Serum antioxidants as predictors of adult respiratory distress syndrome in patients with sepsis

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Adult respiratory distress syndrome (ARDS) can develop as a complication of various disorders, including sepsis, but it has not been possible to identify which of the patients at risk will develop this serious disorder. We have investigated the ability of six markers, measured sequentially in blood, to predict development of ARDS in 26 patients with sepsis.

At the initial diagnosis of sepsis (6–24 h before the development of ARDS), serum manganese superoxide dismutase concentration and catalase activity were higher in the 6 patients who subsequently developed ARDS than in 20 patients who did not develop ARDS. These changes in antioxidant enzymes predicted the development of ARDS in septic patients with the same sensitivity, specificity, and efficiency as simultaneous assessments of serum lactate dehydrogenase activity and factor VIII concentration. By contrast, serum glutathione peroxidase activity and α1-Pi-elastase complexes did not differ at the initial diagnosis of sepsis between patients who did and did not subsequently develop ARDS, and were not as effective in predicting the development of ARDS.

Measurement of manganese superoxide dismutase and catalase, in addition to the other markers, should facilitate identification of patients at highest risk of ARDS and allow prospective treatment.


Introduction

Adult respiratory distress syndrome (ARDS) is an acute inflammatory process characterised by neutrophil accumulation and oedema in the lungs and progressive hypoxaemia. ARDS occurs unpredictably as a complication in patients with sepsis and several other predisposing disorders. Since many common and diverse risk factors can lead to the development of ARDS, but do so only rarely, pretreatment of every patient at risk of ARDS is not practical. Understanding of ARDS is improving and various interventions that can limit inflammation have become available, so it has become important to identify accessible and repeatable markers in at-risk patients that can predict the development of ARDS. Experimental treatments could then be prospectively and effectively assessed in smaller, better-defined groups of patients.

We assessed the abilities of three antioxidant enzymes, manganese superoxide dismutase, catalase, and glutathione peroxidase, and three other potential markers, factor VIII, lactate dehydrogenase, and α1-Pi-elastase complexes, to predict the development of ARDS in patients with sepsis. Oxidant-antioxidant balance changes in ARDS and in other disease states in which oxygen radicals have been implicated. Serum catalase activity is high in patients with established ARDS, but it is not known whether this abnormality arises before the development of ARDS, in which case it could be predictive, or afterwards.

Patients and methods

Written consent was obtained from each patient or a close relative. The subjects were studied with a protocol approved by our hospital’s human subjects review committee. Eligible patients (n = 26) were identified within 8 h of the diagnosis of sepsis, which was defined as a serious bacterial infection and either a rectal or core temperature above 39°C or a peripheral leucocyte count of more than 1·2 × 10⁹/L or more than 20% immature neutrophils. For a diagnosis of sepsis at least one of the following was also required: a positive blood culture of a commonly accepted pathogen, a strongly suspected or proven source of systemic infection, large amounts of pus in a closed space, unexplained systemic arterial hypotension (systolic pressure below 80 mm Hg), systemic vascular resistance less than 800 dyne s cm⁻², or unexplained metabolic acidosis.

Criteria for ARDS were acute respiratory failure requiring mechanical ventilation, bilateral pulmonary infiltrates, pulmonary

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capillary wedge pressure below 18 mm Hg, static pulmonary compliance below 50 mL/cm H2O, and ratio of arterial to alveolar partial pressure of oxygen of less than 0.25. Blood samples were taken at the diagnosis of sepsis (0 h) and also 6, 12, 24, and 48 h later. ARDS developed in 6 patients 6–24 h after the diagnosis of sepsis. Samples were also obtained when ARDS was diagnosed and 6–24 h later either through an indwelling arterial or venous catheter or by direct venipuncture. All patients were prospectively and sequentially studied until death or discharge. Assays were done by staff unaware of the diagnoses. 15 healthy subjects were studied as controls.

Manganese superoxide dismutase, factor VIII antigen, and α,β-Pi-elastase complexes were measured by enzyme-linked immunosorbent assays (ELISAs). The ELISA for manganese superoxide dismutase had a sensitivity of 2 ng/mL and a coefficient of variation of less than 3%. Catalase was assayed by polarographic measurement of oxygen production. 10 μL serum was added to 10 mmol/L hydrogen peroxide in 400 μL Hanks’ balanced salt solution in an airtight, temperature-controlled chamber, which was stirred continuously. Oxygen production was measured with an oxygen electrode, calibrated daily. 1 U catalase activity was defined as the amount of enzyme that consumed 1 μmole of hydrogen peroxide per min at 25°C. The assay had a sensitivity of about 0.5 U/mL and a coefficient of variation below 5%. Glutathione peroxidase was measured spectrophotometrically by the oxidation of NADPH at 340 nm in glutathione reductase, glutathione, and t-butyl hydroperoxide. Lactate dehydrogenase and albumin were assayed spectrophotometrically and uric acid by high-performance liquid chromatography.

The patients with and without ARDS were similar (p > 0.05) in terms of age, sex, packed-cell volume, haemoglobin concentration, blood leucocyte and neutrophil counts, serum alanine aminotransferase activity, serum concentrations of bilirubin, albumin, and uric acid, and APACHE II score, as reported previously. 3 of the 6 patients in whom ARDS developed and 6 of the 20 without ARDS died.

Patients with sepsis had higher serum manganese superoxide dismutase concentrations than did control subjects (figure). However, at the time of diagnosis of sepsis (6–24 h before ARDS was diagnosed in 6 patients) the patients who progressed to ARDS had higher serum concentrations of this enzyme than did patients who did not develop ARDS (p < 0.05). The concentrations remained high for the next 48 h in the ARDS group but returned to control values in the non-ARDS patients. Similarly, at the time of diagnosis of sepsis, serum catalase activity was higher in patients with sepsis than in control subjects (p < 0.05) and in turn patients who later developed ARDS had higher
ARDS but decreased in the other patients. By contrast, we found no significant difference between control subjects and sepsis patients with or without subsequent ARDS in serum glutathione peroxidase activity.

Serum lactate dehydrogenase activity was also significantly higher in patients who subsequently developed ARDS than in patients who did not progress to ARDS (p < 0.05); the latter had activities similar to those of the controls. Lactate dehydrogenase activity increased during the study period in sepsis patients who developed ARDS but not in the other groups.

For factor VIII, serum concentrations were higher in patients with sepsis than in the healthy controls but there were no significant differences between septic patients with and without ARDS. Plasma concentrations of α1-Pi-elastase complexes also were higher in septic patients than controls at diagnosis of sepsis, but the difference between patients who did and did not develop ARDS was significant (p < 0.05) only when ARDS had been diagnosed.

We found no correlations at any time among the six markers. When we calculated indices of the markers' ability to predict ARDS development, based on cutoff points chosen by inspection of the data to provide the best separation of ARDS and non-ARDS subjects, the positive and negative predictive values and the sensitivity and specificity of serum manganese superoxide dismutase concentrations (above or equal to a cutoff of 450 ng/mL or above), lactate dehydrogenase activity (30 U/mL or above), and factor VIII concentrations (above or equal to a cutoff of 445 % of control values or more) were similar in predicting the development of ARDS in patients with sepsis (table). Manganese superoxide dismutase, catalase, and lactate dehydrogenase values exceeded the chosen cutoff values 9 h, 12 h, and 12 h, respectively, before ARDS was diagnosed.

### Discussion

We have found that 9–12 h before the development of ARDS, amounts of two serum antioxidant enzymes were increased in sepsis patients who later developed ARDS compared with patients who did not develop ARDS; both factors predicted the development of ARDS in sepsis patients with sensitivity, specificity, and efficiency equal to those of lactate dehydrogenase and factor VIII measurements.

Assessment of serum manganese superoxide dismutase and catalase could be useful for defining the pathogenesis of ARDS or for identifying patients with similar characteristics. Each measurement is accessible, repeatable, and easy to do. Use of these assessments could facilitate study of prophylactic treatment by reducing the number of at-risk patients needed. Our findings may also have functional importance. These antioxidant enzymes can diminish oxidant insults mediated by superoxide anion, hydrogen peroxide, and their products such as hydroxyl radicals. Accelerated intravascular generation of oxygen radicals from stimulated neutrophils, circulating xanthine oxidase, and other sources has been implicated in the pathogenesis of sepsis and ARDS.

Since the time courses for the various markers differed and no two markers were intercorrelated, our findings suggest that each factor represents a distinct process and that the factors reflect various processes that occur in septic patients with ARDS rather than in ARDS itself. This study focused on sepsis-induced ARDS, and different mechanisms may be present in ARDS that develops after trauma or in other conditions. It is not clear where the markers are produced. Lung tissue injury is a possible source for higher than normal production of lactate dehydrogenase, factor VIII, and the two antioxidant enzymes. Endothelial cells are rich in these substances and, if perturbed, could readily increase the amounts in blood. However, intravascular neutrophil activation must bring about increases in α1-Pi-elastase complexes, since elastase is present only in neutrophils. Changes in α1-Pi-elastase complexes occurred later than the other changes at diagnosis of ARDS, and concentrations had fallen to normal by 48 h after the diagnosis of sepsis, perhaps indicating a decline in neutrophil activity. Red blood cell haemolysis is a plausible source for increased amounts of serum catalase and lactate dehydrogenase, but not manganese superoxide dismutase or factor VIII. Serum catalase activity is increased in the serum of rats subjected to skin burns and in patients with acquired immunodeficiency syndrome, but again the source is unclear. High concentrations of interleukin-1, tumour necrosis factor, and endotoxin have been found in ARDS patients and can cause increases in antioxidants such as manganese superoxide dismutase and catalase.

Perhaps increases in antioxidants reflect activation of intrinsic antioxidant tolerance mechanisms mediated by cytokines.

We thank Ms Jacqueline Smith for graphics, the patient care staff at Denver General Hospital, Mr Michael Owens and Ms Mary Gillespie for technical assistance, and Dr Lisa Beth Ferstenberg and Mr Kyle Davis for expert help with statistical analysis.

J. A. L. was in receipt of a clinician-scientist award from the American Heart Association and P. E. P. a clinical investigator award from the National Institutes of Health (HL-01849). The study was supported by grants from the National Institutes of Health (HL 40784, 45582), Colorado Heart Association, Council for Tobacco Research, Ronald McDonald Children’s Charities, Johnson and Johnson, Williams Family, Swan, Hill, Kleberg, Donner, and American Express Foundations.

### Table: Sensitivity and Specificity of Blood Markers as Predictors of ARDS in Patients with Sepsis

<table>
<thead>
<tr>
<th>Marker</th>
<th>Cutoff</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>Positive Predictive Value (%)</th>
<th>Negative Predictive Value (%)</th>
<th>Efficiency (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>MnSOD</td>
<td>≥450 ng/mL</td>
<td>67 (42–94)</td>
<td>88 (75–98)</td>
<td>67 (42–94)</td>
<td>88 (75–98)</td>
<td>83 (70–94)</td>
</tr>
<tr>
<td>Catalase</td>
<td>≥30 U/mL</td>
<td>83 (61–99)</td>
<td>65 (49–82)</td>
<td>42 (25–68)</td>
<td>93 (81–100)</td>
<td>69 (55–84)</td>
</tr>
<tr>
<td>GPX</td>
<td>≥72 U/mL</td>
<td>72 (57–93)</td>
<td>47 (31–69)</td>
<td>23 (12–53)</td>
<td>73 (33–92)</td>
<td>48 (34–66)</td>
</tr>
<tr>
<td>LDH</td>
<td>≥250 U/L</td>
<td>67 (42–94)</td>
<td>78 (62–92)</td>
<td>50 (29–81)</td>
<td>88 (74–98)</td>
<td>75 (61–89)</td>
</tr>
<tr>
<td>Factor VIII</td>
<td>≥445% control</td>
<td>83 (61–99)</td>
<td>67 (42–94)</td>
<td>45 (27–73)</td>
<td>92 (30–100)</td>
<td>71 (57–85)</td>
</tr>
<tr>
<td>α1-Pi-elastase</td>
<td>≥940 ng/mL</td>
<td>67 (37–98)</td>
<td>64 (30–80)</td>
<td>18 (8–47)</td>
<td>94 (84–100)</td>
<td>64 (51–79)</td>
</tr>
</tbody>
</table>

*MnSOD = manganese superoxide dismutase; GPX = glutathione peroxidase; LDH = lactate dehydrogenase

**True results as a percentage of all results.
REFERENCES


