a 59K protein could be detected. It was stimulated 2-3-fold by the simultaneous presence of Ca²⁺ and TPA (compare the stimulation factors between lanes 3 and 4 vs those between lanes 5 and 6). Both target proteins were membrane associated (lane 1). The 40K phosphoprotein is presumably identical with the inner-membrane protein found to be phosphorylated in a cAMP-dependent fashion in the reconstituted system (Müller & Bandlow, 1987a). The Ca²⁺/TPA-induced stimulation of the cAMP-dependent mitochondrial kinase activity—which not very pronounced—provides the first hint at a new type of kinase activation via regulation of the amphitropic of its regulatory subunit.

The occurrence of the mitochondrial cAMP-binding protein in a complex with a putative catalytic subunit of the mitochondrial cAMP-dependent protein kinase was tested by analyzing its migration behavior in the membrane-bound and soluble states on a native gel. For this purpose mitochondria were incubated with Ca²⁺ and [γ-³²P]ATP since it had been shown that the cAMP receptor is phosphorylated in a Ca²⁺-dependent fashion (Müller & Bandlow, 1989b). After incubation with or without TPA and cAMP, the receptor protein was solubilized with Triton X-100, purified by cAMP affinity chromatography, and analyzed by gel electrophoresis in the native or denatured state.

Figure 9 reveals that, in the absence of cAMP, the membrane-bound form of the ³²P-labeled cAMP receptor, solubilized by detergent, as well as the released form migrated with two different electrophoretic mobilities on the native gel (TX-100) (lanes 1 and 3). However, in the presence of cAMP the upper minor band at the boundary between the stacking and running gel disappeared in the soluble version (lane 4) by contrast to the membrane-bound one (lane 2). This labeled material presumably represented a high molecular weight complex between the cAMP-binding regulatory subunit and a catalytic subunit. Comigration of both versions of the receptor protein on the denaturing gel (SDS), irrespective of whether cAMP had been added (lanes 2 and 4) or not (lanes 1 and 3), provided further evidence for complex formation. Obviously, a portion of this complex disassembled also in the presence of Triton X-100 alone, since some phosphorylated receptor migrated into the running gel, presumably as monomer (lower major band), also in the absence of cAMP (lanes 1 and 3) or TPA (lane 2). Nevertheless, this experiment suggests that the mitochondrial cAMP receptor interacts noncovalently with another protein, most likely the catalytic subunit of the mitochondrial cAMP-dependent protein kinase. Release of the complex from the inner membrane seems to be a prerequisite for the subsequent cAMP-dependent dissociation into its subunits and activation of the kinase.

DISCUSSION

Three different locations have been described in the literature for cAMP-binding proteins in yeast: a soluble R subunit of a soluble protein kinase A was found in the cytoplasm of
this organism (Hixson & Krebs, 1980), several such proteins were described to be associated with the plasma membrane (Jaynes et al., 1980), and one species was detected in mitochondria (Rödel et al., 1985). Apart from location, these proteins differ in molecular weight: 52K was reported for the cytosolic R subunit, 46K for the mitochondrial version, and 25K, 34K, 46K, and 58K for the species from the plasma membrane. In agreement with this latter report, we observed cAMP binding in plasma membrane preparations which accounts for a minor portion of the total membrane-associated cAMP-binding activity in yeast cells (Rödel et al., 1985). But, in contrast to the earlier observations (Jaynes et al., 1980), we could detect only a single species of M, 44 000 in yeast plasma membranes. Its lower molecular weight and its failure to be released from the membrane by treatment with Ca2+/TPA in vitro, the biosynthetic incorporation of glutamine, and its inability to become metabolically labeled by stearic acid in vivo (Müller & Bandlow, 1989a; Bandlow & Müller, in preparation) argue that this species is distinct from the cAMP-binding protein described here both in structure and in their type of membrane anchor. Accordingly, the low cross contamination by plasma membranes of the mitochondrial preparations (about <15%) does not interfere with the observations reported here. In addition, we verified that the soluble cytoplasmic R subunit (52K) was absent from our mitochondrial preparations. In summary, we detected three separate cAMP-binding proteins in yeast, one of which has been shown to occur in mitochondria (Rödel et al., 1985; Müller & Bandlow, 1987a). Their genetic relationship remains to be elucidated.

A large body of experimental data documents the amphiprotic character of the mitochondrial cAMP receptor of yeast. First, in the presence of calcium together with certain phospholipids or lipid derivatives the cAMP-binding activity increases dramatically in the intermembrane space of isolated mitochondria with a concomitant decrease in the inner membrane. Second, under the same conditions photolabile labeled receptor disappears from the inner membrane and emerges in the intermembrane space. Third, according to their electrophoretic mobility both binding proteins have the same apparent molecular weight and share identical electrophoretic patterns of azido-cAMP-labeled peptides after partial proteolysis of the denatured protein. This suggests that the membrane-bound and soluble mitochondrial cAMP receptors are two forms of the same protein with no or only very minor differences in their primary structures. However, both versions exhibit different conformations and possibly posttranslational modifications, as was indicated by two independent criteria: accessibility for proteases of the native proteins and lipophilicity. The structural basis for variation of these properties is elucidated in an accompanying paper (Müller & Bandlow, 1989a). Fourth, movement of the cAMP-binding protein from the inner membrane into the intermembrane space is also observed in vivo. Thus, the described cAMP receptor provides the first example of an amphiprotic protein in mitochondria.

We observed an amazing lack of specificity of phospholipids effecting the release of the membrane-embedded protein. Since lipid head groups alone, e.g., IP3, have no effect, whereas diacylglycerol and especially its nonhydrolyzable analogue TPA, are highly active, the capability of the activator molecule to interact spontaneously with the lipid bilayer may be critical. The importance of hydrophobicity is supported by the fact that phorbol or phorbol acetate, TPA derivatives lacking the myristic acid ester, is incompetent in releasing the receptor, thus emphasizing the necessity of fatty acid side chains. However, it remains to be clarified which phospholipid or lipid derivative acts in vivo and, in addition, from which cellular membrane it is derived.

Lipid compounds are not an absolute prerequisite for the releasing process. They drastically increase its efficiency, provided Ca2+ is present at concentrations approximately found in the cytoplasms of eukaryotic organisms (McCormack & Denton, 1986). On the other hand, calcium alone converted only a limited portion of the membrane-bound cAMP receptor into a soluble form. Surprisingly, it was observed that this form of released binding protein does not exhibit decreased lipophilicity. It is unknown whether the Ca2+-, regulating the amphitropism in vivo, originates from the endoplasmic reticulum or the mitochondrial matrix, compartments considered as intracellular calcium stores (Wang & Waisman, 1979). In an accompanying paper (Müller & Bandlow, 1989b) we studied whether the Ca2+-induced and the Ca2+/phospholipid-induced releases are two mechanistically different processes or whether there exists only a unique releasing pathway wherein Ca2+ and phospholipids affect successive steps.

Eukaryotic cAMP-binding proteins are thought to act exclusively as regulatory subunits of cAMP-dependent protein kinases (Kuo & Greengard, 1969; Greengard, 1978; Cohen, 1982). This assumption is strengthened by the identification of a unique mitochondrial cAMP receptor and mitochondrial cAMP-dependent protein kinase activity in yeast (Müller & Bandlow, 1987a). In detail, it was reported that almost all of the mitochondrial cAMP-binding protein was tightly bound to the inner membrane, whereas cAMP-dependent protein kinase was observed exclusively in the soluble intermembrane space, phosphorylating a 40K inner-membrane protein. This indicates the simultaneous presence of R and C subunits in this compartment. The findings reported here nicely explain the former discrepancy in that both forms of cAMP-binding activity are derived from the same protein, but only the soluble version behaves as an R subunit of a protein kinase A. This implies that the C subunit is bound to R also in the membrane-embedded state of R and is dissociable by cAMP only upon release from the membrane of the regulatory subunit. Consequently, the cAMP-dependent kinase would be active only in the soluble state.

As shown in this study, the stimulation of the mitochondrial cAMP-dependent kinase by cAMP is moderately enhanced in the presence of Ca2+ plus TPA. This Ca2+/TPA-induced kinase activation may be masked by rapid dephosphorylation as part of a downregulation of physiological short-term stimulations. Dissociation of C from the Ca2+/TPA-released but not from the membrane-embedded or detergent-solubilized R subunit, coisolation of R and C by affinity chromatography after solubilization by mild detergents, and the occurrence of casein kinase activity in the eluate (W. Bandlow, unpublished observations) supply additional evidence for the assumption that R is associated with C also in the membrane-bound state. The membrane-bound and the soluble forms of the R subunit were shown to bind cAMP. Obviously, the altered conformation (and lipophilicity) of the released R is a prerequisite for its cAMP-dependent dissociation from C.

The physiological role of the mitochondrial cAMP-dependent protein kinase activity is not yet known. It has, however, repeatedly been communicated that cAMP has some bearing on yeast mitochondrial biogenesis (Chandrasekaran & Jayaraman, 1978; Mahler & Lin, 1978) and respiration (Fang & Butow, 1970). Recently it was noticed that ras1, ras2 double mutants, which have very low endogenous cAMP (Temeles et al., 1985), exhibit a respiratory-deficient phe-
notype (Fraenkel, 1985; Tatchell et al., 1985) whereas overproduction of the cyclic nucleotide in RASm'19 mutants results in giant mitochondria (J. Mattoon, personal communication) capable of enhanced respiration (Kataoka et al., 1984; Toda et al., 1985). So, it may be speculated that mitochondrial protein kinase A is involved in the modulation of cAMP-dependent effects on mitochondria. It might fulfill an essential role like all second messenger-linked kinases found so far [for a review, see Krebs and Beavo (1979) and Pall (1981)] by somehow controlling the biogenesis of mitochondria as a component of a complex signal transduction pathway between the cytoplasm and mitochondria. The direct control by this kinase of mitochondrial transcription, translation, or import of proteins into mitochondria is, however, unlikely (Müller & Bandlow, 1987b).

Registry No. IP, 85166-31-0; PHA, 70470-59-6; 12-myristoyl-PHA, 16561-29-8; PS, 40290-42-4; PC, 63-89-8; DAG, 30334-71-5; Ca, 7440-70-2; protein kinase, 9026-43-1.

References