

1 **Monocyte-chemoattractant protein-1 Levels in Human Atherosclerotic**
2 **Lesions Associate with Plaque Vulnerability**

3 **Running title:** *Georgakis et al.; MCP-1 in human plaque vulnerability*

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1 **Abstract**

2 **Objective:** To determine whether MCP-1 levels in human atherosclerotic plaques associate
3 with plaque vulnerability features.

4 **Approach and Results:** We measured MCP-1 levels in human atherosclerotic plaque
5 samples from 1,199 patients in the Athero-EXPRESS Biobank who underwent endarterectomy
6 for treatment of carotid stenosis. We explored associations with histopathological and
7 molecular features of plaque vulnerability, clinical plaque manifestations, and vascular events
8 up to 3 years after endarterectomy. Following adjustments for age, sex, and vascular risk
9 factors, MCP-1 plaque levels were associated with histopathological markers of plaque
10 vulnerability (large lipid core, low collagen content, high macrophage burden, low smooth
11 muscle cell burden, intraplaque hemorrhage) and with a composite vulnerability index (range
12 0-5, beta per SD-increment in MCP-1: 0.42, 95%CI: 0.30-0.53, $p=5.4 \times 10^{-13}$). We further found
13 significant associations with higher plaque levels of other chemokines and pro-inflammatory
14 molecules, and markers of neovascularization and matrix turnover. When exploring clinical
15 plaque instability, MCP-1 plaque levels were higher among individuals with symptomatic
16 plaques as compared to those with asymptomatic plaques (OR per SD-increment in MCP-1:
17 1.36, 95%CI: 1.09-1.69). MCP-1 levels were further associated with a higher risk of
18 periprocedural major adverse vascular events and strokes occurring in the first 30 days after
19 plaque removal.

20 **Conclusions:** Higher MCP-1 plaque levels are associated with histopathological, molecular,
21 and clinical hallmarks of plaque vulnerability in individuals undergoing carotid endarterectomy.
22 Our findings highlight a role of MCP-1 in clinical plaque instability in humans and complement
23 previous epidemiological, genetic, and experimental studies supporting the translational
24 perspective of targeting MCP-1 signaling in atherosclerosis.

25 **Keywords:** MCP-1; CCL2; atherosclerosis; inflammation; cardiovascular disease; plaque
26 vulnerability; macrophages.

1 **Abbreviations and acronyms**

2	MCP-1	monocyte chemoattractant protein-1
3	IL1 β	interleukin-1 β
4	IL6	interleukin-6
5	CCR2	CC-chemokine receptor 2
6	CCL2	CC-chemokine ligand 2
7	SMC	smooth muscle cell
8	LDL-C	low-density lipoprotein cholesterol
9	eGFR	estimated glomerular filtration rate
10	BMI	body mass index
11	FDR	false discovery rate

1 Introduction

2 Inflammatory mechanisms are critically involved in the pathogenesis of atherosclerosis.^{1, 2}
3 Recent clinical trials on cardiovascular prevention in patients with symptomatic atherosclerotic
4 disease have demonstrated a benefit of anti-inflammatory treatment on top of standard
5 therapy.³⁻⁵ The differences in the efficacy of the tested drugs in these trials risk (canakinumab,
6 methotrexate, colchicine) to lower vascular also highlighted the importance of targeting
7 specific inflammatory pathways.^{3, 5-7} So far, translational efforts mostly focused on the
8 inflammasome- IL1 β - IL6-axis.⁸ Yet, experimental and genetic studies place emphasis on pro-
9 inflammatory mechanisms in atherosclerosis beyond this axis, as has specifically been shown
10 for the chemokine system.^{7, 9}

11 Monocyte chemoattractant protein-1 (MCP-1, also called CC-chemokine ligand 2), is a CC
12 family chemokine that mobilized monocytes from the bone marrow and attracts them to sites
13 of inflammation¹⁰ including the atherogenic arterial wall.¹¹⁻¹⁴ Mice lacking MCP-1 or its receptor
14 CCR2 are protected from atherosclerosis and pharmacological inhibition of the MCP-1/CCR2
15 axis reduces plaque size in experimental atherosclerosis.¹⁵⁻²⁰ Recent genetic and
16 observational data from humans further support associations of circulating MCP-1 levels with
17 the risk of stroke and coronary artery disease.²¹⁻²³ Yet, the translational potential of targeting
18 the MCP-1/CCR2 pathway in human atherosclerosis and specifically whether MCP-1 activity
19 within human plaques is causally involved in atheroprotection remains elusive. To determine
20 the potential clinical utility of targeting MCP-1, it would be critical to clarify associations
21 between MCP-1 levels within plaques and features of plaque vulnerability and instability that
22 underlie the occurrence of clinical events including stroke and myocardial infarction.

23 Here, we analyzed carotid plaque samples from >1000 individuals, who underwent
24 endarterectomy for treatment of asymptomatic or symptomatic carotid stenosis. Our aims were
25 to explore associations of plaque MCP-1 levels with: (i) histopathological features of plaque
26 vulnerability (lipid core, collagen content, macrophage burden, smooth muscle cell (SMC)
27 burden, intraplaque hemorrhage); (ii) plaque inflammation and matrix turnover as assessed by

1 the plaque levels of inflammatory cytokines and metalloproteinase activity; (iii) clinical plaque
2 instability, as defined by a symptomatic plaque causing an acute cerebrovascular event; and
3 (iv) major adverse vascular events occurring after plaque removal (**Figure 1A**).

4 5 **Materials and Methods**

6 The datasets from Athero-Express analyzed for the current study are available upon
7 reasonable request and application to Athero-Express Biobank Study through a Data Transfer
8 Agreement due to consent restrictions and local regulations. Codes used for this study are
9 available here: https://github.com/swvanderlaan/2020_georgakis_vanderlaan_MCP1.

10 *Study population*

11 We used data from the Athero-Express Biobank (<http://www.atheroexpress.nl>), an ongoing
12 prospective study of patients undergoing endarterectomy for manifestations of
13 atherosclerosis.²⁴ Carotid endarterectomy was performed following recommendations by the
14 Asymptomatic Carotid Atherosclerosis Study (ACAS)²⁵ and The North American Symptomatic
15 Carotid Endarterectomy Trial (NASCET).²⁶ Patients were recruited from the St. Antonius
16 Hospital Nieuwegein and University Medical Center Utrecht in Utrecht, Netherlands between
17 2002 and 2019. Individuals who agreed to participate completed questionnaires about medical
18 history and medication use prior to the operation and provided blood samples for biochemical
19 and hematological analyses. Their plaque samples were post-operatively collected and
20 analyzed as described below. Individuals were included in the current study on the basis of
21 having undergone carotid endarterectomy and having available measurements of MCP-1
22 levels in plaque (**Figure 1**). The study protocol conforms to the Declaration of Helsinki and was
23 approved by the ethics committee on research on humans of the University Medical Center
24 Utrecht. All participants provided written informed consent.

1 *Histopathological analysis of atherosclerotic plaque composition*

2 Following carotid endarterectomy, plaque samples were immediately transferred to the
3 laboratory. Plaques were divided in parallel segments of 5-mm thickness perpendicular to the
4 arterial axis and the segment with the greatest plaque burden was subjected to
5 histopathological examination, as previously described.²⁷⁻²⁹ All stained sections were
6 examined microscopically and digitally stored. For the purposes of the current study, we
7 explored five plaque traits that are established hallmarks of plaque vulnerability: lipid content,
8 collagen deposition, macrophages, smooth muscle cells, and intraplaque hemorrhage.^{30, 31}
9 Two independent observers manually scored stainings for these traits using previously defined
10 semi-quantitative methods.^{24, 27-29} In brief, plaque lipid content was quantified visually as a
11 percentage of fat deposition to total plaque area with the use of hematoxylin-eosin and
12 picosirius red stains; a large lipid core was defined as lipid content of >10% of the total plaque
13 area. Collagen deposition (picrosirius red) was manually classified as absent, minor, moderate
14 or heavy staining along the entire luminal border. The burden of macrophages and SMCs was
15 assessed by staining with antibodies against CD68 and α -actin, respectively, and was also
16 manually classified into absent, minor, moderate or heavy staining. In alternative semi-
17 automated computerized analyses, numbers of macrophages and SMCs were quantified on a
18 continuous scale. Specifically, the stainings were scored as percentage of stained area to total
19 plaque area (AnalySiS version 3.2, Soft Imaging GmbH, Munster, Germany).²⁷⁻²⁹ Intraplaque
20 hemorrhage (H&E and fibrin staining) was defined as the composite of plaque bleeding at the
21 luminal side of the plaque as a result of plaque disruption, and was classified as absent or
22 present.

23 To assess the overall vulnerability features of the atherosclerotic plaque, a vulnerability index
24 was created ranging from 0 to 5, as previously described.^{32, 33} Specifically, one point was given
25 to each plaque for the following histopathological features: a lipid core >10%, low collagen load
26 (no/minor), high macrophage burden (moderate/heavy), low SMC burden (no/minor), and
27 presence of intraplaque hemorrhage.

1 *Quantification of plaque levels of MCP-1 and other cytokines*

2 Segments adjacent to those used for histopathological analysis were used for protein isolation.
3 In brief, plaques were manually grinded at -196°C and dissolved in Tris buffer according to an
4 in-house protocol.²⁴ MCP-1 concentrations (pg/mL) were quantified as part of a multiplex assay
5 using the Luminex® platform (Austin, TX, USA) according to the manufacturer's protocol and
6 diagnostic laboratories' standards at the clinical laboratory of the Wilhelmina Children's
7 Hospital (WKZ, Utrecht, the Netherlands).

8 We further quantified the following cytokines and growth factors using established platforms:
9 IL2, IL4, IL5, IL6, IL8, IL9, IL10, IL12, TNF- α , and IFN- γ were measured in multiplex using the
10 human FlowCytomix system from eBioscience (cat.nr.: BMS810FF) in pg/mL. IL13, IL21, MIF,
11 MIP1a, RANTES, MIG, IP10, Eotaxin1, TARC, PARC, MDC, sICAM1, VEGFA, and TGFB
12 were measured in simplex assays using FlowCytomix according to the manufacturer's protocol
13 and diagnostic laboratories' standards at the clinical laboratory of the WKZ. Metalloproteinase
14 (MMP) activity (MMP-2, MMP-8, MMP-9) was assessed with specific Biotrak activity assays
15 (MMP-2 RPN-2631, MMP-8 RPN-2635, and MMP-9 RPN-2634; GE Healthcare LifeSciences,
16 Buckinghamshire, UK). Matrix metalloproteinase levels were corrected for the total protein
17 amount and were expressed as arbitrary units. Given the different platforms used, all protein
18 measurements were inverse-rank transformed to approach normal distributions and to ensure
19 homogeneity in units (per 1-standard deviation [SD]).

20 *Quantification of plasma levels of MCP-1*

21 In a sub-sample of 374 individuals, MCP-1 levels were also quantified in the plasma using the
22 OLINK proximity extension assays (OLINK® Bioscience) in order to explore correlations with
23 MCP-1 plaque concentrations.

24 *Circulating monocyte counts*

25 In a subset of 175 patients, we measured circulating monocyte counts, as previously
26 described.²⁹ In brief, a complete circulating cell profile was determined by a general

1 hematology cell counter (Cell Dyn 1800 Abbott, Minesota, USA). Peripheral blood
2 mononuclear cells (PBMCs) were isolated by Ficoll gradient fractionation and stored at liquid
3 nitrogen. After storage, PBMCs were washed and single cell suspensions were obtained
4 through filtering over a 40 µm cell strainer (542,040 Greiner bio-one). After incubation with
5 fluorescent antibodies, controlling for isotypes, and after exclusion of dead cells, we selected
6 CD11B+CD115+ monocytes using the Kaluza 1.3 gating software. We derived total monocytes
7 counts, as well as identified classical (CD14+CD16-), intermediate (CD14+CD16+) and non-
8 classical (CD14-CD16+) monocytes.

9 *Single-cell RNA-sequencing*

10 In a small sub-sample of 37 individuals we applied single cell-transcriptomics (scRNAseq) to
11 explore the source of MCP-1 levels in the atherosclerotic plaques. Methodological details are
12 described elsewhere.³⁴ In brief, parts of the plaque were minced and enzymatically digested
13 in RPMI 1640 containing 2.5 mg/mL Collagenase IV (ThermoFisher Scientific), 0.25 mg/mL
14 DNase I (Sigma), 2.5 mg/mL Human Albumin Fraction V (MP Biomedicals) and 1 mM
15 Flavopiridol (Selleckchem) at 37°C for 30 minutes. Following filtration through a 70 µm cell
16 strainer and washing, cells were suspended in RPMI 1640 with 1% Fetal Calf Serum, stained
17 with Calcein AM and Hoechst and viable cells were sorted using Beckman Coulter MoFlo
18 Astrios EQ. We used Mosquito® HTS (TTP Labtech) 384 wells plates, filled with 50nL lysis
19 buffer containing CELseq2-primers, spike-ins and dinucleotide triphosphates (dNTPs) and
20 overlaid with mineral oil to prevent evaporation. For downstream analyses we selected cells
21 expressing between 500 and 10.000 genes and genes expressed in at least 3 cells to omit
22 doublets and low-quality cells.³⁴ Data were analyzed using Seurat (Seurat_2.3.4) and log-
23 normalized and scaled with the exclusion of unique molecular identifiers. Subsequently,
24 canonical correlation analysis reduction was performed to identify clusters and to perform t-
25 distributed stochastic neighbor embedding. Cell types were assigned to cell clusters by
26 evaluating gene expression of individual cell clusters using differential gene expression and
27 analysis with SingleR³⁵ against BLUEPRINT reference data.³⁶ Sub-clustering of identified cell

1 clusters was performed using canonical correlation analysis with a resolution of 0.9 or 1.5 for
2 15 dimensions.

3 *Symptomatic vs. asymptomatic plaque*

4 Patients were classified as either having an asymptomatic or symptomatic carotid plaque prior
5 to surgery, based on their answers to a structured questionnaire regarding prior medical history
6 and a detailed review of their medical records. Patients were considered to be symptomatic if
7 they had suffered an acute cerebrovascular event ipsilateral to the plaque within the last 6
8 months. Cerebrovascular events included a supratentorial ischemic stroke, a transient
9 ischemic attack that could be attributed to ischemia in the distribution of the respective artery,
10 an amaurosis fugax, or a central retinal artery occlusion.

11 *Follow-up analysis for major adverse vascular events*

12 Patients were followed up to 3 years after surgery for potential new vascular events. The
13 composite endpoint of any major adverse vascular event included non-fatal stroke (ischemic
14 or hemorrhagic), non-fatal myocardial infarction, ruptured aortic aneurysm, and vascular death,
15 defined as death of presumed vascular origin (fatal stroke, fatal myocardial infarction, sudden
16 death, fatal aortic aneurysm rupture, fatal heart failure, other vascular death). Additional
17 endpoints included stroke (fatal or non-fatal), acute coronary events (fatal or non-fatal
18 myocardial infarction, unstable angina, coronary bypass or percutaneous coronary
19 intervention, and sudden cardiac death), as well as vascular death. Outcomes occurring within
20 the first 30 days after surgery were considered periprocedural events.^{27, 37, 38} All participants
21 underwent clinical follow-up, as detailed elsewhere.²⁷ Clinical endpoints were independently
22 assessed by two clinicians at 1, 2, and 3 years after surgery through patient questionnaires,
23 review of medical records and contact with general practitioners.

24

25

1 *Statistical analysis*

2 Univariable associations between inverse-rank transformed levels of MCP-1 in the plaque with
3 other group variables were explored using Mann-Whitney U-test when comparing two groups
4 and the Kruskal-Wallis test for three or more groups. We explored correlations between plasma
5 and plaque levels of MCP-1 by estimating the Spearman's rho (R). Multivariable models were
6 used to explore associations of plaque levels of MCP-1 (1-SD increment) with (i) plaque protein
7 levels of a panel of 24 cytokines and growth factors (thereafter called cytokines for simplicity)
8 and activity of three metalloproteinases, (ii) histopathological plaque vulnerability phenotypes,
9 (iii) presence of a symptomatic vs. asymptomatic plaque, and (iv) incident major adverse
10 vascular events. Specifically, we performed multivariable logistic regression analyses for
11 symptomatic vs. asymptomatic plaque and for binary histopathological plaque vulnerability
12 traits, as well as linear regression analyses for plaque cytokines and for continuous
13 histopathological plaque vulnerability traits. For the composite vulnerability index (range 0-5),
14 we used ordinal regression analyses. For the prospective analyses for time to new major
15 adverse vascular events, we applied Cox proportional hazard models. Model 1 adjusted for
16 age and sex, whereas Model 2 additionally adjusted for hypertension (self-reported history or
17 antihypertensive medication use), diabetes (defined as self-reported history or glucose-
18 lowering medication use), smoking status (never, former, current smoker), LDL-C levels at time
19 of operation, use of statins or other lipid-lowering drugs, use of antiplatelet agents, estimated
20 glomerular filtration rate (eGFR),³⁹ body mass index (BMI), history of cardiovascular disease
21 (coronary artery disease, stroke, peripheral artery disease), and grade of stenosis (according
22 to NASCET: <70%, 70-90%, 90-99%, complete occlusion). All analyses were corrected for
23 multiple comparisons using the false discovery rate (FDR) approach. Statistical significance
24 threshold was set at a two-sided FDR-adjusted p-value<0.05 across all analyses. Analyses
25 were performed using R (v3.6.3; The R Foundation for Statistical Computing).

26

27

1 **Results**

2 A total of 1,199 patients from the Athero-Express Biobank, who had undergone carotid
3 endarterectomy and had available MCP-1 levels in carotid plaques, were included in the
4 current analysis (mean age 68.6 ± 9.1 years, 36.3% females) (**Figure 1B, Supplemental Table**
5 **I**). MCP-1 levels in carotid plaques were higher among men compared to women, while there
6 was no association with age or vascular risk factors including blood pressure levels, LDL-
7 cholesterol levels, diabetes, smoking, BMI, and kidney function (**Supplemental Figure I**).
8 There was no correlation between plaque and plasma MCP-1 levels in a sub-sample of 374
9 individuals with both measures available (Spearman's $R=0.016$, $p=0.760$, **Supplemental**
10 **Figure II**), thus indicating that plaque MCP-1 may not be a marker of circulating MCP-1 levels.
11 In order to detect the source of MCP-1 levels in the plaque, we applied single-cell
12 transcriptomics in a small sample of 37 individuals and found classical CD14+/CD68+ M1
13 macrophages and to a lesser extent KIT+ mast cells and ACTA2+ smooth muscle cells to be
14 the primary source of expression of *CCL2* in human atherosclerotic plaques (**Supplemental**
15 **Figure III**).

16 *Plaque MCP-1 is associated with histopathological features of plaque vulnerability*

17 First, we explored associations of MCP-1 levels in the plaque with histopathological features
18 of plaque vulnerability (**Figure 2**). MCP-1 levels were significantly associated with all five
19 hallmarks of plaque vulnerability: a large lipid core ($>10\%$), lower collagen content (no/minor),
20 higher macrophage burden (moderate/heavy), lower SMC burden (no/minor), and presence of
21 intraplaque hemorrhage (**Figure 2A-2B; Supplemental Table II**). Similar associations were
22 obtained when considering macrophage and SMC burden as continuous traits (**Supplemental**
23 **Table II**).

24 When combining the five hallmark features of plaque vulnerability traits in a validated
25 aggregate vulnerability index,^{32, 33} plaque MCP-1 levels were gradually higher among
26 individuals with a higher score (ranging from 0-5, $p=6.3 \times 10^{-17}$, **Figure 2C**). In models adjusting

1 for age and sex (model 1), as well as age, sex, and vascular risk factors (model 2), plaque
2 MCP-1 levels were strongly and independently associated with a higher vulnerability index
3 (Model 2: beta 0.42, 95%CI: 0.30-0.53, $p=5.4 \times 10^{-13}$, **Figure 2D**).

4 *Plaque MCP-1 is associated with markers of plaque inflammation and matrix turnover*

5 We then examined whether plaque MCP-1 is associated with markers of plaque inflammation
6 and matrix turnover. Thus, we examined the age- and sex-adjusted associations of plaque
7 MCP-1 levels with multiple cytokines and with metalloproteinase activity in carotid plaques
8 (**Figure 3**). We found significant associations between MCP-1 plaque levels and several
9 cytokines involved in inflammatory cell recruitment. Specifically, we found associations with
10 higher levels of the chemokines IL-8, PARC, TARC, and RANTES, as well with ICAM-1, an
11 adhesion molecule involved in trans-endothelial leukocyte migration^{40, 41} (all FDR-adjusted p-
12 value <0.05 to account for multiple comparisons). Higher MCP-1 levels were further
13 associated with higher levels of VEGF-A, a key driver of plaque neovascularization,⁴² and with
14 higher activity of the matrix metalloproteinases MMP-8, and MMP-9.⁴³ Similar results were
15 obtained when further adjusting for vascular risk factors, although associations with PARC did
16 not remain statistically significant (Model 2, **Supplemental Table III**). Finally, given the role of
17 MCP-1 in monocyte mobilization from the bone marrow beyond monocyte chemotaxis to the
18 plaque, we also explored correlations between MCP-1 levels in the plaque and circulating
19 monocytes and monocyte subtypes in an overlapping sample of 179 individuals. We found no
20 correlation with either total monocyte count or the counts of classical, intermediate, and non-
21 classical monocytes (**Supplemental Figure IV**), thus indicating that MCP-1 plaque levels may
22 not be a marker of monocyte egress from the bone marrow.

23 *Plaque MCP-1 is associated with symptomatic plaques*

24 We further explored associations between plaque MCP-1 levels and clinical plaque instability.
25 MCP-1 levels in the plaque were higher among individuals with a symptomatic plaque (that
26 had caused an acute cerebrovascular event) compared to individuals with asymptomatic

1 plaques ($p=0.0001$, **Figure 4A**). Following adjustments for age and sex, one SD increment in
2 MCP-1 levels in the plaque was associated with higher odds for a symptomatic vs.
3 asymptomatic plaque (OR: 1.31, 95%CI: 1.07-1.60, $p=0.008$). These associations remained
4 significant in a model adjusting for vascular risk factors (OR: 1.36, 95%CI: 1.09-1.69, $p=0.006$,
5 **Figure 4B**).

6 *Plaque MCP-1 is associated with periprocedural stroke during carotid endarterectomy*

7 As a last step, we explored associations between MCP-1 levels in the plaque with vascular
8 events after carotid endarterectomy. Specifically, we looked at events occurring within the first
9 30 days after surgery mostly reflective of periprocedural complications, and at events occurring
10 up to 3 years after surgery (mean follow-up 2.3 years). Plaque MCP-1 levels (1-SD increment)
11 were associated with a higher risk of major adverse vascular events within 30-days after
12 surgery (HR: 2.94, 95%CI: 1.32-6.50, $p=0.007$, **Figure 4C**) independently of age, sex, and
13 vascular risk factors (**Supplemental Table IV**) whereas there was no such association across
14 the entire 3-year period (HR: 1.14, 95%CI: 0.77-1.68, $p=0.519$, **Figure 4D, Supplemental**
15 **Table IV**). MCP-1 plaque levels were further associated with a higher 30-day risk of stroke
16 (HR: 2.32, 95%CI: 1.00-5.36, $p=0.049$, **Figure 4C**).

17

18

1 **Discussion**

2 The present study of 1,199 patients undergoing carotid endarterectomy demonstrated strong
3 associations between plaque MCP-1 levels with multiple features of plaque vulnerability. We
4 found associations between plaque MCP-1 levels and histopathological hallmarks of plaque
5 vulnerability (larger lipid core, low collagen content, high macrophage burden, low SMC
6 burden, intraplaque hemorrhage). We further found higher plaque MCP-1 levels to be
7 associated with higher levels of inflammatory cytokines and higher activity of
8 metalloproteinases within plaques. MCP-1 levels within plaques were higher among individuals
9 with symptomatic plaques, as compared to asymptomatic plaques. Moreover, plaque MCP-1
10 levels were associated with adverse peri-procedural vascular events (time frame: 30 days after
11 plaque removal) independently of traditional vascular risk factors. Collectively, these findings
12 emphasize the importance of plaque MCP-1 levels in plaque vulnerability in human
13 atherosclerosis.

14 While the role of MCP-1 in early stages of atherogenesis through monocyte recruitment in the
15 plaque has been well-established,¹¹⁻¹³ its role in more advanced stages of atherosclerosis
16 remained unknown. The associations reported here between MCP-1 levels in the plaque with
17 all five hallmarks of vulnerable plaques, as well as with a composite vulnerability index, suggest
18 that MCP-1 might be involved in mechanisms related to plaque instability in patients. Our
19 finding of significant associations between plaque MCP-1 levels and symptomatic plaques
20 further support this notion. Importantly, these associations were independent of conventional
21 vascular risk factors, thus suggesting that MCP-1 signaling might contribute to plaque
22 instability on top of established targets for secondary prevention such as LDL-cholesterol,
23 blood pressure, and diabetes.

24 Our current findings on MCP-1 levels in carotid endarterectomy samples complement our
25 recent work showing that circulating MCP-1 levels associate with ischemic stroke, coronary
26 artery disease, and vascular death.²¹⁻²³ Whether MCP-1 levels within atherosclerotic plaques
27 are reflective of MCP-1 activity and active monocyte recruitment remains unknown. Yet, our

1 finding of an association between MCP-1 levels in the plaque with multiple other cell-recruiting
2 chemokines with a proven role in atherosclerosis,⁴⁴⁻⁴⁷ as well as with macrophage burden in
3 the plaque strongly suggest an association with inflammatory cell recruitment.

4 While MCP-1 levels within plaques were not associated with future vascular events up to 3
5 years after plaque removal, we found significant associations with periprocedural events,
6 mainly stroke, occurring within the first 30 days after the procedure. Periprocedural strokes
7 during carotid endarterectomy might have multiple causes, but have been shown to frequently
8 originate from plaque thrombosis and embolization.⁴⁸ Although the mechanisms underlying the
9 observed associations remain unknown, they might relate to the higher risk of periprocedural
10 microembolization previously reported to be more common in patients with features of
11 vulnerable plaques.^{27, 49}

12 To our knowledge, there has been only one small phase II trial targeting MCP-1 signaling in
13 the context of human atherosclerosis. Among 108 patients with vascular risk factors and high
14 CRP levels, treatment with a single intravenous infusion of a humanized monoclonal antibody
15 against the receptor of MCP-1 (CCR2), led to significant reductions in CRP levels.⁵⁰ Of note,
16 the residual risk for clinical events among individuals with carotid plaques currently managed
17 by best medical treatment (statins and anti-platelet agents) remains non-negligible.⁵¹⁻⁵³ Our
18 current data in conjunction with recent genetic,²¹ experimental,¹⁵⁻²⁰ and observational^{22, 23} data
19 on MCP-1 support moving towards clinical trials that target MCP-1 signaling in populations
20 with established atherosclerotic disease.

21 Our study has limitations. First, the cross-sectional nature of most analyses precludes causal
22 inferences. For example, the association between plaque MCP-1 levels and symptomatic
23 plaques could relate to a secondary increase in plaque MCP-1 levels following the acute
24 clinical event.⁵⁴ Similarly, the associations of plaque MCP-1 levels with other pro-inflammatory
25 cytokines, growth factors and metalloproteinase activity can only be interpreted as correlations
26 and not as providing a mechanistic explanation for the associations of MCP-1 with plaque
27 vulnerability. A more detailed mechanistic exploration of the findings would require

1 experimental studies. Second, plaque MCP-1 levels were available only in around half of the
2 Athero-Express population, thus potentially leading to selection bias. Yet, the demographic
3 characteristics of our study sample matched those of the entire Athero-Express sample
4 **(Supplemental Table I)**. Similarly, there might be selection bias due to the
5 underrepresentation of patients with asymptomatic plaques, as they are less likely to undergo
6 carotid endarterectomy. Third, our longitudinal analyses are limited by power due to the low
7 number of new events in the current cohort. Fourth, the pathology and mechanisms of plaque
8 vulnerability might differ between vascular beds and these results from carotid plaques might
9 not be extrapolated to other atherosclerotic lesions. Interestingly, previous studies have also
10 shown involvement of MCP-1 in other vascular diseases, such as venous thromboembolism
11 which however could not be explored in the context of the current study.

12 In conclusion, our study shows that among individuals undergoing carotid endarterectomy,
13 plaque MCP-1 levels are associated with histopathological hallmarks of plaque vulnerability, a
14 pro-inflammatory plaque profile, plaque matrix turnover, clinical plaque instability, and higher
15 risk of periprocedural events. As such, our findings provide evidence for an involvement of
16 MCP-1 in carotid plaque instability and further complement previous evidence from
17 epidemiological, genetic, and experimental studies supporting the translational perspective of
18 targeting the MCP-1/CCR2 axis in atherosclerosis.

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20

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8

9 **References**

- 10 1. Esenwa CC and Elkind MS. Inflammatory risk factors, biomarkers and associated therapy in
11 ischaemic stroke. *Nat Rev Neurol*. 2016;12:594-604.
- 12 2. Libby P, Ridker PM, Hansson GK and Leducq Transatlantic Network on Atherothrombosis.
13 Inflammation in atherosclerosis: from pathophysiology to practice. *J Am Coll Cardiol*. 2009;54:2129-
14 38.
- 15 3. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, Fonseca F,
16 Nicolau J, Koenig W, Anker SD, Kastelein JJP, Cornel JH, Pais P, Pella D, Genest J, Cifkova R,
17 Lorenzatti A, Forster T, Kobalava Z, Vida-Simiti L, Flather M, Shimokawa H, Ogawa H, Dellborg M,
18 Rossi PRF, Troquay RPT, Libby P, Glynn RJ and Cantos Trial Group. Antiinflammatory Therapy with
19 Canakinumab for Atherosclerotic Disease. *N Engl J Med*. 2017;377:1119-1131.
- 20 4. Tardif JC, Kouz S, Waters DD, Bertrand OF, Diaz R, Maggioni AP, Pinto FJ, Ibrahim R,
21 Gamra H, Kiwan GS, Berry C, Lopez-Sendon J, Ostadal P, Koenig W, Angoulvant D, Gregoire JC,
22 Lavoie MA, Dube MP, Rhoads D, Provencher M, Blondeau L, Orfanos A, L'Allier PL, Guertin MC and
23 Roubille F. Efficacy and Safety of Low-Dose Colchicine after Myocardial Infarction. *N Engl J Med*.
24 2019;381:2497-2505.
- 25 5. Ridker PM, Everett BM, Pradhan A, MacFadyen JG, Solomon DH, Zaharris E, Mam V, Hasan
26 A, Rosenberg Y, Iturriaga E, Gupta M, Tsigoulis M, Verma S, Clearfield M, Libby P, Goldhaber SZ,
27 Seagle R, Ofori C, Saklayen M, Butman S, Singh N, Le May M, Bertrand O, Johnston J, Paynter NP,
28 Glynn RJ and Investigators C. Low-Dose Methotrexate for the Prevention of Atherosclerotic Events. *N*
29 *Engl J Med*. 2019;380:752-762.
- 30 6. Ridker PM. Anticytokine Agents: Targeting Interleukin Signaling Pathways for the Treatment of
31 Atherothrombosis. *Circ Res*. 2019;124:437-450.
- 32 7. Aday AW and Ridker PM. Targeting Residual Inflammatory Risk: A Shifting Paradigm for
33 Atherosclerotic Disease. *Front Cardiovasc Med*. 2019;6:16.
- 34 8. Ridker PM. From C-Reactive Protein to Interleukin-6 to Interleukin-1: Moving Upstream To
35 Identify Novel Targets for Atheroprotection. *Circ Res*. 2016;118:145-56.
- 36 9. Koenen RR and Weber C. Therapeutic targeting of chemokine interactions in atherosclerosis.
37 *Nat Rev Drug Discov*. 2010;9:141-53.
- 38 10. Deshmane SL, Kremlev S, Amini S and Sawaya BE. Monocyte chemoattractant protein-1
39 (MCP-1): an overview. *J Interferon Cytokine Res*. 2009;29:313-26.
- 40 11. Lin J, Kakkar V and Lu X. Impact of MCP-1 in atherosclerosis. *Curr Pharm Des*.
41 2014;20:4580-8.
- 42 12. Nelken NA, Coughlin SR, Gordon D and Wilcox JN. Monocyte chemoattractant protein-1 in
43 human atheromatous plaques. *J Clin Invest*. 1991;88:1121-7.
- 44 13. Lutgens E, Faber B, Schapira K, Evelo CT, van Haften R, Heeneman S, Cleutjens KB,
45 Bijnens AP, Beckers L, Porter JG, Mackay CR, Rennert P, Bailly V, Jarpe M, Dolinski B, Kotliansky
46 V, de Fougères T and Daemen MJ. Gene profiling in atherosclerosis reveals a key role for small

1 inducible cytokines: validation using a novel monocyte chemoattractant protein monoclonal antibody.
2 *Circulation*. 2005;111:3443-52.

3 14. Soehnlein O, Drechsler M, Doring Y, Lievens D, Hartwig H, Kemmerich K, Ortega-Gomez A,
4 Mandl M, Vijayan S, Projahn D, Garlachs CD, Koenen RR, Hristov M, Lutgens E, Zernecke A and
5 Weber C. Distinct functions of chemokine receptor axes in the atherogenic mobilization and
6 recruitment of classical monocytes. *EMBO Mol Med*. 2013;5:471-81.

7 15. Liehn EA, Piccinini AM, Koenen RR, Soehnlein O, Adage T, Fatu R, Curaj A, Popescu A,
8 Zernecke A, Kungl AJ and Weber C. A new monocyte chemotactic protein-1/chemokine CC motif
9 ligand-2 competitor limiting neointima formation and myocardial ischemia/reperfusion injury in mice. *J*
10 *Am Coll Cardiol*. 2010;56:1847-57.

11 16. Bot I, Ortiz Zacarias NV, de Witte WE, de Vries H, van Santbrink PJ, van der Velden D,
12 Kroner MJ, van der Berg DJ, Stamos D, de Lange EC, Kuiper J, AP IJ and Heitman LH. A novel CCR2
13 antagonist inhibits atherogenesis in apoE deficient mice by achieving high receptor occupancy. *Sci*
14 *Rep*. 2017;7:52.

15 17. Gu L, Okada Y, Clinton SK, Gerard C, Sukhova GK, Libby P and Rollins BJ. Absence of
16 monocyte chemoattractant protein-1 reduces atherosclerosis in low density lipoprotein receptor-
17 deficient mice. *Mol Cell*. 1998;2:275-81.

18 18. Boring L, Gosling J, Cleary M and Charo IF. Decreased lesion formation in CCR2^{-/-} mice
19 reveals a role for chemokines in the initiation of atherosclerosis. *Nature*. 1998;394:894-7.

20 19. Combadiere C, Potteaux S, Rodero M, Simon T, Pezard A, Esposito B, Merval R, Proudfoot
21 A, Tedgui A and Mallat Z. Combined inhibition of CCL2, CX3CR1, and CCR5 abrogates Ly6C(hi) and
22 Ly6C(lo) monocytosis and almost abolishes atherosclerosis in hypercholesterolemic mice. *Circulation*.
23 2008;117:1649-57.

24 20. Winter C, Silvestre-Roig C, Ortega-Gomez A, Lemnitzer P, Poelman H, Schumski A, Winter J,
25 Drechsler M, de Jong R, Immler R, Sperandio M, Hristov M, Zeller T, Nicolaes GAF, Weber C, Viola
26 JR, Hidalgo A, Scheiermann C and Soehnlein O. Chrono-pharmacological Targeting of the CCL2-
27 CCR2 Axis Ameliorates Atherosclerosis. *Cell Metab*. 2018;28:175-182 e5.

28 21. Georgakis MK, Gill D, Rannikmae K, Traylor M, Anderson CD, Lee JM, Kamatani Y, Hopewell
29 JC, Worrall BB, Bernhagen J, Sudlow CLM, Malik R and Dichgans M. Genetically Determined Levels
30 of Circulating Cytokines and Risk of Stroke. *Circulation*. 2019;139:256-268.

31 22. Georgakis MK, Malik R, Bjorkbacka H, Pana TA, Demissie S, Ayers C, Elhadad MA, Fornage
32 M, Beiser AS, Benjamin EJ, Boekholdt SM, Engstrom G, Herder C, Hoogeveen RC, Koenig W,
33 Melander O, Orho-Melander M, Schiopu A, Soderholm M, Wareham N, Ballantyne CM, Peters A,
34 Seshadri S, Myint PK, Nilsson J, de Lemos JA and Dichgans M. Circulating Monocyte
35 Chemoattractant Protein-1 and Risk of Stroke: Meta-Analysis of Population-Based Studies Involving
36 17 180 Individuals. *Circ Res*. 2019;125:773-782.

37 23. Georgakis MK, de Lemos JA, Ayers C, Wang B, Bjorkbacka H, Pana TA, Thorand B, Sun C,
38 Fani L, Malik R, Dupuis J, Engström G, Orho-Melander M, Melander O, Boekholdt MS, Zierer A,
39 Elhadad MA, Koenig W, Herder C, Hoogeveen RC, Kavousi M, Ballantyne CM, Peters A, Myint PK,
40 Nilsson J, Benjamin EJ and Dichgans M. Circulating monocyte chemoattractant protein-1 levels are
41 associated with 1 cardiovascular mortality: a meta-analysis of population-based studies. *JAMA*
42 *Cardiology (Accepted)*. 2020.

43 24. Verhoeven BA, Velema E, Schoneveld AH, de Vries JP, de Bruin P, Seldenrijk CA, de Kleijn
44 DP, Busser E, van der Graaf Y, Moll F and Pasterkamp G. Athero-express: differential atherosclerotic
45 plaque expression of mRNA and protein in relation to cardiovascular events and patient
46 characteristics. Rationale and design. *Eur J Epidemiol*. 2004;19:1127-33.

47 25. Toole JF. ACAS recommendations for carotid endarterectomy. ACAS Executive Committee.
48 *Lancet*. 1996;347:121.

49 26. Coyne TJ and Wallace MC. Surgical referral for carotid artery stenosis--the influence of
50 NASCET. North American Symptomatic Carotid Endarterectomy Trial. *Can J Neurol Sci*. 1994;21:129-
51 32.

52 27. Hellings WE, Peeters W, Moll FL, Piers SR, van Setten J, Van der Spek PJ, de Vries JP,
53 Seldenrijk KA, De Bruin PC, Vink A, Velema E, de Kleijn DP and Pasterkamp G. Composition of
54 carotid atherosclerotic plaque is associated with cardiovascular outcome: a prognostic study.
55 *Circulation*. 2010;121:1941-50.

56 28. Hellings WE, Pasterkamp G, Vollebregt A, Seldenrijk CA, De Vries JP, Velema E, De Kleijn
57 DP and Moll FL. Intraobserver and interobserver variability and spatial differences in histologic
58 examination of carotid endarterectomy specimens. *J Vasc Surg*. 2007;46:1147-54.

59 29. Meeuwssen JAL, de Vries JJ, van Duijvenvoorde A, van der Velden S, van der Laan SW, van
60 Koeverden ID, van de Weg SM, de Borst GJ, de Winther MPJ, Kuiper J, Pasterkamp G, Hofer IE, de
61 Jager SCA and Queen of Hearts C. Circulating CD14(+)CD16(-) classical monocytes do not associate

1 with a vulnerable plaque phenotype, and do not predict secondary events in severe atherosclerotic
2 patients. *J Mol Cell Cardiol.* 2019;127:260-269.

3 30. Finn AV, Nakano M, Narula J, Kolodgie FD and Virmani R. Concept of vulnerable/unstable
4 plaque. *Arterioscler Thromb Vasc Biol.* 2010;30:1282-92.

5 31. Bentzon JF, Otsuka F, Virmani R and Falk E. Mechanisms of plaque formation and rupture.
6 *Circ Res.* 2014;114:1852-66.

7 32. Timmerman N, de Kleijn DPV, de Borst GJ, den Ruijter HM, Asselbergs FW, Pasterkamp G,
8 Haitjema S and van der Laan SW. Family history and polygenic risk of cardiovascular disease:
9 Independent factors associated with secondary cardiovascular events in patients undergoing carotid
10 endarterectomy. *Atherosclerosis.* 2020.

11 33. Verhoeven B, Hellings WE, Moll FL, de Vries JP, de Kleijn DP, de Bruin P, Busser E,
12 Schoneveld AH and Pasterkamp G. Carotid atherosclerotic plaques in patients with transient ischemic
13 attacks and stroke have unstable characteristics compared with plaques in asymptomatic and
14 amaurosis fugax patients. *J Vasc Surg.* 2005;42:1075-81.

15 34. Depuydt MAC, Prange KHM, Slenders L, Ord T, Elbersen D, Boltjes A, de Jager SCA,
16 Asselbergs FW, de Borst GJ, Aavik E, Lonnberg T, Lutgens E, Glass CK, den Ruijter HM, Kaikkonen
17 MU, Bot I, Slutter B, van der Laan SW, Yla-Herttuala S, Mokry M, Kuiper J, de Winther MPJ and
18 Pasterkamp G. Microanatomy of the Human Atherosclerotic Plaque by Single-Cell Transcriptomics.
19 *Circ Res.* 2020;127:1437-1455.

20 35. Aran D, Looney AP, Liu L, Wu E, Fong V, Hsu A, Chak S, Naikawadi RP, Wolters PJ, Abate
21 AR, Butte AJ and Bhattacharya M. Reference-based analysis of lung single-cell sequencing reveals a
22 transitional profibrotic macrophage. *Nat Immunol.* 2019;20:163-172.

23 36. Martens JH and Stunnenberg HG. BLUEPRINT: mapping human blood cell epigenomes.
24 *Haematologica.* 2013;98:1487-9.

25 37. Randomised trial of endarterectomy for recently symptomatic carotid stenosis: final results of
26 the MRC European Carotid Surgery Trial (ECST). *Lancet.* 1998;351:1379-87.

27 38. Barnett HJ, Taylor DW, Eliasziw M, Fox AJ, Ferguson GG, Haynes RB, Rankin RN, Clagett
28 GP, Hachinski VC, Sackett DL, Thorpe KE, Meldrum HE and Spence JD. Benefit of carotid
29 endarterectomy in patients with symptomatic moderate or severe stenosis. North American
30 Symptomatic Carotid Endarterectomy Trial Collaborators. *N Engl J Med.* 1998;339:1415-25.

31 39. Levey AS, Stevens LA, Schmid CH, Zhang YL, Castro AF, 3rd, Feldman HI, Kusek JW,
32 Eggers P, Van Lente F, Greene T, Coresh J and Ckd EPI. A new equation to estimate glomerular
33 filtration rate. *Ann Intern Med.* 2009;150:604-12.

34 40. Kitagawa K, Matsumoto M, Sasaki T, Hashimoto H, Kuwabara K, Ohtsuki T and Hori M.
35 Involvement of ICAM-1 in the progression of atherosclerosis in APOE-knockout mice. *Atherosclerosis.*
36 2002;160:305-10.

37 41. Nakashima Y, Raines EW, Plump AS, Breslow JL and Ross R. Upregulation of VCAM-1 and
38 ICAM-1 at atherosclerosis-prone sites on the endothelium in the ApoE-deficient mouse. *Arterioscler*
39 *Thromb Vasc Biol.* 1998;18:842-51.

40 42. Camare C, Pucelle M, Negre-Salvayre A and Salvayre R. Angiogenesis in the atherosclerotic
41 plaque. *Redox Biol.* 2017;12:18-34.

42 43. Peeters W, Moll FL, Vink A, van der Spek PJ, de Kleijn DP, de Vries JP, Verheijen JH, Newby
43 AC and Pasterkamp G. Collagenase matrix metalloproteinase-8 expressed in atherosclerotic carotid
44 plaques is associated with systemic cardiovascular outcome. *Eur Heart J.* 2011;32:2314-25.

45 44. Simonini A, Moscucci M, Muller DW, Bates ER, Pagani FD, Burdick MD and Strieter RM. IL-8
46 is an angiogenic factor in human coronary atherectomy tissue. *Circulation.* 2000;101:1519-26.

47 45. Veillard NR, Kwak B, Pelli G, Mulhaupt F, James RW, Proudfoot AE and Mach F. Antagonism
48 of RANTES receptors reduces atherosclerotic plaque formation in mice. *Circ Res.* 2004;94:253-61.

49 46. Weber C, Meiler S, Doring Y, Koch M, Drechsler M, Megens RT, Rowinska Z, Bidzhekov K,
50 Fecher C, Ribechini E, van Zandvoort MA, Binder CJ, Jelinek I, Hristov M, Boon L, Jung S, Korn T,
51 Lutz MB, Forster I, Zenke M, Hieronymus T, Junt T and Zernecke A. CCL17-expressing dendritic cells
52 drive atherosclerosis by restraining regulatory T cell homeostasis in mice. *J Clin Invest.*
53 2011;121:2898-910.

54 47. Orekhov AN, Nikiforov NG, Elizova NV, Korobov GA, Aladinskaya AV, Sobenin IA and
55 Bobryshev YV. Tumor Necrosis Factor-alpha and C-C Motif Chemokine Ligand 18 Associate with
56 Atherosclerotic Lipid Accumulation In situ and In vitro. *Curr Pharm Des.* 2018;24:2883-2889.

57 48. Jacobowitz GR, Rockman CB, Lamparello PJ, Adelman MA, Schanzer A, Woo D, Landis R,
58 Gagne PJ, Riles TS and Imparato AM. Causes of perioperative stroke after carotid endarterectomy:
59 special considerations in symptomatic patients. *Ann Vasc Surg.* 2001;15:19-24.

- 1 49. Verhoeven BA, de Vries JP, Pasterkamp G, Ackerstaff RG, Schoneveld AH, Velema E, de
2 Kleijn DP and Moll FL. Carotid atherosclerotic plaque characteristics are associated with
3 microembolization during carotid endarterectomy and procedural outcome. *Stroke*. 2005;36:1735-40.
- 4 50. Gilbert J, Lekstrom-Himes J, Donaldson D, Lee Y, Hu M, Xu J, Wyant T, Davidson M and
5 Group MLNS. Effect of CC chemokine receptor 2 CCR2 blockade on serum C-reactive protein in
6 individuals at atherosclerotic risk and with a single nucleotide polymorphism of the monocyte
7 chemoattractant protein-1 promoter region. *Am J Cardiol*. 2011;107:906-11.
- 8 51. Spence JD, Coates V, Li H, Tamayo A, Munoz C, Hackam DG, DiCicco M, DesRoches J,
9 Bogiatzi C, Klein J, Madrenas J and Hegele RA. Effects of intensive medical therapy on microemboli
10 and cardiovascular risk in asymptomatic carotid stenosis. *Arch Neurol*. 2010;67:180-6.
- 11 52. Inzitari D, Eliasziw M, Gates P, Sharpe BL, Chan RK, Meldrum HE and Barnett HJ. The
12 causes and risk of stroke in patients with asymptomatic internal-carotid-artery stenosis. North
13 American Symptomatic Carotid Endarterectomy Trial Collaborators. *N Engl J Med*. 2000;342:1693-
14 700.
- 15 53. Nadareishvili ZG, Rothwell PM, Beletsky V, Pagniello A and Norris JW. Long-term risk of
16 stroke and other vascular events in patients with asymptomatic carotid artery stenosis. *Arch Neurol*.
17 2002;59:1162-6.
- 18 54. Roth S, Singh V, Tiedt S, Schindler L, Huber G, Geerlof A, Antoine DJ, Anfray A, Orset C,
19 Gauberti M, Fournier A, Holdt LM, Harris HE, Engelhardt B, Bianchi ME, Vivien D, Haffner C,
20 Bernhagen J, Dichgans M and Liesz A. Brain-released alarmins and stress response synergize in
21 accelerating atherosclerosis progression after stroke. *Sci Transl Med*. 2018;10.

22

23

1 **Highlights**

2

3 • Robust evidence from epidemiological, genetic, and preclinical studies supports a role of
4 MCP-1 in atherosclerosis. Still, the role of MCP-1 in human plaque vulnerability is
5 unknown

6 • In 1,199 individuals undergoing endarterectomy due to carotid stenosis, MCP-1 plaque
7 levels were associated with histopathological and molecular markers of plaque
8 vulnerability

9 • MCP-1 levels were also associated with clinical plaque instability, as indicated by
10 symptomatic plaques and a higher risk of periprocedural major adverse vascular events
11 and strokes

12 • Our findings provide support for a role of MCP-1 in plaque vulnerability in humans

13

1 **Figure legends**

2

3 **Figure 1. Study design.** (A) Graphical illustration of the study design. (B) Flowchart detailing
4 the number of individuals included in the respective analyses. Also shown are the number of
5 individuals excluded from the current analyses and the reasons for exclusion.

6

7 **Figure 2. Plaque MCP-1 levels associate with histopathological hallmarks of plaque**
8 **vulnerability.** (A) Graphical depiction of histopathological features of a vulnerable plaque. (B)
9 Associations of plaque MCP-1 levels (1-SD increment) with the individual vulnerability features
10 (binary traits) as derived from logistic regression analyses (adjusted for age, sex, hypertension,
11 diabetes, current smoking, LDL-C levels at time of operation, use of lipid-lowering agents, use
12 of antiplatelet agents, estimated glomerular filtration rate, body mass index, history of
13 cardiovascular disease, and grade of stenosis). Shown are Odds Ratios (OR) and error bars
14 correspond to their 95% confidence intervals (CI). (C) Plaque MCP-1 levels (inverse-rank
15 transformed) in study participants across plaque vulnerability index scores (p-value derived
16 from Kruskal-Wallis test). Shown are the median values (central line), the upper and lower
17 quartiles (box limits) and the 1.5x interquartile range (whiskers). (D) Multivariable associations
18 between MCP-1 levels in plaque (1 SD-increment) with the composite plaque vulnerability
19 index score, as derived from ordinal regression analyses (Model 1 adjusted for age and sex,
20 Model 2 additionally adjusted for the abovementioned vascular risk factors). Shown are beta
21 coefficients and the error bars correspond to their 95%CI. MCP-1 plaque levels are inverse-
22 rank transformed in all analyses.

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25

1 **Figure 3. Plaque MCP-1 levels associate with markers of plaque inflammation and matrix**
2 **turnover.** Shown are the beta coefficients and 95% confidence intervals (95%CI) derived from
3 linear regression models for 1-SD increment in plaque MCP-1 levels (inverse-rank
4 transformed) adjusted for age and sex. Stars indicate statistically significant results (false
5 discovery rate-adjusted p-value <0.05).

6

7 **Figure 4. Plaque MCP-1 levels associate with symptomatic versus asymptomatic plaque**
8 **and periprocedural events after plaque removal. (A)** Plaque MCP-1 levels (inverse-rank
9 transformed) in patients with symptomatic versus asymptomatic plaques (p-value derived from
10 Mann-Whitney U-test). Shown are the median values (central line), the upper and lower
11 quartiles (box limits) and the 1.5x interquartile range (whiskers) **(B)** Multivariable associations
12 between MCP-1 plaque levels (1 SD-increment) with symptomatic plaque as derived from
13 logistic regression analyses (vascular risk factors in the second model include hypertension,
14 diabetes, current smoking, LDL-C levels at time of operation, use of lipid-lowering agents, use
15 of antiplatelet agents, estimated glomerular filtration rate, body mass index, history of
16 cardiovascular disease, and grade of stenosis). Shown are Odds Ratios (OR) and error bars
17 correspond to their 95% confidence intervals (CI). **(C, D)** Associations of plaque MCP-1 levels
18 (1-SD increment) with new adverse vascular events within **(C)** 30 days (periprocedural events)
19 and **(D)** 3 years after surgery, as derived from Cox regression analyses (adjusted for age, sex,
20 and the abovementioned vascular risk factors). Shown are Hazard Ratios (OR) derived from
21 Cox regression analyses and error bars correspond to their 95%CI. MCP-1 plaque levels are
22 inverse-rank transformed in all analyses.

23